

## SYNTHESIS OF 5-*O*- $\alpha$ - AND - $\beta$ -D-GLUCOPYRANOSYL-D-GLUCOFURANOSE AND 5-*O*- $\alpha$ -D-GLUCOPYRANOSYL-D-FRUCTOPYRANOSE (LEUCROSE)

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### ABSTRACT

Reaction of 1,2-*O*-cyclopentylidene- $\alpha$ -D-glucofuranurono-6,3-lactone (**2**) with 2,3,4,6-tetra-*O*-acetyl- $\alpha$ -D-glucofuranosyl bromide (**1**) gave 1,2-*O*-cyclopentylidene-5-*O*-(2,3,4,6-tetra-*O*-acetyl- $\alpha$ -D-glucofuranosyl)- $\alpha$ -D-glucofuranurono-6,3-lactone (**3**, 45%) and 1,2-*O*-cyclopentylidene-5-*O*-(2,3,4,6-tetra-*O*-acetyl- $\beta$ -D-glucofuranosyl)- $\alpha$ -D-glucofuranurono-6,3-lactone (**4**, 38%). Reduction of **3** and **4** with lithium aluminium hydride, followed by removal of the cyclopentylidene group, afforded 5-*O*- $\alpha$ - (**9**) and - $\beta$ -D-glucofuranosyl-D-glucofuranose (**12**), respectively. Base-catalysed isomerization of **9** yielded crystalline 5-*O*- $\alpha$ -D-glucofuranosyl-D-fructopyranose (leucrose, 53%).

### INTRODUCTION

The disaccharide leucrose (**13**) has been isolated from dextran-producing cultures of *L. mesenteroides*<sup>1</sup> and *S. bovis*<sup>2,3</sup>. The latter bacterium also produces 5-*O*- $\alpha$ -isomaltosyl-D-fructopyranose<sup>3</sup>. In addition, 5-*O*- $\alpha$ -D-glucofuranosyl-D-glucofuranose (**9**), which is closely related structurally to **13**, was erroneously believed to be present in manioc flour<sup>4–6</sup>.

We have prepared<sup>6</sup> 5-*O*-(2-acetamido-2-deoxy- $\alpha$ -D-glucofuranosyl)- $\beta$ -D-glucofuranose (**11**) *via* condensation of dimeric 3,4,6-tri-*O*-acetyl-2-deoxy-2-nitroso- $\alpha$ -D-glucofuranosyl chloride with 1,2-*O*-isopropylidene- $\alpha$ -D-glucofuranurono-6,3-lactone. However, attempted preparation of **9** by a similar route failed. The cyclopentylidene acetal is a better protecting group for the glucofuranose residue than is the isopropylidene acetal<sup>7</sup>, and we now report the synthesis of **9**, **12**, and **13** using 1,2-*O*-cyclopentylidene- $\alpha$ -D-glucofuranurono-6,3-lactone<sup>7</sup> (**2**) as the aglycon precursor.

### RESULTS AND DISCUSSION

Condensation of **1** with **2**<sup>7</sup> in acetonitrile in the presence of mercuric cyanide and bromide was poorly stereospecific. However, the yield of products was high in comparison with that (10–11%) from 3,6-di-*O*-acetyl-1,2-*O*-isopropylidene- $\alpha$ -D-glucofuranose<sup>8,9</sup>. The coupling products **3** (45%) and **4** (38%) were readily separated

by fractional crystallization from ethanol. Reduction of **3** and **4** with lithium aluminium hydride in 1,2-dimethoxyethane afforded the cyclopentylidene-protected disaccharides, which were isolated and characterized as the hexa-acetates **5** and **6**, respectively. Deacetylation of **5** and **6** in methanolic sodium methoxide gave 1,2-*O*-cyclopentylidene-5-*O*- $\alpha$ - (**7**) and - $\beta$ -*D*-glucopyranosyl- $\alpha$ -*D*-glucofuranose (**8**), respectively. The  $^1\text{H-n.m.r.}$  spectra obtained at various stages of the preparation of **7** and **8** were in agreement with the structures assigned. The anomeric configuration of the acetates **3-6** was established on the basis of the chemical shift of H-5'; H-5 in 2,3,4,6-tetra-*O*-acetyl- $\beta$ -*D*-glucopyranosides resonates at higher field than in the  $\alpha$  anomers. The interglycosidic, anomeric proton resonances were discernible after deacetylation.

The removal of the cyclopentylidene group from **7** and **8** was effected with 0.52M sulphuric acid<sup>7</sup>, yielding the free sugars **9** and **12** (~96%), respectively. The change in optical rotation accompanying the hydrolyses was consistent with the presence of a (1  $\rightarrow$  5)-linkage<sup>7</sup> in both **9** and **12**. Compound **12** was characterized as its known toluene-*p*-sulphonylhydrazone<sup>9,10</sup>. The g.l.c. data (Table I) for the

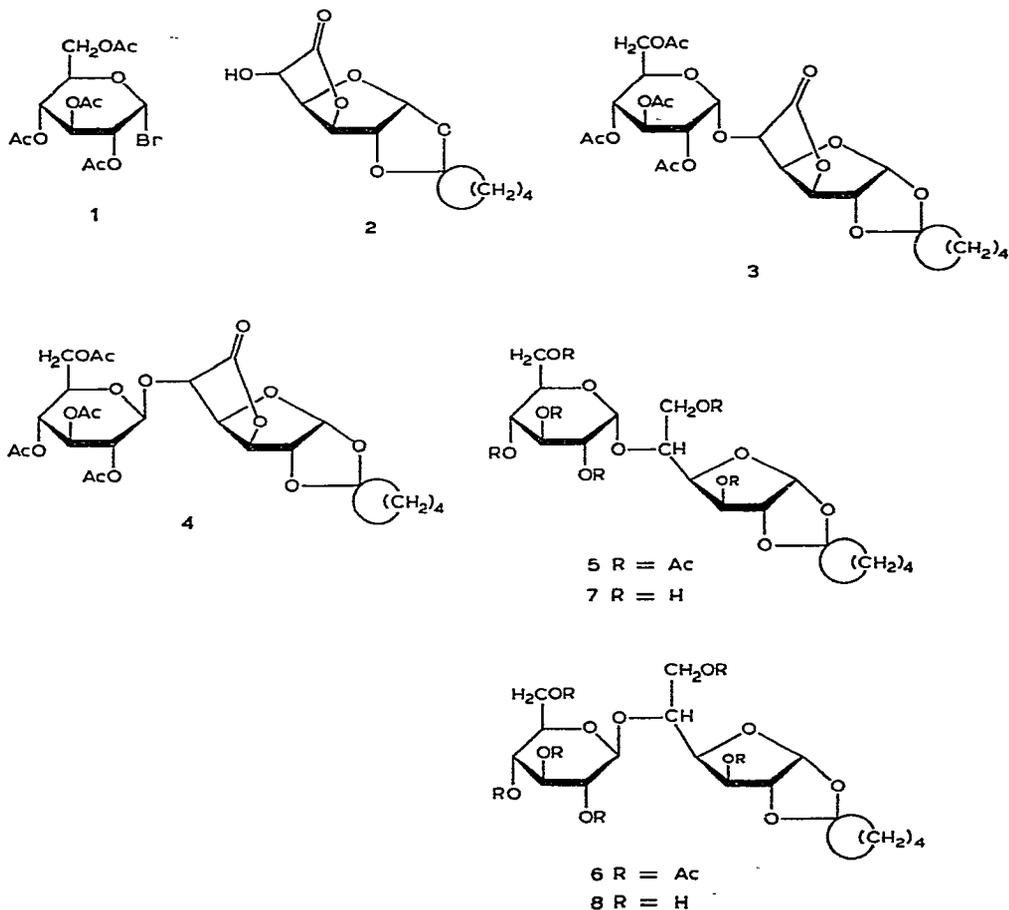


TABLE I

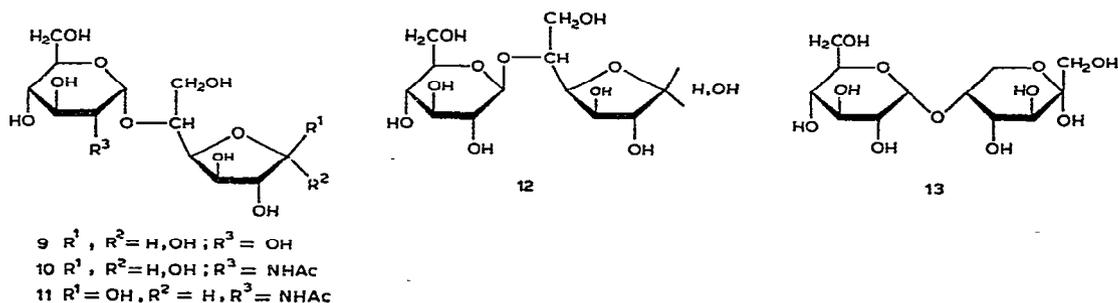
G.L.C. DATA<sup>a</sup> FOR THE *O*-TRIMETHYLSILYL DERIVATIVES OF **9**, **12**, AND **13**, AND THE RESPECTIVE ALDITOLS<sup>b</sup> (IN PARENTHESES)

	SE-30 (3.8%) T <sub>s</sub>	OV-17 (3%) T <sub>s</sub>
<b>12</b>	1.12 1.24 (1.58)	1.15 1.26 (1.61)
<b>9</b>	1.13 1.22 (1.98)	1.13 1.23 (2.01)
synthetic <b>13</b>	1.46 1.67 (1.98) <sup>c</sup>	1.47 1.73 (2.01) <sup>c</sup>
authentic <b>13</b>	1.46 1.67 (1.98) <sup>c</sup>	1.47 1.73 (2.01) <sup>c</sup>

<sup>a</sup>The retention times (T<sub>s</sub>) are given relative to per-*O*-Me<sub>3</sub>Si-sucrose; error ±0.03. <sup>b</sup>Prepared by sodium borohydride reduction of the respective disaccharides. <sup>c</sup>A shoulder attributable to per-*O*-Me<sub>3</sub>Si-5-*O*- $\alpha$ -D-glucopyranosyl-D-mannitol was observed on the positive slope of the peak.

trimethylsilylated disaccharide alditols derived from **9** and authentic leucrose were identical. The  $J_{1,2}$  values for the furanose moieties in **9** and **12** accorded with those observed<sup>6</sup> for **10**.

The isomerization of **9** in saturated, oxygen-free calcium hydroxide solution was monitored directly by t.l.c., and by g.l.c. of the trimethylsilylated derivatives, polarimetry, and <sup>1</sup>H-n.m.r. spectroscopy [Ca(OD)<sub>2</sub> in deuterium oxide]. The <sup>1</sup>H-n.m.r. spectrum immediately recorded after dissolution of **9** revealed the  $\alpha$ -furanoid form exclusively, presumably because of complex formation with calcium ions at O-1 and O-2, which are *cis* in the  $\alpha$ -form. A similar phenomenon has been observed for 5-*O*-methyl-D-ribofuranose<sup>11</sup>. In the isomerization reaction, no anomeric protons attributable to mannofuranose were observed, and **9** was almost quantitatively consumed within 3 h, yielding mainly **13**. At intermediate reaction times, the <sup>1</sup>H-n.m.r. spectra and g.l.c. traces were those of **9** and **13** superimposed. However, longer reaction times produced other components (t.l.c.) which were not further investigated. No mannose could be detected after methanolysis of crude **13** and subsequent g.l.c. analysis<sup>12</sup>.



Thus, the formation of the 1,2-enediol from **9** is extremely rapid in comparison to its back-formation from **13**. The anomerisation of **11** (*cf.* **9**) is instantaneous in water<sup>6</sup>, indicating that the furanose hemiacetal is in rapid equilibrium with the corresponding pseudo-acyclic intermediate, which was postulated<sup>13</sup> to be the immediate precursor for the 1,2-enediol produced in alkaline media. Conversely, the anomerization of fructose has been ascribed largely to pyranose–furanose interconversion rather than to  $\alpha \rightarrow \beta$  pyranose change<sup>14</sup>. The slow mutarotations of leucrose<sup>1</sup> and other 5-substituted fructopyranoses<sup>14,15</sup> have been cited to support this hypothesis. The apparently negligible back-formation of the 1,2-enediol from **13** substantiates the concept that the main factor controlling the rate of enolization is the concentration of an intermediate pseudo-acyclic form<sup>13</sup>.

Selection of optimal reaction conditions permitted the isolation of a product (53%) which was identical with authentic **13** (see also the data in Table I).

The foregoing procedure offers an attractive route for the preparation of leucrose-type oligosaccharides<sup>3</sup>, and increases the utility of D-glucofuranurono-6,3-lactone<sup>6,7,16</sup> for the preparation of (1  $\rightarrow$  5)-linked disaccharides.

#### EXPERIMENTAL

*Materials.* — Leucrose was a gift from Dr. F. H. Stodola (Peoria, Illinois).

*General methods.* — Melting points were determined on a Mettler FP5/FP51 photoelectric apparatus. Specific rotations were determined at ambient temperature with a Perkin–Elmer 141 Polarimeter. Half-lives of the cyclopentylidene acetals were determined as described earlier<sup>7</sup>. <sup>1</sup>H-N.m.r. spectra (internal Me<sub>4</sub>Si or sodium 2,2-dimethyl-2-silapentane-5-sulphonate, as appropriate) were recorded with a Varian EM390 (90 MHz) spectrometer, and i.r. spectra with a Pye Unicam SP1100 spectrophotometer. Solutions were concentrated at 40° (bath)/~14 mmHg. T.l.c. was performed on silica gel (Schleicher & Schüll TLC Ready Plastic Foil FR-1500) with conventional detection by charring with sulphuric acid. Column chromatography was performed on silica gel (Merck Kieselgel 60, 230–400 mesh) with *A*, chloroform; *B*, chloroform–methanol (25:1); *C*, propan-2-ol–ethyl acetate–water (2:2:1); and *D*, acetic acid–ethyl acetate–water–butan-1-ol (6:3:1:8). G.l.c. of trimethylsilylated derivatives<sup>17</sup> of sugars was carried out on a Pye 104 instrument equipped with flame-ionization detector and glass columns (1.60 m  $\times$  4 mm) packed with 3.8% of SE-30 or 3% of OV-17 on Chromosorb W-AW DMCS (80–100 mesh) (oven temperature, 227°; nitrogen flow-rate, 40 ml/min). The retention times (*T*<sub>S</sub>) are given relative to that of trimethylsilylated sucrose.

*1,2-O-Cyclopentylidene-5-O-(2,3,4,6-tetra-O-acetyl- $\alpha$ -D-glucopyranosyl)- $\alpha$ -D-glucofuranurono-6,3-lactone (3) and 1,2-O-cyclopentylidene-5-O-(2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl)- $\alpha$ -D-glucofuranurono-6,3-lactone (4).* — A mixture of 2,3,4,6-tetra-O-acetyl- $\alpha$ -D-glucopyranosyl bromide (**1**; 16.4 g, 40 mmol), mercuric cyanide (5.04 g, 20 mmol), mercuric bromide (7.2 g, 20 mmol), 1,2-O-cyclopentylidene- $\alpha$ -D-glucofuranurono-6,3-lactone<sup>7</sup> (**2**; 10 g, 41.3 mmol), and dry acetonitrile

(100 ml) was kept at room temperature for 1.5 h in the dark, and then concentrated. The residue was extracted with chloroform (4 × 50 ml), and the combined extracts were shaken with M potassium bromide (3 × 100 ml), dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated to a syrup. Addition of ether afforded a crystalline 1 : 1 mixture (16 g) of 3 and 4. The remaining mother liquor was subjected to column chromatography (solvent *A*) and gave an additional crop of 3 + 4 (6 g) (Found: C, 51.08; H, 5.36. C<sub>25</sub>H<sub>32</sub>O<sub>15</sub> calc.: C, 52.45; H, 5.63%).

Fractional crystallization of 3 + 4 from ethanol (2 g/100 ml), in which the former was more soluble, was monitored by t.l.c. (solvent *B*) and yielded 3 (10.6 g, 45%; *R<sub>F</sub>* 0.6), m.p. 129–131°, [ $\alpha$ ]<sub>D</sub> +136° (*c* 2.5, chloroform),  $\nu_{\max}^{\text{KBr}}$  1820 (C=O, lactone) and 1750 cm<sup>-1</sup> (C=O, acetyl). <sup>1</sup>H-N.m.r. data (CDCl<sub>3</sub>):  $\delta$  1.6–1.9 (m, 8 H, cyclopentylidene-CH<sub>2</sub>); 2.00, 2.03, 2.06, and 2.09 (4 s, 12 H, 4 AcO); 5.97 (d, *J*<sub>1,2</sub> 3.8 Hz, H-1); 4.74 (d, *J*<sub>2,1</sub> 3.8 Hz, H-2); 4.53 (d, *J*<sub>5,4</sub> 4.4 Hz, H-5); 5.36 (d, *J*<sub>1',2'</sub> 3.9 Hz, H-1'); and 4.0–5.7 (m, 8 H, other protons) (Found: C, 52.40; H, 5.59%). Compound 4 (9.0 g, 38%; *R<sub>F</sub>* 0.5) had m.p. 108–110°, [ $\alpha$ ]<sub>D</sub> +6.5° (*c* 2.5, chloroform),  $\nu_{\max}^{\text{KBr}}$  as for 3. <sup>1</sup>H-N.m.r. data (CDCl<sub>3</sub>):  $\delta$  1.6–1.9 (m, 8 H, cyclopentylidene-CH<sub>2</sub>); 2.00, 2.02, 2.06, and 2.08 (4 s, 12 H, 4 AcO); 5.96 (d, *J*<sub>1,2</sub> 3.8 Hz, H-1); 4.71 (d, *J*<sub>2,1</sub> 3.8, *J*<sub>2,3</sub> ~0 Hz, H-2); 4.79 (d, *J*<sub>3,4</sub> 3.0 Hz, H-3); 5.59 (d, *J*<sub>5,4</sub> 4.4 Hz, H-5); 4.20 (d, 2 H, *J*<sub>6a',5'</sub> = *J*<sub>6b',5'</sub> = 3.6 Hz, H-6a'b'); 3.73 (m, 1 H, H-5'β); and 4.9–5.2 (m, 5 H, other protons) (Found: C, 52.30; H, 5.61%).

*3,6-Di-O-acetyl-1,2-O-cyclopentylidene-5-O-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-α-D-glucofuranose* (6). — A solution of 4 (5.1 g, 8.9 mmol) in 1,2-dimethoxyethane (180 ml) was flushed with nitrogen, and lithium aluminium hydride (2.2 g, 58 mmol) was added in portions with stirring. The mixture was then stirred for 3 h at 45–50°, and cooled to 10°, and the excess of hydride was decomposed with ethyl acetate (27 ml). The addition of 50% aqueous methanol (220 ml) gave a precipitate that was collected over diatomaceous earth and thoroughly washed with methanol. The filtrates were combined, the pH was adjusted to 5–6 with hydrochloric acid, and the solution was concentrated. A solution of the residue in water (1 litre) was freeze-dried, and the residue was conventionally acetylated with pyridine (80 ml) and acetic anhydride (40 ml) for 16 h at 5°, and 6 h at room temperature, to yield 6 (3.46 g, 59%), m.p. 111–113° (from ether), [ $\alpha$ ]<sub>D</sub> -16° (*c* 3, chloroform),  $\nu_{\max}^{\text{KBr}}$  1750 cm<sup>-1</sup> (C=O, acetyl); no lactone absorption was observed (Found: C, 52.86; H, 6.07. C<sub>29</sub>H<sub>40</sub>O<sub>17</sub> calc.: C, 52.73; H, 6.10%).

*3,6-Di-O-acetyl-1,2-O-cyclopentylidene-5-O-(2,3,4,6-tetra-O-acetyl-α-D-glucopyranosyl)-α-D-glucofuranose* (5). — Compound 3 (4.6 g, 8.0 mmol) was reduced with lithium aluminium hydride and acetylated as described above for 4. The resulting, viscous syrup was eluted from a column (15 × 5 cm) of silica gel with solvent *A* to afford 5 (4.6 g, 86%), as a brittle foam, [ $\alpha$ ]<sub>D</sub> +50° (*c* 2.5, chloroform),  $\nu_{\max}^{\text{film}}$  as for 6 (Found: C, 52.95; H, 6.15%).

*1,2-O-Cyclopentylidene-5-O-α-D-glucopyranosyl-α-D-glucofuranose* (7). — A solution of 5 (5 g, 7.6 mmol) in 0.02M methanolic sodium methoxide (200 ml) was monitored by t.l.c. (solvents *B* and *C*) until reaction was complete, neutralized with

Amberlite IR-120 ( $H^+$ ) resin, and then treated with a mixed-bed ( $H^+$ ,  $HO^-$ ) resin (100 mg), filtered, and concentrated. A solution of the residue in water (500 ml) was passed through a fine sintered-glass funnel and freeze-dried, affording amorphous **7** (3.0 g, 97%),  $[\alpha]_D +81^\circ$  (*c* 1.5, water).  $^1H$ -N.m.r. data ( $D_2O$ ):  $\delta$  6.10 (d,  $J_{1,2}$  3.9 Hz, H-1 $\alpha$ ), 5.25 (d,  $J_{1',2'}$  3.8 Hz, H-1' $\alpha$ ), and 4.75 (d,  $J_{1,2}$  3.9 Hz, H-2) (Found: C, 49.77; H, 6.80,  $C_{17}H_{28}O_{11}$  calc. C, 50.00; H, 6.91 %).

*1,2-O-Cyclopentylidene-5-O- $\beta$ -D-glucopyranosyl- $\alpha$ -D-glucofuranose (8)*. — Treatment of **6** (3.4 g, 5.2 mmol) as described above for **5** yielded **8** (2.0 g, 95%),  $[\alpha]_D +3^\circ$  (*c* 1.5, water).  $^1H$ -N.m.r. data ( $D_2O$ ):  $\delta$  6.01 (d,  $J_{1,2}$  3.8 Hz, H-1 $\alpha$ ), 4.64 (d,  $J_{2,1}$  3.8 Hz, H-2), and 4.66 (d,  $J_{1',2'}$  7.5 Hz, H-1' $\beta$ ) (Found: C, 49.88; H, 6.80 %).

*5-O- $\alpha$ -D-Glucopyranosyl- (9) and 5-O- $\beta$ -D-glucopyranosyl-D-glucofuranose (12)*. — The hydrolyses of **7** and **8** (1.5 g, 3.7 mmol) in 0.52M sulphuric acid (75 ml) were monitored at  $20^\circ$  by t.l.c. (solvent *D*) and polarimetry<sup>7</sup> [ $t_{0.5} \sim 1.7$  h,  $\partial\alpha/\partial t < 0$ , consistent with a (1  $\rightarrow$  5)-linkage<sup>7</sup> in **7** and **8**; Freudenberg<sup>9</sup> reported  $\partial\alpha/\partial t < 0$  for the isopropylidene analogue of **8**]. After 9.5–10 h, the reaction was quenched with ice and subsequent neutralization with Dowex 1 X2 ( $HCO_3^-$ ) resin. Each mixture was filtered and freeze-dried to yield **9** (95%),  $[\alpha]_D +69.5^\circ$  (*c* 2, water), and **12** (94%),  $[\alpha]_D -25^\circ$  (*c* 2, water). Each product was subjected to ion-exchange chromatography<sup>18</sup> on a column (1.5 m  $\times$  2 cm) of Dowex 50 X4 ( $K^+$ ) resin to remove any traces of glucose and **7** or **8**, and isolated by freeze-drying.

Compound **9** had  $[\alpha]_D +71^\circ$  (*c* 2, water); for g.l.c. data, see Table I. The  $^1H$ -n.m.r. spectrum was similar to that of **10**<sup>6</sup> and indicated a 1:1  $\alpha\beta$ -mixture:  $\delta$  5.51 (d,  $J_{1,2}$  4.0 Hz, furanoid H-1 $\alpha$ ), 5.14 (d,  $J_{1',2'}$  3.6 Hz, H-1' $\alpha'$ ), 5.24 (d,  $J_{1,2}$  0.8 Hz, furanoid H-1 $\beta$ ), and 5.18 (d,  $J_{1',2'}$  3.6 Hz, H-1' $\alpha$ ). Compound **9** is best characterized by its facile conversion into crystalline **13**.

Compound **12** had  $[\alpha]_D -28^\circ$  (*c* 2, water); lit.<sup>9,10</sup>  $[\alpha]_D -22^\circ$  and  $-23^\circ$  (*c* 1, water) for syrupy **12**. For g.l.c. data, see Table I. The  $^1H$ -n.m.r. spectrum ( $D_2O$ ) was consistent with a 1:1  $\alpha\beta$ -mixture:  $\delta$  5.53 (d,  $J_{1,2}$  3.6 Hz, furanoid H-1 $\alpha$ ), 4.66 (d,  $J_{1',2'}$  7.5 Hz, H-1' $\beta$ ), 5.22 (d,  $J_{1,2} < 0.5$  Hz, furanoid H-1 $\beta$ ), and 4.70 (d,  $J_{1',2'}$  7.5 Hz, H-1' $\beta$ ). Treatment<sup>9</sup> of **12** with toluene-*p*-sulphonylhydrazine gave the hydrazone, m.p. 178–179° (dec.),  $[\alpha]_D -21^\circ$  (*c* 1, 4:1 pyridine–water); lit.<sup>10</sup> m.p. 180–181° (dec.),  $[\alpha]_D -22^\circ$ .

*5-O- $\alpha$ -D-Glucopyranosyl-D-fructopyranose (leucrose, 13)*. — Compound **9** (630 mg, 1.84 mmol) was treated in an atmosphere of nitrogen with oxygen-free 0.044M calcium hydroxide<sup>19</sup> (7 ml) at  $35^\circ$ . The  $[\alpha]_{546}$  value rapidly dropped from  $+7^\circ$  (extrapolated  $t_0$  value) to  $\sim 0.5^\circ$  in 3 h. At intervals, aliquots (5  $\mu$ l) were neutralized with 50% aqueous acetic acid and subjected to t.l.c. [solvent *C*; **9** ( $R_F$  0.4)  $\rightarrow$  **13** ( $R_F$  0.23)] and g.l.c. analysis (trimethylsilyl derivatives, Table I). After 3 h, the ratio of **9** and **13** was  $\sim 1:9$ . The reaction mixture was quenched with ice and the required amount of Amberlite IR-120 ( $H^+$ ) resin, and then filtered and freeze-dried. The product was subjected to ion-exchange chromatography<sup>18</sup> as described above, isolated by freeze-drying, and crystallized from methanol–ethanol to give **13** (331 mg, 53%), m.p. and mixture m.p. 156–157.5°,  $[\alpha]_D -7^\circ$  (*c* 2, water); lit.<sup>1</sup> m.p. 156–157°,

$[\alpha]_D -7.6^\circ$  (c 4, water) (Found: C, 41.70; H, 6.50.  $C_{12}H_{22}O_{11}$  calc.: C, 42.11; H, 6.48%). The  $^1H$ -n.m.r. data ( $D_2O$ ) [ $\delta$  5.13 (d,  $J_{1',2'}$ , 3.9 Hz, H-1' $\alpha$ ) and 3.2–4.2 (13 H, other protons)] for synthetic and authentic **13** were indistinguishable.

Compound **9** (50 mg) was treated with an oxygen-free, saturated solution (0.6 ml) of calcium deuterioxide in deuterium oxide<sup>19</sup>.  $^1H$ -N.m.r. spectroscopy [1,4-dioxane ( $\delta$  3.56) as internal reference] revealed the  $\alpha$  form of **9** exclusively:  $\delta$  5.18 (d, 1 H,  $J_{1,2}$  2.3 Hz, H-1 $\alpha$ ) and 4.94 (d, 1 H,  $J_{1',2'}$  3.6 Hz, H-1' $\alpha$ ). Both doublets disappeared during 3 h, and a new doublet at  $\delta$  4.89 ( $J_{1',2'}$  3.8 Hz, H-1' $\alpha$  in **13**) reached maximum intensity. No other signals were observed in the anomeric region of the spectra. The reaction was quenched as described above, and a sample (1 mg) of crude **13** was subjected to methanolysis and g.l.c. analysis as devised by Clamp *et al.*<sup>12</sup>; no mannose was detected.

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#### REFERENCES

- 1 F. H. STODOLA, E. S. SHARPE, AND J. KOEPSSELL, *J. Am. Chem. Soc.*, **78** (1956) 2514–2518.
- 2 R. W. BAILEY AND E. J. BOURNE, *Nature (London)*, **184** (1959) 904–905.
- 3 E. J. BOURNE, D. H. HUTSON, AND H. WEIGEL, *Biochem. J.*, **79** (1961) 549–553.
- 4 G. MAGHUIN-ROGISTER, *Bull. Soc. Chim. Belg.*, **77** (1968) 575–578.
- 5 J. JADOT AND G. MAGHUIN-ROGISTER, *Bull. Soc. Chim. Belg.*, **77** (1968) 569–574.
- 6 W. A. R. VAN HEESWIJK, P. DE HAAN, AND J. F. G. VLIEGENTHART, *Carbohydr. Res.*, **48** (1976) 187–196.
- 7 W. A. R. VAN HEESWIJK, J. B. GOEDHART, AND J. F. G. VLIEGENTHART, *Carbohydr. Res.*, **58** (1977) 337–344.
- 8 N. K. KOCHETKOV, A. J. KHORLIN, AND A. F. BOCHKOV, *Tetrahedron*, **23** (1967) 693–707.
- 9 K. FREUDENBERG AND K. OERTZEN, *Justus Liebig's Ann. Chem.*, **574** (1951) 37–53.
- 10 J. C. SOWDEN AND A. S. SPRIGGS, *J. Am. Chem. Soc.*, **78** (1956) 2503–2505.
- 11 S. J. ANGYAL AND K. P. DAVIES, *Chem. Commun.*, (1971) 500–501.
- 12 J. P. KAMERLING, G. J. GERWIG, J. F. G. VLIEGENTHART, AND J. R. CLAMP, *Biochem. J.*, **151** (1975) 491–495.
- 13 H. S. ISBELL, H. L. FRUSH, C. W. R. WADE, AND C. E. HUNTER, *Carbohydr. Res.*, **9** (1969) 163–175.
- 14 H. S. ISBELL AND W. W. PIGMAN, *J. Res. Natl. Bur. Stand.*, **20** (1938) 773–776.
- 15 D. J. BELL, *J. Chem. Soc.*, (1953) 1231–1233.
- 16 H. WEIDMANN, M. APPENROTH, R. LEIPERT-KLUG, K. DAX, AND P. STÖCKL, *J. Carbohydr. Nucleos. Nucleot.*, **3** (1976) 235–260.
- 17 J. P. KAMERLING, J. F. G. VLIEGENTHART, J. VINK, AND J. J. DE RIDDER, *Tetrahedron*, **27** (1971) 4275–4288.
- 18 R. M. SAUNDERS, *Carbohydr. Res.*, **7** (1968) 76–79.
- 19 E. M. MONTGOMERY AND C. S. HUDSON, *J. Am. Chem. Soc.*, **52** (1930) 2101–2105.