

SYNTHESIS OF 5-*O*- β -D-GALACTOFURANOSYL-D-GALACTOFURANOSE

WOLFGANG A. R. VAN HEESWIJK, HENNY G. J. VISSER, AND JOHANNES F. G. Vliegenthart

Laboratory of Organic Chemistry, University of Utrecht (The Netherlands)

(Received December 30th, 1976; accepted for publication March 23rd, 1977)

ABSTRACT

Conversion of benzyl α - β -D-galactofuranoside into the 5,6-*O*-[α -(dimethylamino)benzylidene] derivative, followed by acetylation of HO-2 and HO-3, and selective ring opening of the acetal, gave benzyl 2,3-di-*O*-acetyl-6-*O*-benzoyl- α - β -D-galactofuranoside (**4**). The title disaccharide was synthesised from **4** by reaction with 3,4,6-tri-*O*-acetyl- α -D-galactofuranose 1,2-(methyl orthoacetate) followed by removal of protecting groups.

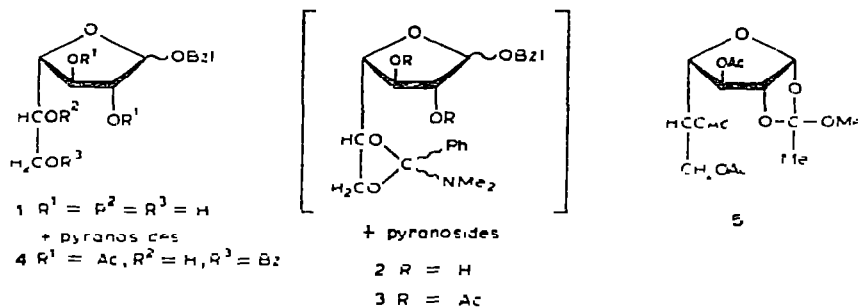
INTRODUCTION

The occurrence of (1 \rightarrow 5)-linked D-galactofuranoid units in oligosaccharides has been demonstrated by the isolation of 5-*O*- β -D-glucopyranosyl-D-galactofuranose¹ and 5-*O*- β -D-galactofuranosyl-D-galactofuranose² from partial acid hydrolysates of the type-specific substance of *Pneumococcus* 33B and galactocarlose, respectively. We now report a synthesis of the latter disaccharide.

RESULTS AND DISCUSSION

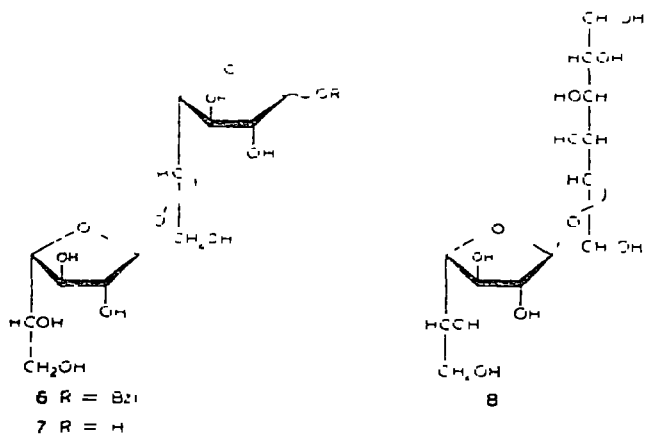
Syrupy benzyl α - β -D-galactofuranoside (**1**) was prepared in 62% yield by treatment of D-galactose diethyl dithioacetal with benzyl alcohol in the presence of mercuric oxide and chloride³. Furanosides obtained by this method are frequently syrups⁴, although crystalline benzyl β -D-galactofuranoside has been isolated⁵ (12%) from **1**. The ¹H-n.m.r. spectrum of **1** in methyl sulfoxide-*d*₆ contained signals for anomeric protons at δ ~4.9 for the furanose forms, made up of an overlapping singlet ($J_{1,2}$ ~0 Hz, H-1 β) and a doublet ($J_{1,2}$ 3-4 Hz, H-1 α), as observed for 6-aminoethyl D-galactofuranoside⁶, and accounting for at least 90% of the mixture. A small doublet at δ 4.73 ($J_{1,2}$ 3-4 Hz) was tentatively assigned to H-1 of the α -pyranoside, but its quantification was hampered by the proximity of resonances for the benzyl methylene group.

Following the procedure of Hanessian and Moralioglu⁷, **1** was treated with an excess of *N,N*-dimethylbenzamide dimethyl acetal in chloroform to give a mixture (1:1) of two diastereomeric forms of benzyl 5,6-*O*-[α -(dimethylamino)benzylidene]- α - β -D-galactofuranoside (**2**) in virtually quantitative yield. Reaction of **2** with acetic



anhydride in pyridine afforded the 2,3-diacetate **3** which with aqueous acetic acid, gave benzyl 2,3-di-*O*-acetyl-6-*O*-benzoyl- α -D-galactofuranoside (**4**). An overall yield of 69% was obtained for the sequence $1 \rightarrow 2 \rightarrow 3 \rightarrow 4$ when the intermediates were not isolated. Attempts to purify **2** and **3** resulted in considerable losses due to the lability of the 5,6-acetal groups. Compound **4** was purified by column chromatography, and its 1H -n.m.r. spectrum was consistent with the assigned structure.

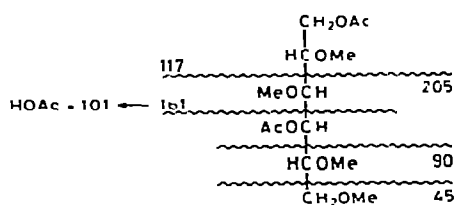
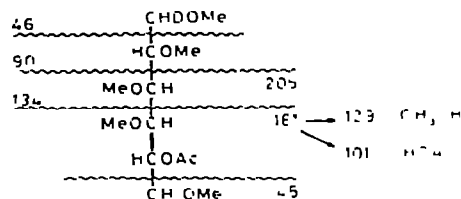
Condensation of 3,5,6-tri-*O*-acetyl- α -D-galactofuranose 1,2-(methyl orthoacetate) (**5**) with **4** in nitromethane in the presence of mercuric bromide, with deacetylation of the product, gave a mixture from which benzyl 5-*O*- β -D-galactofuranosyl-D-galactofuranoside (**6**) was isolated. Although **6** gave one spot in t.l.c., it contained impurities that were revealed after debenzylation which gave mainly 5-*O*- β -D-galactofuranosyl-D-galactofuranose (**7**). The contaminants were reducing disaccharides containing galactopyranose residues (1H -n.m.r. spectroscopy). Purification of **7** was readily achieved by ion-exchange column chromatography.⁸



The presence of the (1 \rightarrow 5)-linkage in **7** was confirmed by 1H -n.m.r. spectroscopy, although assignment of the furanoid anomeric protons was only possible on the basis of a 360-MHz spectrum.

The physical constants of **7** and those of crystalline 5-*O*- β -D-galactofuranosyl-D-galactitol (**8**), obtained upon borohydride reduction of **7** corresponded closely to those reported by Gorin and Spencer². The retention times of trimethylsilylated synthetic and authentic **8** in g.l.c. were identical (co-chromatography) (see Experimental).

Reduction of **7** with sodium borodeuteride followed by methylation⁹ and then^{9,10} hydrolysis, reduction, and acetylation gave 1,4-di-*O*-acetyl-2,3,5,6-tetra-*O*-methyl-D-galactitol (**9**) and 5-*O*-acetyl-1,2,3,4,6-penta-*O*-methyl-D-galactitol-1-*d* (**10**), which were identified by g.l.c.-m.s. and proved the presence of a furanosyl (1 \rightarrow 5)-linkage in **7**. The ¹H-n.m.r. and optical rotation data for **7** and **8** established the intersugar glycosidic linkage to be β .

**9****10**

EXPERIMENTAL

General — 5-*O*- β -D-Galactofuranosyl-D-galactitol was a gift from Dr. P. A. J. Gorin. Pyridine (1 litre) was purified by distillation from chlorosulphonic acid (10 ml) and kept over molecular sieve 5A. Nitromethane was purified as described earlier.¹¹ Solutions were concentrated at 40 (bath)°/14 mmHg. Specific rotations were determined at ambient temperature with a Perkin-Elmer model 141 polarimeter. ¹H-N.m.r. spectra were recorded with Varian EM-390 (90 MHz) and Bruker HX-360 (360 MHz) spectrometers, with tetramethylsilane or sodium 2,2-dimethyl-2-silapentane-5-sulphonate (DSS) as the internal standard. Directions for the preparation of partially methylated per-*O*-acetylated hexitols and for the interpretation of their g.l.c.-m.s. data are given in the literature.^{9,10} Mass spectra (75 eV) were recorded on a Jeol JGC-100/JMS-07 combination [column of 3% of SE-30 on Chromosorb W-AW DMCS (80–100 mesh) at 158°, ion-source temperature 250°, accelerating voltage 3 kV, ionizing current 300 μ A]. G.l.c. of trimethylsilyl derivatives¹² of sugars was carried out at 227° with a Pye 104 instrument equipped with flame-ionisation detector and glass columns (1.60 m \times 4 mm) packed with 3.8% of SE-30 or 3% of OV-17 on Chromosorb W-AW DMCS (80–100 mesh) and a nitrogen flow-rate of 30 ml/min. Retention times (T_R) are given relative to that of Me₃Si-sucrose.

T.l.c. was performed on silica gel (Schleicher & Schull TLC Ready Plastic Foil FR-1500). All reactions were monitored by t.l.c., and mobilities are expressed as R_F .

where X denotes a reference compound or a starting material. The spots were detected by charring with sulphuric acid. Benzyl derivatives were also detected under u.v. light (254 nm). Column chromatography was performed on silica gel (Merck Kieselgel 60, 230–400 mesh) with *A*, benzene–propan-2-ol–water (30:14:1), *B*, chloroform–methanol (25:1), *C*, chloroform–methanol (100:1), *D*, acetic acid–ethyl acetate–butan-1-ol–water (6:3:8:1), *E*, water, *F*, ethyl acetate–acetic acid–water (9:2:2), and *G*, propan-2-ol–ethyl acetate–water (5:5:2).

Benzyl D-galactofuranoside (1) — The crude syrupy product⁴ was purified by column chromatography (solvent *A*). The first fraction was concentrated several times from its ethanolic solution to remove residual solvent *A* and yielded **1** as a syrup (62%), $[\alpha]_D^{25} -23$ (c 1, methanol). The material gave one spot in t.l.c. (solvents *A* and *G*), and g.l.c. of the Me₃Si derivative gave one peak with a shoulder on the negative slope. ¹H-N.m.r. data (methyl sulphoxide-*d*₆) δ 7.2–7.5 (m, 5 H, aromatic protons), 4.9 (m, 0.9 H, $J_{1,2} \sim 0$ and 3–4 Hz, H-1 β and H-1 α of furanosides), 4.73 [d, $J_{1,2}$ 3.4 Hz, H-1 α (?)] of pyranoside], and 3.3–4.8 (other protons). [Found (for freeze-dried **1**) C, 57.70, H, 6.67. C₁₃H₁₈O₆ calc. C, 57.77, H, 6.71%].

Benzyl 2,3-di-O-acetyl-6-O-benzoyl-D-galactofuranoside (4) — A mixture of **1** (3.2 g, 11.8 mmol), *N,N*-dimethylbenzamide dimethyl acetal⁶ (5 g, 25.6 mmol), and chloroform (20 ml) was stirred in the dark at room temperature. After 16 h, t.l.c. (solvent *B*) revealed the absence of **1** and the formation of two closely migrating components at R_f 4 in approximately equal amounts. The solution was cooled, and diluted with chloroform (80 ml) and ice-water (80 ml), and the organic layer was separated, dried (Na₂SO₄), filtered, and concentrated to ~10 ml at 30° (bath). To the solution (containing **2***), pyridine (20 ml) was immediately added followed by acetic anhydride (3.5 ml, 37 mmol). After 24 h, t.l.c. (solvent *B*) revealed the formation of two closely migrating components (**3**, R_f 2.5) in approximately equal amounts and the absence of **3**. The solution was diluted with chloroform (80 ml), extracted with ice-water (80 ml), dried at 0° (Na₂SO₄), and concentrated, and pyridine was removed from the residue by evaporation of toluene three times therefrom. The product (**3***, 7 g) was immediately dissolved in acetic acid–water (19 l, 40 ml), and the solution was kept at room temperature for 2 h and then poured into 8% aqueous sodium hydrogen carbonate (1 litre). The mixture was extracted with chloroform (3 \times 200 ml) and the extract was dried (Na₂SO₄) and concentrated. The residual syrup consisted (t.l.c., solvent *B*) of a major component (**4**) and four minor components having R_f 1.29, 1.23, 0.86, and 0.80. Compound **4** was purified by column chromatography (gradient elution, solvents *C* \rightarrow *B*) and obtained as a colourless oil (3.75 g, 8.18 mmol, 69% based on **1**), $[\alpha]_D^{25} -36$ (c 4, chloroform). ¹H-N.m.r. data (chloroform-*d*) δ 7.1–7.6 and 7.9–8.2 (2 m, 5 H, OBz), 7.3 (m, 5 H, Ph), 2.0 (s, 6 H, 2 AcO), 3.23 (d, 1 H, $J_{HO,5}$ 8 Hz, HO-5, disappeared upon deuterium-exchange), and 3.8–5.8 (m, 9 H, sugar ring protons and benzyl-CH₂) (Found C, 63.31, H, 5.96. C₂₄H₂₆O₉ calc. C, 62.88, H, 5.72%).

⁴Solutions containing **2** or **3** gave a positive reaction⁶ for α -(dimethylamino)benzylidene acetal.

Benzyl 5-O- β -D-galactofuranosyl-D-galactofuranoside (6) — 3,5,6-Tri-*O*-acetyl- α -D-galactofuranose 1,2-(methyl orthoacetate)⁷ (**5**, 2.28 g, 6.29 mmol) and **4** (2.3 g, 5.02 mmol) were dissolved in nitromethane (25 ml). The solvent was distilled at atmospheric pressure with simultaneous addition of nitromethane to keep the volume constant. Mercuric bromide (84 mg) was added after a few ml of solvent had distilled, and distillation was then continued for 4 h. The mixture was cooled, pyridine (1 ml) was added, the precipitate was removed, and the filtrate was concentrated. The resulting syrup (6 g) was deacetylated in 0.02M methanolic sodium methoxide (100 ml). After completion of the reaction (t.l.c., solvents *A* and *B*), the solution was neutralized with Amberlite IR-120 (H⁺) resin, and concentrated to dryness. The residue was eluted from a column⁸ (1.5 m \times 2 cm) of Dowex 50 X4 (K⁺) resin with solvent *E* at 0.4 ml/min. D-Galactose, methyl β -D-galactofuranoside, **6**, and **1** were separately eluted. The fraction containing **6** was freeze-dried to yield a syrup (262 mg, 12%) [α]_D -113 (c 8, methanol) which was homogeneous in t.l.c. (solvent *A*, *R*_{GAL} 2). ¹H-N.m.r. data (deuterium oxide) δ 7.5 (m, 5 H, Ph), 5.0–5.3 (m, 2 H, anomeric protons), 3.6–4.3 (m, sugar ring protons), and 4.7–4.9 (m, benzyl-CH₂) partially masked by an HOD signal [Found: C, 50.10, H, 6.78. Calc. for C₁₆H₂₈O₁₁ · 1.3 H₂O: C, 50.06, H, 6.77%].

5-O- β -D-Galactofuranosyl-D-galactofuranose (7) — A vigorously stirred solution of **6** (250 mg, 0.58 mmol) in 90% aqueous ethanol (100 ml) was hydrogenated at atmospheric pressure over 10% palladium-on-charcoal (200 mg) for 2.5 h. Removal of the catalyst and concentration left a residue which consisted mainly of **7** (t.l.c., solvent *D*; *R*_{GAL} 0.85) with minor components having *R*_{GAL} 0.60, 0.40 and 0.30. These impurities proved to be at least partly, disaccharides containing galactopyranoses, as shown by ¹H-n.m.r. spectroscopy (deuterium oxide) δ 4.63 (d, 0.2 H, *J*_{1,2} 7.5 Hz, H-1 β pyranose), 5.0–5.3 (m, 1.8 H, overlapping signals for H-1 of α - and β -furanose and α -pyranose) and 3.4–4.3 (m, 12 H, sugar ring protons). Ion-exchange chromatography⁸ (solvent *E*), as described for **6**, yielded **7** (120 mg, 61%) as a syrup. [α]_D -64 (c 2, water), lit.² [α]_D -65 (c 3.8, water). The freeze-dried sugar had [α]_D -74 (c 2, water) and *R*_{GAL} 0.85 (descending p.c., solvent *F*), lit.² *R*_{GAL} 0.9. The 360-MHz ¹H-n.m.r. spectrum of **7** in deuterium oxide is consistent with a 2:3 α : β -mixture δ 5.197 (*J*_{1,2} 3–4 Hz, H-1 α), 5.231 (*J*_{1,2} \sim 0 Hz, H-1' β), and 5.244 (*J*_{1,2} \sim 0 Hz, H-1 β), 5.206 (*J*_{1,2} \sim 0 Hz, H-1' β) [Found (for freeze-dried **7**): C, 41.62, H, 6.37. C₁₂H₂₂O₁₁ calc.: C, 42.11; H, 6.48%].

5-O- β -D-Galactofuranosyl-D-galactitol (8) — Reduction² of **7** (40 mg, 0.12 mmol) with sodium borohydride (40 mg) gave a crude alditol (38 mg, 94%) which was homogeneous in t.l.c. (solvent *G*, *R*_{GAL} 1) and by ¹H-n.m.r. spectroscopy (deuterium oxide) δ 5.27 (s, 1 H, *J*_{1,2} \sim 0 Hz, H-1') and 3.4–4.3 (m, 14 H, other protons). When recrystallized from methanol-ethanol, **8** had m.p. 148–150, [α]_D -63 (c 2, water), lit.² m.p. 149–151, [α]_D -65 (water).

On g.l.c. of the Me₃Si derivatives on SE-30 and OV-17, the following *T*_S values were observed: 7.090, 1.13 and 0.90, 1.15, 8.137 and 1.38.

Methanolysis of **8** (1 mg) followed by g.l.c., as devised by Clamp *et al.*^{13,14},

showed the presence of galactose and galactitol in a 1:1 ratio. Alditol acetate analysis^{9, 10} of 8-d₁ was in accordance with the assigned structure.

ACKNOWLEDGMENTS

We thank Mr. G. J. Gerwig for conducting the methanolysis experiments, Mr. C. Versluis (Laboratory of Analytical Chemistry, State University, Utrecht) for recording the mass spectra, and Mr. A. V. E. George and Dr. L. Dorland for recording the 90- and 360-MHz ¹H-nmr spectra. This investigation was supported by the Netherlands Foundation for Chemical Research (S.O.N.) with financial aid from the Netherlands Organisation for the Advancement of Pure Research (Z.W.O.).

REFERENCES

1. M. J. WATSON, *Biochem. J.* **137** (1974) 603-606.
2. P. A. J. GORIN AND I. F. T. SPENCER, *Can. J. Chem.* **37** (1959) 499-501.
3. J. W. GREEN AND E. PACSU, *J. Am. Chem. Soc.* **59** (1937) 1205-1210.
4. J. W. GREEN AND E. PACSU, *J. Am. Chem. Soc.*, **60** (1938) 2056-2057.
5. J. E. SCHNEIDER AND Y. C. LEE, *Carbohydr. Res.* **43** (1975) 79-91.
6. S. HANESSIAN AND E. MORALIOGLU, *Can. J. Chem.*, **50** (1972) 233-245.
7. J. JACQUINET AND P. SINAY, *Carbohydr. Res.* **34** (1974) 343-349.
8. R. M. SAUNDERS, *Carbohydr. Res.*, **7** (1968) 76-79.
9. J. LONNGREN AND S. SVENSSON, *Adv. Carbohydr. Chem. Biochem.* **29** (1974) 41-106.
10. H. BJØRNDAL, B. LINDBERG, A. PILOTTI AND S. SVENSSON, *Carbohydr. Res.* **15** (1970) 339-349.
11. N. K. KOCHETKOV AND A. F. BOCHKOV, *Methods Carbohydr. Chem.* **6** (1972) 480-485.
12. J. P. KAMERLING, J. F. G. VliegENTHART, J. VINK AND J. J. DE RIDDER, *Tetrahedron* **27** (1971) 4275-4288.
13. J. R. CLAMP, T. BHATTI AND R. E. CHAMBERS, *Methods Biochem. Anal.*, **19** (1971) 229-344.
14. J. P. KAMERLING, G. J. GERWIG, J. F. G. VliegENTHART, AND J. R. CLAMP, *Biochem. J.* **151** (1975) 491-495.