

Target cell availability and the successful suppression of HIV by hydroxyurea and didanosine

Rob J. De Boer, Charles A.B. Boucher* and Alan S. Perelson†

AIDS 1998, 12:1567–1570

Keywords: Hydroxyurea, immunosuppression, target cell availability, resistance

Introduction

Surprisingly, immunosuppressive treatment can enhance the efficacy of conventional HIV-1 antiretroviral treatment, and can be beneficial for HIV-1-infected patients. This argues for a role of target cell availability in limiting the HIV-1 infection, and is in agreement with mathematical models suggesting that immunosuppression may limit the outgrowth of drug-resistant escape mutants. Immunosuppressive drugs like hydroxyurea (HU) may therefore be powerful and affordable supplements to HIV-1 antiretroviral therapy.

Clinical trials

Recent clinical trials in HIV-1-infected patients have investigated the long-term synergistic effect of HU on conventional antiretroviral therapy with the nucleoside analogue didanosine (ddI). Vila *et al.* [1] treated ddI-naive individuals with CD4 cell counts above $200 \times 10^6/l$ with HU and ddI, and reported that after 1 year, 10 out of 20 patients had no detectable virus in plasma or lymphoid tissue. Two of these patients stopped therapy and had extracellular virus remain undetectable in both lymph nodes and plasma for 1 year [2]. Similarly, Lori *et al.* [3] reported that after

72 weeks of ddI–HU treatment, three out of six patients had no detectable plasma virus, and that there was no rebound of the plasma viral load in any patient on uninterrupted treatment. There is also an intriguing anecdotal report of a patient on indinavir, ddI and HU, who after having had HIV driven down to an undetectable level stopped taking these drugs, and remained undetectable for 9 months [4].

Short-term studies report similar encouraging results of the ddI–HU combination in patients naive for ddI. During the first month of treatment the viral load decreases sharply by 1–2 \log_{10} copies/ml, and several patients had undetectable virus levels after 3 months [5], 4 months [6], or 6 months [7] of treatment. In another study, 1000 mg daily HU treatment added to chronic ddI therapy decreased viral load by approximately 1 \log_{10} copies/ml and decreased CD4 cell count by 25% [8]. The combination of HU with ddI is more potent than combinations with other nucleoside analogues [9], probably because HU preferentially depletes intracellular dATP concentrations [10,11]. However, monotherapy with HU failed to have a beneficial effect on plasma HIV RNA load (but may decrease CD4 cell count) [12,13]. Two studies have compared ddI monotherapy with the ddI–HU combination. They either failed to find a difference [14], or reported a significantly stronger decrease in plasma viraemia with the

From Theoretical Biology, Utrecht University, the *Department of Virology, Eijkman-Winkler Institute, Utrecht University Hospital, Utrecht, The Netherlands, and the †Theoretical Division, Los Alamos National Laboratory, Los Alamos, New Mexico, USA.

Note: Portions of this work were performed under the auspices of the US Department of Energy and the Santa Fe Institute.

Sponsorship: This work was supported by NIH grant RR06555, NATO grant GRC960019, Dutch AIDS foundation (PccO grant 1317), and the Jeanne P. and Joseph M. Sullivan Foundation.

Requests for reprints to: Dr Rob J. De Boer, Theoretical Biology, Utrecht University, Padualaan 8, 3584 CH Utrecht, The Netherlands.

Date of receipt: 13 February 1998; revised: 13 May 1998; accepted: 19 May 1998.

ddI-HU combination [7]. Patients on ddI-HU treatment routinely develop mutations known to confer resistance to ddI [7,11]. However, since ddI-resistant mutants grow poorly in the presence of HU [11], these patients have a lower plasma virus concentration than those on ddI monotherapy [7,11].

Efficacy of ddI-HU

Why is long-term treatment with ddI-HU effective? HU blocks the cellular enzyme ribonucleotide reductase, which thus decreases the intracellular concentrations of nucleosides required for DNA synthesis [9,15]. By decreasing the intracellular dATP pool, HU may favour the incorporation of ddI [11]. HU is also a cell cycle-specific toxin [16] that inhibits the S-phase of the cell cycle [17]. It is a routinely prescribed cytostatic drug used in the treatment of leukaemia [18] and to kill dividing T cells [19,20]. Although the HU dosage employed in ddI-HU trials should be low enough to avoid haematological side-effects [18], most (but not all [5]) of the HU trials in HIV patients report suppression [8,12,13] or poor recovery [1,3,6] of peripheral blood CD4 cell counts. This negative impact on peripheral blood CD4 cell counts is an important difference between the ddI-HU combination and other forms of antiretroviral therapy. By killing dividing CD4+ T cells and by depleting intracellular dATP concentrations HU reduces the availability of suitable target cells for HIV. Using mathematical models we have shown that such a reduction of target cell availability during antiretroviral treatment can strongly reduce the growth rates of drug-resistant escape mutants [21]. This effect, we believe, explains the encouraging long-term effects of the ddI-HU combination [1-3], even in the apparent presence of ddI-resistance mutations [7,11].

There is ample evidence that the availability of activated CD4+ T cells limits HIV-1 levels during clinical latency. Stimulating the immune system with interleukin-2 in the absence of potent antiretroviral therapy may increase the viral load [22]. Immunization of HIV-1-infected patients with either influenza vaccine [23,24], hepatitis B vaccine [25], pneumococcal vaccine [26], or tetanus toxoid [27], which should all activate T cells, tends to increase the viral load. A similar increase in HIV levels is seen during infection with pathogenic organisms [28,29]. The early rebound of wild-type virus observed during zidovudine treatment [30] finds a straightforward explanation in the increased target cell availability when the CD4 cell counts recover [30-33]. Finally, the high CD4+ T-cell production in children [34] may explain the high viral loads that HIV-infected children tend to have [35,36]. If HIV is target-cell-limited during clinical latency [37,38], one should be able to exploit this by immunosuppressive therapies

decreasing target cell levels. Suppression with cyclosporine [39-42], prednisolone [43,44] and HU [1,3,5,6,8] indeed have beneficial effects.

Mathematical models

Analysing mathematical models in which the HIV infection is target-cell-limited one finds that for any strain of HIV there exists a minimum target cell number below which the strain cannot be maintained [21]. This threshold number is set by various viral characteristics, such as its infection rate, burst size, and lifespan [21]. This finding is identical to classical results in epidemiology stating that any infectious disease has a critical host density below which the infection cannot maintain itself. Because HIV-1 infection is at quasi-steady state during clinical latency [45,46], the steady-state target cell level should be close to this epidemiological threshold. Target cell numbers higher than this would allow a target-cell-limited virus to expand, which is consistent with the data reviewed above, while target cell numbers below this threshold will lead to viral decay [21,47].

Analysing antiretroviral therapy in the same mathematical model, we have predicted precisely the long-term effects that are observed now with the ddI-HU combination: the major beneficial effect of supplementing antiretroviral therapy with target cell suppression should be a reduced expansion of drug-resistant mutants [21]. Pre-existing drug-resistant variants, having a lower fitness than the pretreatment wild-type virus [48,49], require higher target cell levels than the wild-type virus in order to expand. Likewise, novel mutants arising under drug pressure are unlikely to attain a fitness higher than that of wild-type virus before the onset of treatment. Thus, the recovery of the CD4+ target cell population seems the 'Achilles heel' of conventional antiretroviral therapy: the increased target cell availability allows drug-resistant mutants to escape [21,30,33,47]. The encouraging long-term effect of ddI-HU treatment on the viral load, allowing in most cases only for a limited CD4 cell recovery, is therefore in good agreement with our conjecture that HU decreases target cell availability and consequently reduces, or even prevents, the outgrowth of drug resistant escape mutants [21].

Conclusion

Importantly, our results suggest that similar long-term beneficial effects are to be expected from the combination of HU, or other immunosuppressive agents, with other antiretroviral drugs. Obviously this should be

tested carefully because lowering CD4+ T cells may put patients at risk of even more opportunistic infections, and because immunosuppression would be harmful if the HIV infection is largely controlled by immune responses rather than by target cell availability. The current encouraging results with the ddi-HU combination nevertheless supports our conjecture that some degree of target cell depletion could be very beneficial by preventing drug resistance [21]. If this turns out to be true, it would open up inexpensive and well-tolerated new therapeutic strategies for patients not responding to current therapies, and for countries unable to afford them.

References

- Vila J, Biron F, Nugier F, Vallet T, Peyramond D: **1-year follow-up of the use of hydroxycarbamide and didanosine in HIV infection.** *Lancet* 1996, **348**:203–204.
- Vila J, Nugier F, Bargues G, et al.: **Absence of viral rebound after treatment of HIV-infected patients with didanosine and hydroxycarbamide.** *Lancet* 1997, **350**:635–636.
- Lori F, Jessen H, Foli A, Lisziewicz J, Matteo PS: **Long-term suppression of HIV-1 by hydroxyurea and didanosine.** *JAMA* 1997, **277**:1437–1438.
- Cohen J: **HIV suppressed long after treatment [news].** *Science* 1997, **277**:1927.
- Biron F, Lucht F, Peyramond D, et al.: **Pilot clinical trial of the combination of hydroxyurea and didanosine in HIV-1 infected individuals.** *Antiviral Res* 1996, **29**:111–113.
- Clotet B, Ruiz L, Cabrera C, et al.: **Short term anti-HIV activity, at three month interval, of the combination didanosine and hydroxyurea.** *Antiviral Ther* 1996, **1**:189–193.
- De Antoni A, Foli A, Lisziewicz J, Lori F: **Mutations in the *pol* gene of the human immunodeficiency virus type 1 in infected patients receiving didanosine and hydroxyurea combination therapy.** *J Infect Dis* 1997, **176**:899–903.
- Montaner JS, Zala C, Conway B, et al.: **A pilot study of hydroxyurea among patients with advanced human immunodeficiency virus (HIV) disease receiving chronic didanosine therapy: Canadian HIV Trials Network Protocol 080.** *J Infect Dis* 1997, **175**:801–806.
- Lori F, Malykh A, Cara A, et al.: **Hydroxyurea as an inhibitor of human immunodeficiency virus-type 1 replication.** *Science* 1994, **266**:801–805.
- Gao WY, Johns DG, Chokekuchai S, Mitsuya H: **Disparate actions of hydroxyurea in potentiation of purine and pyrimidine 2',3'-dideoxynucleoside activities against replication of human immunodeficiency virus.** *Proc Natl Acad Sci USA* 1995, **92**:8333–8337.
- Lori F, Malykh AG, Foli A, et al.: **Combination of a drug targeting the cell with a drug targeting the virus controls human immunodeficiency virus type 1 resistance.** *AIDS Res Hum Retroviruses* 1997, **13**:1403–1409.
- Simonelli C, Nasti G, Vaccher E, et al.: **Hydroxyurea treatment in HIV-infected patients.** *J Acquir Immune Defic Syndr Hum Retrovirol* 1996, **13**:462–464.
- Giacca M, Zanussi S, Comar M, et al.: **Treatment of human immunodeficiency virus infection with hydroxyurea: virologic and clinical evaluation.** *J Infect Dis* 1996, **174**:204–209.
- Simonelli C, Comar M, Zanussi S, De Paoli P, Tirelli U, Giacca M: **No therapeutic advantage from didanosine (ddi) and hydroxyurea versus ddi alone in patients with HIV infection.** *AIDS* 1997, **11**:1299–1300.
- Meyerhans A, Vartanian JP, Hultgren C, et al.: **Restriction and enhancement of human immunodeficiency virus type 1 replication by modulation of intracellular deoxynucleoside triphosphate pools.** *J Virol* 1994, **68**:535–540.
- Cousens LP, Orange JS, Biron CA: **Endogenous IL-2 contributes to T cell expansion and IFN-gamma production during lymphocytic choriomeningitis virus infection.** *J Immunol* 1995, **155**:5690–5699.
- Buchkovich KJ, Greider CW: **Telomerase regulation during entry into the cell cycle in normal human T cells.** *Mol Biol Cell* 1996, **7**:1443–1454.
- Donehower RC: **An overview of the clinical experience with hydroxyurea.** *Semin Oncol* 1992, **19**:11–19.
- Rocha B, Freitas AA, Coutinho AA: **Population dynamics of T lymphocytes. Renewal rate and expansion in the peripheral lymphoid organs.** *J Immunol* 1983, **131**:2158–2164.
- Ropke C: **Renewal rates of murine T-lymphocyte subsets.** *Cell Immunol* 1990, **128**:185–197.
- De Boer RJ, Boucher CA: **Anti-CD4 therapy for AIDS suggested by mathematical models.** *Proc R Soc Lond B Biol Sci* 1996, **263**:899–905.
- Kovacs JA, Baseler M, Dewar RJ, et al.: **Increases in CD4 T lymphocytes with intermittent courses of interleukin-2 in patients with human immunodeficiency virus infection. A preliminary study.** *N Engl J Med* 1995, **332**:567–575.
- Staprans SI, Hamilton BL, Follansbee SE, et al.: **Activation of virus replication after vaccination of HIV-1-infected individuals.** *J Exp Med* 1995, **182**:1727–1737.
- O'Brien WA, Grovit-Ferbas K, Namazi A, et al.: **Human immunodeficiency virus-type 1 replication can be increased in peripheral blood of seropositive patients after influenza vaccination.** *Blood* 1995, **86**:1082–1089.
- Cheeseman SH, Davaro RE, Ellison RT III: **Hepatitis B vaccination and plasma HIV-1 RNA [letter].** *N Engl J Med* 1996, **334**:1272.
- Brichacek B, Swindells S, Janoff EN, Pirruccello S, Stevenson M: **Increased plasma human immunodeficiency virus type 1 burden following antigenic challenge with pneumococcal vaccine.** *J Infect Dis* 1996, **174**:1191–1199.
- Stanley SK, Ostrowski MA, Justement JS, et al.: **Effect of immunization with a common recall antigen on viral expression in patients infected with human immunodeficiency virus type 1.** *N Engl J Med* 1996, **334**:1222–1230.
- Goletti D, Weissman D, Jackson RW, et al.: **Effect of *Mycobacterium tuberculosis* on HIV replication. Role of immune activation.** *J Immunol* 1996, **157**:1271–1278.
- Orenstein JM, Fox C, Wahl SM: **Macrophages as a source of HIV during opportunistic infections.** *Science* 1997, **276**:1857–1861.
- De Jong MD, Veenstra J, Stilianakis NI, et al.: **Host-parasite dynamics and outgrowth of virus containing a single K70R amino acid change in reverse transcriptase are responsible for the loss of human immunodeficiency virus type 1 RNA load suppression by zidovudine.** *Proc Natl Acad Sci USA* 1996, **93**:5501–5506.
- McLean AR, Emery VC, Webster A, Griffiths PD: **Population dynamics of HIV within an individual after treatment with zidovudine.** *AIDS* 1991, **5**:485–489.
- McLean AR, Nowak MA: **Competition between zidovudine-sensitive and zidovudine-resistant strains of HIV.** *AIDS* 1992, **6**:71–79.
- Stilianakis NI, Boucher CA, De Jong MD, Van Leeuwen R, Schuurman R, De Boer RJ: **Clinical data sets of human immunodeficiency virus type 1 reverse transcriptase-resistant mutants explained by a mathematical model.** *J Virol* 1997, **71**:161–168.
- Mackall CL, Fleisher TA, Brown MR, et al.: **Age, thymopoiesis, and CD4+ T-lymphocyte regeneration after intensive chemotherapy.** *N Engl J Med* 1995, **332**:143–149.
- Steketee RW, Abrams EJ, Thea DM, et al.: **Early detection of perinatal human immunodeficiency virus (HIV) type 1 infection using HIV RNA amplification and detection. New York City Perinatal HIV Transmission Collaborative Study.** *J Infect Dis* 1997, **175**:707–711.
- Shearer WT, Quinn TC, LaRussa P, et al.: **Viral load and disease progression in infants infected with human immunodeficiency virus type 1.** *N Engl J Med* 1997, **336**:1337–1342.
- Coffin JM: **HIV population dynamics *in vivo*: implications for genetic variation, pathogenesis, and therapy.** *Science* 1995, **267**:483–489.
- Feinberg MB, McLean AR: **AIDS: decline and fall of immune surveillance?** *Curr Biol* 1997, **7**:R136–R140.
- Andrieu JM, Even P, Venet A, et al.: **Effects of cyclosporin on T-cell subsets in human immunodeficiency virus disease.** *Clin Immunol Immunopathol* 1988, **47**:181–198.

40. Schwarz A, Offermann G, Keller F, *et al.*: **The effect of cyclosporine on the progression of human immunodeficiency virus type 1 infection transmitted by transplantation: data on four cases and review of the literature.** *Transplantation* 1993, **55**:95–103.
41. Weber J, Galpin S: **HIV results in the frame. Cyclosporin A [letter].** *Nature* 1995, **375**:198.
42. Martin LN, Murphey-Corb M, Mack P, *et al.*: **Cyclosporin A modulation of early virologic and immunologic events during primary simian immunodeficiency virus infection in rhesus monkeys.** *J Infect Dis* 1997, **176**:374–383.
43. Andrieu JM, Lu W, Levy R: **Sustained increases in CD4 cell counts in asymptomatic human immunodeficiency virus type 1-seropositive patients treated with prednisolone for 1 year.** *J Infect Dis* 1995, **171**:523–530.
44. Corey L: **Reducing T cell activation as a therapy for human immunodeficiency virus infection.** *J Infect Dis* 1995, **171**:521–522.
45. Ho DD, Neumann AU, Perelson AS, Chen W, Leonard JM, Markowitz M: **Rapid turnover of plasma virions and CD4 lymphocytes in HIV-1 infection.** *Nature* 1995, **373**:123–126.
46. Wei X, Ghosh SK, Taylor ME, *et al.*: **Viral dynamics in human immunodeficiency virus type 1 infection.** *Nature* 1995, **373**:117–122.
47. Wein LM, D'Amato RM, Perelson AS: **Mathematical analysis of antiretroviral therapy aimed at HIV-1 eradication or maintenance of low viral loads.** *J Theor Biol* 1998, **192**:81–98.
48. Goudsmit J, De Ronde A, Ho DD, Perelson AS: **Human immunodeficiency virus fitness *in vivo*: calculations based on a single zidovudine resistance mutation at codon 215 of reverse transcriptase.** *J Virol* 1996, **70**:5662–5664.
49. Goudsmit J, De Ronde A, De Rooij E, De Boer RJ: **Broad spectrum of *in vivo* fitness of human immunodeficiency virus type 1 subpopulations differing at reverse transcriptase codons 41 and 215.** *J Virol* 1997, **71**:4479–4484.