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Detectable A Disintegrin and Metalloproteinase With Thrombospondin Motifs-1 in Serum Is Associated With Adverse Outcome in Pediatric Sepsis

IMPORTANCE: A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 is hypothesized to play a role in the pathogenesis of invasive infection, but studies in sepsis are lacking.

OBJECTIVES: To study A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 protein level in pediatric sepsis and to study the association with outcome.

DESIGN: Data from two prospective cohort studies.

SETTING AND PARTICIPANTS: Cohort 1 is from a single-center study involving children admitted to PICU with meningococcal sepsis (samples obtained at three time points). Cohort 2 includes patients from a multicenter study involving children admitted to the hospital with invasive bacterial infections of differing etiologies (samples obtained within 48 hr after hospital admission).

MAIN OUTCOMES AND MEASURES: Primary outcome measure was mortality. Secondary outcome measures were PICU-free days at day 28 and hospital length of stay.

RESULTS: In cohort 1 ($n = 59$), nonsurvivors more frequently had A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 levels above the detection limit than survivors at admission to PICU (8/11 [73%] and 6/23 [26%], respectively; $p = 0.02$) and at $t = 24$ hours (2/3 [67%] and 3/37 [8%], respectively; $p = 0.04$). In cohort 2 ($n = 240$), A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 levels in patients within 48 hours after hospital admission were more frequently above the detection limit than in healthy controls (110/240 [46%] and 14/64 [22%], respectively; $p = 0.001$). Nonsurvivors more often had detectable A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 levels than survivors (16/21 [76%] and 94/219 [43%], respectively; $p = 0.003$), which was mostly attributable to patients with *Neisseria meningitidis*.

CONCLUSIONS AND RELEVANCE: In children with bacterial infection, detection of A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 within 48 hours after hospital admission is associated with death, particularly in meningococcal sepsis. Future studies should confirm the prognostic value of A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 and should study pathophysiologic mechanisms.

KEY WORDS: A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1 protein; bacterial infections; biomarkers; inflammation; mortality; sepsis

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Prevalence and outcome of bacterial infections are determined by host (e.g., genetic predisposition, immune response to bacteria), pathogen, and healthcare system factors (1). The European Childhood Life-threatening Infectious Diseases Study (EUCLIDS) aims to identify genetic

factors and biological pathways associated with susceptibility and/or severity of life-threatening bacterial infections (2–4). Preliminary EUCLIDS genetic studies in meningococcal sepsis patients identified a SNP in *A Disintegrin and Metalloproteinase with Thrombospondin Motifs-1* (*ADAMTS-1*; rs9975310) to be associated with disease severity, although this association did not reach genome-wide significance (unpublished data). Furthermore, animal studies showed that *ADAMTS-1* is increased in the host inflammatory response, and therefore we hypothesize that *ADAMTS-1* plays a role in the pathogenesis of invasive infection (5–7). Studies on *ADAMTS-1* in sepsis, either adult or pediatric, are currently lacking.

The *ADAMTS* family includes 19 proteases with a variety of functions, for example, in coagulation and inflammation (8–11). *ADAMTS-13*, the von Willebrand factor (vWF)-cleaving protease, cleaves ultra large prothrombotic multimeric vWF into an optimal size for normal coagulation (12) and is the most extensively studied *ADAMTS* protease in sepsis. Previous studies demonstrated that decreased *ADAMTS-13* levels, presumably leading to increased formation of thrombi, are associated with more severe disease and poor outcome (13, 14).

Other *ADAMTS*-proteins have not been studied in sepsis yet despite animal studies hinting toward an important role in inflammation and sepsis. *ADAMTS-1* is an inhibitor of angiogenic activity, is associated with acute inflammatory processes, and is involved in the process of extracellular matrix damage and repair (7, 15). In rats and mice, a dramatic increase of *ADAMTS-1* was detected after lipopolysaccharide (LPS) induced systemic inflammation, suggesting that the *ADAMTS-1* gene is an inflammation-associated gene (5–7). An immunomodulatory role for *ADAMTS-1* is also indicated by the pro-inflammatory phenotype observed in *ADAMTS-1*-deficient mice (16).

We studied *ADAMTS-1* serum protein levels in pediatric sepsis and studied the association with mortality, illness severity, coagulation, and infecting pathogen.

MATERIALS AND METHODS

This study comprises data from two independent cohorts; a single-center cohort of children admitted to PICU with meningococcal sepsis (cohort 1) and an international, multicenter cohort of children admitted to hospital with invasive bacterial infections of differing etiologies (cohort 2).

Cohort 1

Children 1 month to 18 years old with meningococcal sepsis presenting to the PICU of Erasmus MC-Sophia Children's Hospital between October 1991 and February 2000 were prospectively enrolled in meningococcal sepsis studies (17–19). All patients fulfilled internationally agreed criteria for sepsis with petechial rash and/or purpura (20). Blood samples were collected at admission to PICU, at 24 hours, and at 1 month after PICU admission.

Serum samples were processed on ice and stored at -80°C until analysis. In remaining serum samples available from these studies, we measured *ADAMTS-1* levels using a commercially available human enzyme-linked immunosorbent assay (ELISA) kit as described by the manufacturer (*ADAMTS-1* ELISA kit, MBS2021525; MyBioSource, San Diego, CA). The lower limit of detection (LLOD) of this assay was 1.6 ng/mL (1,600 pg/mL). *ADAMTS-1* levels measured below the LLOD were considered 1.6 ng/mL.

The samples obtained 1 month after PICU admission were considered as convalescent samples.

Cohort 2

Children suspected of community-acquired bacterial infection at hospital admission were prospectively enrolled between July 2012 and December 2016. This multicenter cohort study (EUCLIDS) involves 195 hospitals from 10 countries. Detailed information on consortium and enrollment strategy has been published elsewhere (2, 3). Patients were recruited as early as possible in the illness within a time window from admission to hospital to the time when microbiology results became available.

For this laboratory study, we selected children 1 month to 18 years old recruited in five countries (United Kingdom, Spain, Austria, The Netherlands, and Switzerland) with an invasive infection caused by *Neisseria meningitidis*, *Streptococcus pneumoniae*, *Staphylococcus aureus*, or group A streptococcus, from whom serum samples obtained within 48 hours after hospital admission were available. Invasive bacterial infection was defined as isolation by culture or polymerase chain reaction of a bacterial organism from a normally sterile site. We considered blood, cerebrospinal fluid, urine, bronchoalveolar lavage, joint aspirate, abscess aspirate, intraoperative swabs, and pleural aspirate as sterile sites. Positive cultures from sites

such as endotracheal tube aspirate, nasopharyngeal aspirate, throat/nasal swabs, and wounds were not considered as sterile sites.

ADAMTS-1 levels were measured with a custom-made Luminex assay based on a capture antibody, detection antibody, and recombinant human ADAMTS-1 (ADAMTS-1 DuoSet ELISA assay; R&D Systems, Abingdon, United Kingdom). This Luminex assay, being a far more sensitive assay than the ELISA used for cohort 1, had a LLOD of 7.0 pg/mL.

For comparison, EUCLIDS recruited healthy controls from whom serum was obtained prior to elective surgical procedures. The controls did not have any underlying inflammatory comorbidity.

Ethical Aspects

This study was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice guidelines. The Erasmus MC—meningococcal sepsis study was approved by the ethical committee of Erasmus MC (MEC-2015-497), and the EUCLIDS study protocol was approved by at least one ethical review board in every participating country (Coordinating Center Research Ethics Committee reference: 11/LO/1982). Written informed consent was obtained from parents or legal guardians.

Clinical Data Collection

Data for both cohorts were collected prospectively. Illness severity and risk of mortality were assessed by the Pediatric Risk of Mortality (PRISM) score (21), Pediatric Index of Mortality 2 (PIM 2) (22), need for ventilation and/or inotropes, predicted death based on the base excess and platelet count at presentation score (23), predicted death based on the Rotterdam score (17), and disseminated intravascular coagulation (DIC) score (24). Coagulation and inflammation markers were measured for clinical reasons or measured as requirement for other meningococcal sepsis studies to which patients had been recruited (17, 18, 25). For the multicenter EUCLIDS study, monthly telephone conferences, biannual meetings, clinical protocols including case definitions, data audits, and monitoring ensured uniform procedures among study sites.

Outcome Measures

The primary outcome measure was mortality. Patients were classified as deceased if death occurred during

hospital stay. Secondary outcome measures were PICU-free days at day 28 (days alive and free from the need for intensive care) and hospital length of stay. PICU-free days in patients who died were considered zero.

Statistical Analysis

Categorical variables are presented as counts (percentages). We used the chi-square test (or Fisher exact test in case the number of events in one group was < 5) to compare frequency distributions between two categorical variables. Continuous variables with normal distribution are presented as mean (\pm SD); non-normally distributed variables are reported as median (interquartile range [IQR]). We tested differences between groups with analysis of variance or Kruskal-Wallis and Student *t* test or Mann-Whitney *U* test, as appropriate. In the cohort 1, Friedman tests were used to compare ADAMTS-1 levels between three time points. Correlations between ADAMTS-1 level and secondary outcome measures, illness severity, coagulation markers, and inflammatory markers were assessed by Spearman rank correlation. Post hoc Bonferroni correction for multiple testing was applied. Statistical analyses were performed with SPSS Version 21 (Armonk, NY). Graphs were created with GraphPad Prism 8.4.0 (GraphPad Software, Inc.). A *p* value of less than 0.05 was considered statistically significant.

RESULTS

Cohort 1

We included 59 children admitted to PICU with meningococcal sepsis, of whom 11 (19%) died, who had 109 samples available for ADAMTS-1 measurements. Patient characteristics are shown in **Table 1**.

Because ADAMTS-1 levels of 90 of 108 samples (83%) were below the LLOD of the assay (1.6 ng/mL), we compared the number of samples with detectable ADAMTS-1 (designated as ADAMTS-1 \geq 1.6 ng/mL) with the number of samples with undetectable ADAMTS-1. At admission to PICU, ADAMTS-1 was detectable in 14 of 34 patients (41%), which was more frequent than in patients at *t* = 24 hours (5/40 [12%]; *p* = 0.005) and at *t* = 1 month (0/35 [0%]; *p* < 0.001) (**Table 2**). Nonsurvivors more frequently had detectable ADAMTS-1 levels compared with survivors at admission to PICU (nonsurvivors 8/11 [73%], survivors 6/23 [26%]; *p* = 0.02) and at *t* = 24 hours (nonsurvivors 2/3 [67%], survivors 3/37 [8%]; *p* = 0.04).

TABLE 1.
Baseline Characteristics Cohort 1 and Cohort 2

Variable	Cohort 1	Cohort 2	Cohort 2	Cohort 2
	Patients (n = 59)	Patients (n = 240)	Controls (n = 64)	p
Male	35 (59%)	131 (55%)	34 (53%)	NS
Age	3.0 yr (1.8–9.7 yr)	3.4 yr (15 mo–9.2 yr)	5.4 yr (2.8–12.4 yr)	< 0.01
Ethnicity ^a				NS
African/North African	–	12 (5%)	8 (13%)	
Asian	–	13 (6%)	4 (6%)	
European	–	185 (81%)	47 (75%)	
Meso/South American	–	4 (2%)	0 (0%)	
Middle Eastern	–	3 (1%)	2 (3%)	
Other/mixed	–	12 (5%)	2 (3%)	
Number of underlying conditions				< 0.01
None	–	138 (58%)	25 (39%)	
≥ 1	–	102 (42%)	39 (61%)	
Immunizations up to date ^b	–	178 (95%)	58 (95%)	NS
Illness severity				
Sepsis	–	159 (66%)		
PICU admission	59 (100%)	177 (74%)		
Need for inotropes ^c	42 (95%)	107 (53%)		
Days on inotropes	–	3 (2–5)		
Need for invasive ventilation ^d	31 (60%)	106 (52%)		
Days on invasive ventilation	–	4 (3–7)		
Need for extracorporeal membrane oxygenation ^e	–	4 (3%)		
Pediatric Risk of Mortality (21) score ^f	20 (14–26)	11 (7–16)		
Pediatric Index of Mortality 2 (22) score ^g (predicted death, %)	–	3.5% (0.8–11.6)		
Predicted death rate based on the base excess and platelet count at presentation score (23) ^h	6.1 (3.4–19.8)	–		
Predicted death rate based on the Rotterdam score (17) ⁱ	12.2 (1.6–77.0)	–		
Lactate (mmol/L) ^h	4.5 (3.3–6.5)	–		
DIC score ^j	5 (4–7)	2 (0–2)		
Presence of DIC (DIC score ≥ 5) ⁱ	22 (61%)	18 (10%)		

(Continued)

TABLE 1.(Continued).
Baseline Characteristics Cohort 1 and Cohort 2

Variable	Cohort 1	Cohort 2	Cohort 2	Cohort 2
	Patients (n = 59)	Patients (n = 240)	Controls (n = 64)	p
Outcome				
PICU-free days at day 28 (d) ^k	23 (13–25)	23 (19–25)		
Hospital length of stay (d) ^l	7 (3–12)	10 (6–17)		
Death	11 (19%)	21 (9%)		

DIC = disseminated intravascular coagulation (24, 26), NS = not significant.

^aEthnicity data were available for 229/240 patients and 63/64 controls.

^bImmunization data were available for 188/240 patients and 61/64 controls.

^cData on inotropes were available for 44/59 patients in cohort 1 and 202/240 patients in cohort 2.

^dData on invasive ventilation were available for 52/59 patients in cohort 1 and 203/240 patients in cohort 2.

^eData on extracorporeal membrane oxygenation were available for 161/240 patients.

^fPediatric Risk of Mortality score (21) was available for 50/59 patients in cohort 1 and 150/240 patients in cohort 2.

^gPediatric Index of Mortality 2 score (22) was available for 177/240 patients.

^hPredicted death rate based on the base excess and platelet count at presentation score (23) and lactate were available for 52/59 patients.

ⁱPredicted death rate based on the Rotterdam score (17) was available for 48/59 patients.

^jData on DIC were available for 36/59 patients in cohort 1 and 187/240 patients in cohort 2.

^kData on PICU-free days at day 28 were available for 59/59 patients in cohort 1 and 177/177 PICU patients in cohort 2.

^lData on hospital length of stay were available for 53/59 patients in cohort 1 and 240/240 patients in cohort 2.

Values are reported as counts (percentages) or medians (interquartile ranges) unless stated otherwise. Dashes indicate data is on this variable is not available and no statistical test has been done.

TABLE 2.
Cohort 1; A Disintegrin and Metalloproteinase With Thrombospondin Motifs-1 Levels in Survivors and Nonsurvivors at Admission to PICU, at t = 24 Hours and at t = 1 Month

Time Point	All Patients (n = 59)	Survivors (n = 48)	Nonsurvivors (n = 11)	p
PICU admission	n = 34, 1.6 (1.6–2.1)	n = 23, 1.6 (1.6–1.6)	n = 11, 2.0 (1.6–3.1)	0.02 ^a
n < 1.6	n = 20 (59%)	n = 17 (74%)	n = 3 (27%)	0.02 ^b
n > 1.6	n = 14 (41%)	n = 6 (26%)	n = 8 (73%)	
t = 24 hr	n = 40, 1.6 (1.6–1.6)	n = 37, 1.6 (1.6–1.6)	n = 3, 2.1 (1.6–3.3)	Not significant ^a
n < 1.6	n = 35 (88%)	n = 34 (92%)	n = 1 (33%)	0.04 ^b
n > 1.6	n = 5 (12%)	n = 3 (8%)	n = 2 (67%)	
t = 1 mo	n = 35, 1.6 (1.6–1.6)	n = 35, 1.6 (1.6–1.6)	n = 0	–
n < 1.6	n = 35 (100%)	n = 35 (100%)	n = 0	–
n > 1.6	n = 0	n = 0	n = 0	

^aMann-Whitney *U* test.

^bFisher exact test.

A Disintegrin and Metalloproteinase With Thrombospondin Motifs-1 levels are presented as median (interquartile range).

For each time point, we additionally analyzed the number of samples below and above the lowest level of detection (1.6 ng/mL).

Dashes indicate data is on this variable is not available and no statistical test has been done.

Median ADAMTS-1 level at admission to PICU ($n = 34$; 1.6 ng/mL [IQR, 1.6–2.1 ng/mL]) did not differ from ADAMTS-1 level at $t = 24$ hours ($n = 40$; 1.6 ng/mL [IQR, 1.6–1.6 ng/mL]) or at $t = 1$ month ($n = 35$; 1.6 ng/mL [IQR, 1.6–1.6 ng/mL]; Friedman test $p = 0.37$) (Table 2). At admission to PICU, ADAMTS-1 levels in nonsurvivors ($n = 11$; 2.0 ng/mL [IQR, 1.6–3.1 ng/mL]) were significantly higher than in survivors ($n = 23$; 1.6 ng/mL [IQR, 1.6–1.6 ng/mL]; $p = 0.02$). Although numbers were low ($n = 40$), after 24 hours, there still was a trend for higher ADAMTS-1 levels in nonsurvivors ($n = 3$; 2.1 ng/mL [IQR, 1.6–3.3 ng/mL]) compared with survivors ($n = 37$; 1.6 ng/mL [IQR, 1.6–1.6 ng/mL]; $p = 0.09$).

ADAMTS-1 level at admission to PICU was not significantly correlated to PICU-free days at day 28 ($r = -0.31$; $p = 0.08$) and hospital length of stay ($r = -0.48$; $p = 0.01$) nor did they correlate with illness severity, coagulation markers, or inflammatory markers (Supplemental Digital Content, <http://links.lww.com/CCX/A845>).

Cohort 2

We included 240 children with an invasive infection caused by *N. meningitidis* ($n = 83$), *S. pneumoniae* ($n = 63$), *S. aureus* ($n = 50$), or group A streptococcus ($n = 44$), of which 21 children died (9%). Additionally, we included 64 controls (age ranged from 1 mo to 18 yr). Baseline characteristics are shown in Table 1, and baseline characteristics per pathogen are shown in Table 3.

ADAMTS-1 level within 48 hours after admission to hospital was more frequently detectable in patients (110/240 [46%]) compared with controls (14/64 [22%]; $p = 0.001$). Furthermore, although median values were similar, ADAMTS-1 level analyzed by the rank-sum test differed between patients and controls (patients: median 7.0 pg/mL [IQR, 7.0–118 pg/mL]; controls: median 7.0 pg/mL [IQR, 7.0–7.0 pg/mL]; $p < 0.001$) (Fig. 1). The elevation in ADAMTS-1 was more pronounced in PICU patients ($n = 177$; median, 11.7 pg/mL [IQR, 7.0–166 pg/mL]) than in non-PICU patients ($n = 63$; median, 7.0 pg/mL [IQR, 7.0–18.5 pg/mL]; $p = 0.001$).

Detection of ADAMTS-1 was more frequent in patients with *N. meningitidis* (41/83 [49%]; $p = 0.001$), *S. aureus* (21/50 [42%]; $p = 0.02$), and group A streptococcus (25/44 [57%]; $p < 0.001$) compared

with controls (14/64 [22%]). Detection in patients with *S. pneumoniae* (23/63 [37%]; $p = 0.07$) did not differ from controls. ADAMTS-1 level per pathogen group is depicted in Figure 1.

In samples taken within 48 hours after hospital admission, ADAMTS-1 was detected in nonsurvivors (16/21 [76%]) more frequently than in survivors (94/219 [43%]; $p = 0.003$). Furthermore, ADAMTS-1 levels in nonsurvivors (median, 260 pg/mL [IQR, 45–1,548 pg/mL]) were higher compared with survivors (median, 7.0 pg/mL [IQR, 7.0–96 pg/mL]; $p < 0.001$) (Fig. 2A).

This was attributable to patients with *N. meningitidis* (detectable nonsurvivors 7/7 [100%], detectable survivors 34/76 [45%]; $p = 0.005$). Median ADAMTS-1 level also differed between meningococcal infection nonsurvivors and survivors (nonsurvivors: median, 687 pg/mL [IQR, 120–4,108 pg/mL]; survivors: median, 7.0 pg/mL [IQR, 7.0–111 pg/mL]; $p < 0.001$) (Fig. 2B). In children with meningococcal infections, ADAMTS-1 level at admission to hospital was correlated to PICU-free days at day 28 ($r = -0.54$; $p < 0.001$), PRISM score ($r = 0.47$; $p < 0.001$), PIM 2 score ($r = 0.42$; $p < 0.001$), and plasminogen activator inhibitor-1 (PAI-1) ($r = 0.42$; $p < 0.001$), but not to hospital length of stay ($r = 0.12$; $p = 0.30$).

ADAMTS-1 detection did not differ significantly between survivors and nonsurvivors of infections with *S. pneumoniae* (detectable nonsurvivors 3/7 [43%], detectable survivors 20/56 [36%]; $p = 0.71$), *S. aureus* (detectable nonsurvivors 3/4 [75%], detectable survivors 18/46 [39%]; $p = 0.16$), and group A streptococcus (detectable nonsurvivors 3/3 [100%], detectable survivors 22/41 [54%]; $p = 0.12$). ADAMTS-1 levels in survivors and nonsurvivors per pathogen group are depicted in Figure 2B.

With regard to secondary outcome measures, ADAMTS-1 levels at admission to hospital were strongly correlated to PICU-free days at day 28 ($r = -0.36$; $p < 0.001$), PRISM score ($r = 0.37$; $p < 0.001$), DIC score ($r = 0.27$; $p < 0.001$), need for invasive ventilation ($r = 0.27$; $p < 0.001$), platelets ($r = -0.30$; $p < 0.001$), protein C ($r = -0.24$; $p < 0.001$), and PAI-1 ($r = 0.33$; $p < 0.001$), but less strongly with hospital length of stay ($r = -0.13$; $p = 0.05$) (Supplemental Digital Content, <http://links.lww.com/CCX/A845>).

Thus, in both cohorts, ADAMTS-1 in nonsurvivors was more frequently detectable and showed a

TABLE 3.
Baseline Characteristics Cohort 2 by Pathogen

Variable	All Patients (n = 240)	<i>Neisseria meningitidis</i> (n = 83)	<i>Streptococcus pneumoniae</i> (n = 63)	<i>Staphylococcus aureus</i> (n = 50)	Group A <i>Streptococcus</i> (n = 44)	p
Gender, male	131 (55%)	50 (60%)	39 (62%)	25 (50%)	17 (39%)	NS
Age	3.4 yr (15 mo–9.2 yr)	1.8 yr (8 mo–5.3 yr)	2.6 yr (16 mo–5.6 yr)	9.9 yr (4.2 yr–13.1 yr)	3.7 yr (18 mo–7.9 yr)	< 0.001
Ethnicity ^a						NS
African/North African	12 (5%)	2 (3%)	4 (7%)	4 (9%)	2 (5%)	
Asian	13 (6%)	1 (1%)	2 (3%)	4 (9%)	6 (14%)	
European	185 (81%)	69 (89%)	49 (79%)	35 (76%)	32 (74%)	
Meso/South American	4 (2%)	0 (0%)	2 (3%)	2 (4%)	0 (0%)	
Middle Eastern	3 (1%)	2 (3%)	0 (0%)	1 (2%)	0 (0%)	
Other/mixed	12 (5%)	4 (5%)	5 (8%)	0 (0%)	3 (7%)	
Number of underlying conditions						NS
None	138 (58%)	56 (68%)	33 (52%)	23 (46%)	26 (59%)	
≥ 1	102 (42%)	27 (32%)	30 (48%)	27 (54%)	18 (41%)	
Immunizations up to date ^b	178 (95%)	63 (96%)	49 (96%)	32 (89%)	34 (97%)	NS
Illness severity						
Sepsis	159 (66%)	65 (78%)	36 (57%)	25 (50%)	33 (75%)	< 0.01
PICU admission	177 (74%)	73 (88%)	40 (64%)	24 (48%)	40 (91%)	< 0.001
Need for inotropes ^c	107 (53%)	52 (71%)	12 (26%)	15 (36%)	28 (70%)	< 0.001
Days on inotropes	3 (2–5)	3 (2–4)	4 (1–6)	3 (3–7)	4 (2–5)	NS
Need for invasive ventilation ^d	106 (52%)	45 (62%)	18 (38%)	13 (30%)	30 (75%)	< 0.001
Days on invasive ventilation	4 (3–7)	5 (3–6)	3 (2–9)	4 (3–19)	4 (2–8)	NS
Need for extracorporeal membrane oxygenation ^e	4 (3%)	1 (2%)	0 (0%)	2 (9%)	1 (3%)	NS
Pediatric Risk of Mortality (21) score ^f	11 (7–16)	13 (7–16)	10 (6–17)	11 (7–19)	12 (7–15)	NS
Pediatric Index of Mortality 2 (22) score ^g (predicted death, %)	3.5% (0.8–11.6)	3.5% (0.8–13.0)	2.7% (0.8–9.3)	2.6% (0.8–4.5)	6.2% (1.0–13.0)	NS
DIC score ^h	2 (0–2)	2 (2–4)	2 (0–2)	2 (0–2)	2 (0–2)	< 0.01
Presence of DIC (DIC score ≥ 5) ^h	18 (10%)	10 (14%)	2 (4%)	3 (8%)	3 (9%)	NS

(Continued)

TABLE 3.(Continued).
Baseline Characteristics Cohort 2 by Pathogen

Variable	All Patients (n = 240)	<i>Neisseria meningitidis</i> (n = 83)	<i>Streptococcus pneumoniae</i> (n = 63)	<i>Staphylococcus aureus</i> (n = 50)	Group A <i>Streptococcus</i> (n = 44)	p
Outcome						
PICU-free days at day 28 (d) ⁱ	23 (19–25)	24 (21–25)	22 (18–26)	19 (2–25)	23 (16–25)	NS
Hospital length of stay (d)	10 (6–17)	8 (5–13)	10 (4–15)	10 (7–19)	14 (8–21)	< 0.05
Death	21 (9%)	7 (8%)	7 (11%)	4 (8%)	3 (7%)	NS

DIC = disseminated intravascular coagulation (26), NS = not significant.

^aEthnicity data were available for 229/240 patients; 78/83 *Neisseria meningitidis*, 62/63 *Streptococcus pneumoniae*, 46/50 *Staphylococcus aureus*, and 43/44 group A streptococcus (GAS) patients.

^bImmunization data were available for 188/240 patients; 66/83 *N. meningitidis*, 51/63 *S. pneumoniae*, 36/50 *S. aureus*, and 35/44 GAS patients.

^cData on inotropes were available for 202/240 patients; 73/83 *N. meningitidis*, 47/63 *S. pneumoniae*, 42/50 *S. aureus*, and 40/44 GAS patients.

^dData on invasive ventilation were available for 203/240 patients; 73/83 *N. meningitidis*, 47/63 *S. pneumoniae*, 43/50 *S. aureus*, and 40/44 GAS patients.

^eData on extracorporeal membrane oxygenation were available for 161/240 patients; 67/83 *N. meningitidis*, 37/63 *S. pneumoniae*, 22/50 *S. aureus*, and 35/44 GAS patients.

^fPediatric Risk of Mortality score (21) was available for 150/240 patients; 66/83 *N. meningitidis*, 33/63 *S. pneumoniae*, 19/50 *S. aureus*, and 32/44 GAS patients.

^gPediatric Index of Mortality 2 (22) score was available for 177/240 patients; 73/83 *N. meningitidis*, 40/63 *S. pneumoniae*, 24/50 *S. aureus*, and 40/44 GAS patients.

^hData on DIC were available for 187/240 patients; 72/83 *N. meningitidis*, 45/63 *S. pneumoniae*, 36/50 *S. aureus*, and 34/44 GAS patients.

ⁱData on PICU-free days at day 28 were available for 177/177 PICU patients; 73/73 *N. meningitidis*, 40/40 *S. pneumoniae*, 24/24 *S. aureus*, and 40/40 GAS patients.

Values are reported as counts (percentages) or medians (interquartile ranges) unless stated otherwise.

higher level than in survivors. And in cohort 2 only, ADAMTS-1 levels were correlated to PICU-free days at day 28.

DISCUSSION

This study is the first to show that ADAMTS-1 serum levels are elevated in children admitted to hospital with bacterial infection and sepsis. Importantly, our study demonstrates that in nonsurvivors ADAMTS-1 serum levels were more often detectable than in survivors, especially in patients with *N. meningitidis* disease. Additionally, ADAMTS-1 levels were correlated to PICU-free days and other markers for illness severity.

Our findings are in line with observations in experimental sepsis models where plasma levels of ADAMTS-1 in rats increased after injection with

Escherichia coli LPS (5). Furthermore, interleukin (IL)-1 β , a pro-inflammatory cytokine implicated in pediatric sepsis (27), was found to induce ADAMTS-1 production in human decidual stromal cells in vitro (28). Apart from sepsis-induced inflammation, inflammation related to nerve injury and cancer is also associated with increased ADAMTS-1 production (7, 29).

The role of ADAMTS-1 in the pathophysiology of bacterial infection, and in particular meningococcal disease, is mostly unclear so far. The association of ADAMTS-1 on sepsis mortality may be due to interference with vascular endothelial growth factor (VEGF) and VEGF receptor-2 signaling that have been involved in the pathophysiology of sepsis (30–35). ADAMTS-1 binds VEGF and blocks the VEGF receptor-2 (36), thus potentially contributing to sepsis-induced organ dysfunction (31). Furthermore, an immune-modulatory/

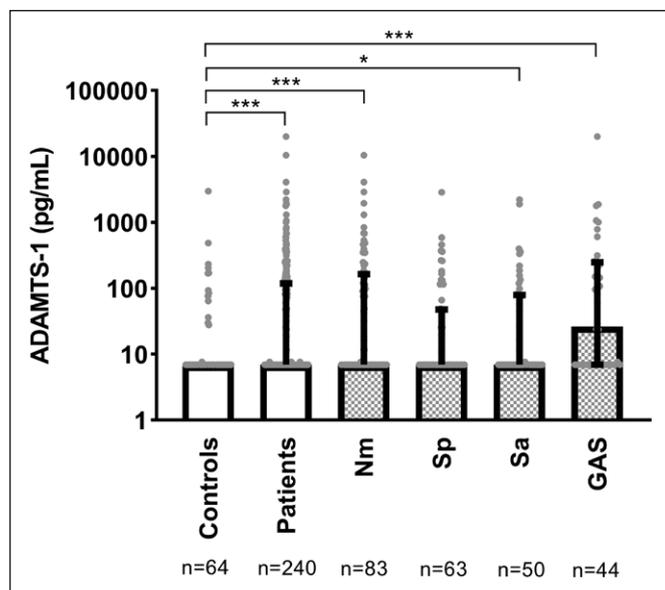


Figure 1. Cohort 2; A Disintegrin and Metalloproteinase With Thrombospondin Motifs-1 (ADAMTS-1) levels in controls and patients. Patients are further split into pathogen groups; *Neisseria meningitidis* (Nm; median, 7.0 pg/mL; interquartile range [IQR], 7.0–165 pg/mL; $p < 0.001$), *Streptococcus pneumoniae* (Sp; median, 7.0 pg/mL; IQR, 7.0–48 pg/mL; $p = 0.08$), *Staphylococcus aureus* (Sa; median, 7.0 pg/mL; IQR, 7.0–80 pg/mL; $p = 0.02$), and *group A streptococcus* (GAS; median, 26.5 pg/mL; IQR, 7.0–249; $p < 0.001$). Bar indicates median value, and whiskers indicate IQRs. Y-axis (ADAMTS-1 level) has a logarithmic scale ($*p \leq 0.05$, $***p \leq 0.001$).

suppressive role for ADAMTS-1 has also been proposed (16), possibly resulting in high levels of anti-inflammatory cytokines (e.g., IL-10, IL-1 receptor antagonist, and soluble tumor necrosis factor receptors) that are associated with sepsis mortality (37, 38). In line with these hypotheses, we found that a higher ADAMTS-1 serum level was associated with increased mortality. Although future studies should further elucidate the pathophysiological role of ADAMTS-1, our current data indicate that ADAMTS-1 can be part of the inflammatory response to pediatric sepsis.

When comparing ADAMTS-1 levels in different pathogens, our findings in nonsurvivors versus survivors were most pronounced in patients with *N. meningitidis* and partly in *Group A streptococcus* infections. Although mortality across the pathogen groups was comparable, patients with *N. meningitidis* and *group A streptococcus* infections more often had sepsis, including the need for inotropes and invasive ventilation. The systemic inflammatory response in these patients might have contributed to higher ADAMTS-1 levels. Additionally, pathogen-specific properties interfere

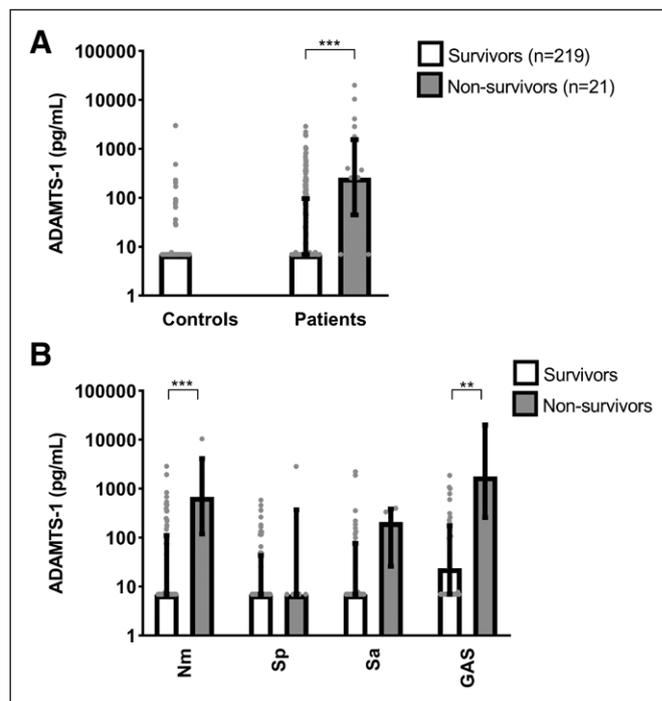


Figure 2. Cohort 2; A Disintegrin and Metalloproteinase With Thrombospondin Motifs-1 (ADAMTS-1) by mortality. **A**, ADAMTS-1 levels in survivors and nonsurvivors. **B**, ADAMTS-1 levels in survivors and nonsurvivors of invasive infections with *Neisseria meningitidis* (Nm; survivors: $n = 76$, median 7.0 pg/mL, interquartile range [IQR] 7.0–111 pg/mL; nonsurvivors: $n = 7$, median 688 pg/mL, IQR 120–4,108 pg/mL; $p < 0.001$), *Streptococcus pneumoniae* (Sp; survivors: $n = 56$, median 7.0 pg/mL, IQR 7.0–43 pg/mL; nonsurvivors: $n = 7$, median 7.0 pg/mL, IQR 7.0–371 pg/mL; $p = 0.42$), *Staphylococcus aureus* (Sa; survivors: $n = 46$, median 7.0 pg/mL, IQR 7.0–76 pg/mL; nonsurvivors: $n = 4$, median 209 pg/mL, IQR 26–384 pg/mL; $p = 0.07$), and *group A streptococcus* (GAS; survivors: $n = 41$, median 24 pg/mL, IQR 7.0–175 pg/mL; nonsurvivors: $n = 3$, median 1,793 pg/mL, IQR 257–20,052 pg/mL; $p = 0.008$). Bar indicates median value, and whiskers indicate IQRs. Y-axis (ADAMTS-1 level) has a logarithmic scale ($**p \leq 0.01$, $***p \leq 0.001$).

with the host response to infection (39, 40). *N. meningitidis* and/or *group A streptococcus* could possess properties interacting with ADAMTS-1.

A major strength of our study is that we used two independent cohorts that both revealed comparable changes in ADAMTS-1 serum level. Because the assays in both cohorts differ, we are not able to compare absolute values of ADAMTS-1. Other strengths of our study were that we examined ADAMTS-1 levels in sepsis caused by different pathogens and correlations of ADAMTS-1 with illness severity, coagulation, and inflammatory markers. Our study is possibly limited by the long-time storage of samples from cohort 1. The stability of ADAMTS-1 proteins in stored samples

is unknown. However, if samples would be affected, we assume that all samples would be affected equally. Also, we did not compare ADAMTS-1 levels measured in cohort 1 with controls. We considered convalescent samples (taken at 1 mo after PICU admission) as appropriate control for the initial measurements, but ADAMTS-1 levels after critical illness are unknown.

Comparisons between cohort 1 and cohort 2 are also limited by the variation in time from hospital onset to blood sampling. Cohort 1 collected samples at admission to PICU, at 24 hours, and at 1 month after PICU admission, while cohort 2 included all blood samples taken within 48 hours after hospital admission. Because the course of ADAMTS-1 protein levels in human sepsis is unknown, we do not know the impact of clustering of samples from cohort 2 for analysis. However, ADAMTS-1 level was not correlated to the time interval between hospital admission and the time of blood sample (data not shown).

CONCLUSIONS

Detectable ADAMTS-1 is associated with disease severity in sepsis, particularly in meningococcal sepsis, with higher ADAMTS-1 levels in nonsurvivors than in survivors. Future studies should confirm the prognostic value of ADAMTS-1 in adult sepsis and should study possible pathophysiologic mechanisms to identify potential therapeutic targets.

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REFERENCES

- Cohen J, Vincent JL, Adhikari NK, et al: Sepsis: A roadmap for future research. *Lancet Infect Dis* 2015; 15:581–614
- Boeddha NP, Schlapbach LJ, Driessen GJ, et al; EUCLIDS consortium: Mortality and morbidity in community-acquired sepsis in European pediatric intensive care units: A prospective cohort study from the European Childhood Life-threatening Infectious Disease Study (EUCLIDS). *Crit Care* 2018; 22:143
- Martinón-Torres F, Salas A, Rivero-Calle I, et al; EUCLIDS Consortium: Life-threatening infections in children in Europe (the EUCLIDS Project): A prospective cohort study. *Lancet Child Adolesc Health* 2018; 2:404–414
- Borghini L, Png E, Binder A, et al; EUCLIDS consortium: Identification of regulatory variants associated with genetic susceptibility to meningococcal disease. *Sci Rep* 2019; 9:6966
- Oveland E, Karlsen TV, Haslene-Hox H, et al: Proteomic evaluation of inflammatory proteins in rat spleen interstitial fluid and lymph during LPS-induced systemic inflammation reveals increased levels of ADAMST1. *J Proteome Res* 2012; 11:5338–5349
- Strand ME, Aronsen JM, Braathen B, et al: Shedding of syndecan-4 promotes immune cell recruitment and mitigates cardiac dysfunction after lipopolysaccharide challenge in mice. *J Mol Cell Cardiol* 2015; 88:133–144
- Kuno K, Kanada N, Nakashima E, et al: Molecular cloning of a gene encoding a new type of metalloproteinase-disintegrin family protein with thrombospondin motifs as an inflammation associated gene. *J Biol Chem* 1997; 272:556–562
- Kelwick R, Desanlis I, Wheeler GN, et al: The ADAMTS (A Disintegrin and Metalloproteinase with Thrombospondin motifs) family. *Genome Biol* 2015; 16:113
- Porter S, Clark IM, Kevorkian L, et al: The ADAMTS metalloproteinases. *Biochem J* 2005; 386:15–27
- Apte SS: A disintegrin-like and metalloprotease (reprolysin-type) with thrombospondin type 1 motif (ADAMTS) superfamily: Functions and mechanisms. *J Biol Chem* 2009; 284:31493–31497
- Mead TJ, Apte SS: ADAMTS proteins in human disorders. *Matrix Biol* 2018; 71–72:225–239
- Fujikawa K, Suzuki H, McMullen B, et al: Purification of human von Willebrand factor-cleaving protease and its identification as a new member of the metalloproteinase family. *Blood* 2001; 98:1662–1666
- Martin K, Borgel D, Lerolle N, et al: Decreased ADAMTS-13 (A disintegrin-like and metalloprotease with thrombospondin type 1 repeats) is associated with a poor prognosis in sepsis-induced organ failure. *Crit Care Med* 2007; 35:2375–2382
- Bongers TN, Emonts M, de Maat MP, et al: Reduced ADAMTS13 in children with severe meningococcal sepsis is associated with severity and outcome. *Thromb Haemost* 2010; 103:1181–1187
- Vázquez F, Hastings G, Ortega MA, et al: METH-1, a human ortholog of ADAMTS-1, and METH-2 are members of a new family of proteins with angio-inhibitory activity. *J Biol Chem* 1999; 274:23349–23357
- Rodríguez-Baena FJ, Redondo-García S, Peris-Torres C, et al: ADAMTS1 protease is required for a balanced immune cell repertoire and tumour inflammatory response. *Sci Rep* 2018; 8:13103
- Kornelisse RF, Hazelzet JA, Hop WC, et al: Meningococcal septic shock in children: Clinical and laboratory features, outcome, and development of a prognostic score. *Clin Infect Dis* 1997; 25:640–646
- de Kleijn ED, de Groot R, Hack CE, et al: Activation of protein C following infusion of protein C concentrate in children with severe meningococcal sepsis and purpura fulminans: A randomized, double-blinded, placebo-controlled, dose-finding study. *Crit Care Med* 2003; 31:1839–1847
- Derkx B, Wittes J, McCloskey R: Randomized, placebo-controlled trial of HA-1A, a human monoclonal antibody to endotoxin, in children with meningococcal septic shock. European Pediatric Meningococcal Septic Shock Trial Study Group. *Clin Infect Dis* 1999; 28:770–777
- Goldstein B, Giroir B, Randolph A; International Consensus Conference on Pediatric Sepsis: International pediatric sepsis consensus conference: Definitions for sepsis and organ dysfunction in pediatrics. *Pediatr Crit Care Med* 2005; 6:2–8
- Pollack MM, Ruttimann UE, Getson PR: Pediatric risk of mortality (PRISM) score. *Crit Care Med* 1988; 16:1110–1116
- Slater A, Shann F, Pearson G; Paediatric Index of Mortality (PIM) Study Group: PIM2: A revised version of the paediatric index of mortality. *Intensive Care Med* 2003; 29:278–285
- Couto-Alves A, Wright VJ, Perumal K, et al: A new scoring system derived from base excess and platelet count at presentation predicts mortality in paediatric meningococcal sepsis. *Crit Care* 2013; 17:R68
- Taylor FB Jr, Toh CH, Hoots WK, et al; Scientific Subcommittee on Disseminated Intravascular Coagulation (DIC) of the International Society on Thrombosis and Haemostasis (ISTH): Towards definition, clinical and laboratory criteria, and a

- scoring system for disseminated intravascular coagulation. *Thromb Haemost* 2001; 86:1327–1330
25. Hermans PW, Hibberd ML, Booy R, et al: 4G/5G promoter polymorphism in the plasminogen-activator-inhibitor-1 gene and outcome of meningococcal disease. Meningococcal research group. *Lancet* 1999; 354:556–560
 26. Khemani RG, Bart RD, Alonzo TA, et al: Disseminated intravascular coagulation score is associated with mortality for children with shock. *Intensive Care Med* 2009; 35:327–333
 27. Zhang AQ, Pan W, Gao JW, et al: Associations between interleukin-1 gene polymorphisms and sepsis risk: A meta-analysis. *BMC Med Genet* 2014; 15:8
 28. Ng YH, Zhu H, Pallen CJ, et al: Differential effects of interleukin-1beta and transforming growth factor-beta1 on the expression of the inflammation-associated protein, ADAMTS-1, in human decidual stromal cells in vitro. *Hum Reprod* 2006; 21:1990–1999
 29. Sasaki M, Seo-Kiryu S, Kato R, et al: A disintegrin and metalloprotease with thrombospondin type1 motifs (ADAMTS-1) and IL-1 receptor type 1 mRNAs are simultaneously induced in nerve injured motor neurons. *Brain Res Mol Brain Res* 2001; 89:158–163
 30. Karlsson S, Pettilä V, Tenhunen J, et al; Finnsepsis Study Group: Vascular endothelial growth factor in severe sepsis and septic shock. *Anesth Analg* 2008; 106:1820–1826
 31. Zhang RY, Liu YY, Qu HP, et al: The angiogenic factors and their soluble receptors in sepsis: Friend, foe, or both? *Crit Care* 2013; 17:446
 32. Pickkers P, Sprong T, Eijk Lv, et al: Vascular endothelial growth factor is increased during the first 48 hours of human septic shock and correlates with vascular permeability. *Shock* 2005; 24:508–512
 33. Yano K, Liaw PC, Mullington JM, et al: Vascular endothelial growth factor is an important determinant of sepsis morbidity and mortality. *J Exp Med* 2006; 203:1447–1458
 34. van der Flier M, van Leeuwen HJ, van Kessel KP, et al: Plasma vascular endothelial growth factor in severe sepsis. *Shock* 2005; 23:35–38
 35. van der Flier M, Baerveldt EM, Miedema A, et al: Decreased expression of serum and microvascular vascular endothelial growth factor receptor-2 in meningococcal sepsis*. *Pediatr Crit Care Med* 2013; 14:682–685
 36. Luque A, Carpizo DR, Iruela-Arispe ML: ADAMTS1/METH1 inhibits endothelial cell proliferation by direct binding and sequestration of VEGF165. *J Biol Chem* 2003; 278:23656–23665
 37. Osuchowski MF, Welch K, Siddiqui J, et al: Circulating cytokine/inhibitor profiles reshape the understanding of the SIRS/CARS continuum in sepsis and predict mortality. *J Immunol* 2006; 177:1967–1974
 38. Gogos CA, Drosou E, Bassaris HP, et al: Pro- versus anti-inflammatory cytokine profile in patients with severe sepsis: A marker for prognosis and future therapeutic options. *J Infect Dis* 2000; 181:176–180
 39. Gasparini R, Panatto D, Bragazzi NL, et al: How the knowledge of interactions between meningococcus and the human immune system has been used to prepare effective Neisseria meningitidis vaccines. *J Immunol Res* 2015; 2015:189153
 40. Loof TG, Deicke C, Medina E: The role of coagulation/fibrinolysis during *Streptococcus pyogenes* infection. *Front Cell Infect Microbiol* 2014; 4:128