

Opinion

The Induced Resistance Lexicon:
Do's and Don'ts

Jonas De Kesel,¹ Uwe Conrath,² Víctor Flors,³ Estrella Luna,⁴ Melissa H. Mageroy,⁵ Brigitte Mauch-Mani,⁶ Victoria Pastor,³ María J. Pozo,⁷ Corné M.J. Pieterse,⁸ Jurriaan Ton,⁹ and Tina Kynedt^{1,*}

To be protected from biological threats, plants have evolved an immune system comprising constitutive and inducible defenses. For example, upon perception of certain stimuli, plants can develop a conditioned state of enhanced defensive capacity against upcoming pathogens and pests, resulting in a phenotype called 'induced resistance' (IR). To tackle the confusing lexicon currently used in the IR field, we propose a widely applicable code of practice concerning the terminology and description of IR phenotypes using two main phenotypical aspects: local versus systemic resistance, and direct versus primed defense responses. Our general framework aims to improve uniformity and consistency in future scientific communication, which should help to avoid further misinterpretations and facilitate the accessibility and impact of this research field.

A History of Terminology Confusion

'Induced resistance' (IR; see [Glossary](#)) is a phenotype in which plants, once stimulated by certain pathogens, pests, beneficial microbes, chemical agents, physical wounding, or herbivory, exhibit lowered susceptibility for future **challenges** when compared with naïve control plants [1]. Apart from the observation of lowered susceptibility to pests and diseases, IR phenotypes can be confirmed at the level of defense responses, for instance, by augmented production of reactive oxygen species, enforced callose deposition, and altered epigenomes, transcriptomes, proteomes, metabolomes, etc.. These effects are similar to effects stimulated by pathogen-associated molecular patterns (PAMPs), damage-associated molecular patterns (DAMPs), or pathogen effectors.

Since the first recognition of IR phenotypes in the early 1900s [2–5], several terms and concepts have been introduced to describe IR. Terms introduced in the early days, such as 'plant immunization' [2], were continued to be used by some [6], while others introduced a variety of new terms. Sequeira [7] was among the first who raised the issue of a confusing terminology in this field. Certain terms were (re-)defined at the First International Symposium on Induced Resistance to Plant Diseases in 2000 [8]. Nevertheless, in 2006, Tuzun [9] published a manuscript entitled 'Terminology related to induced systemic resistance: Incorrect use of synonyms may lead to a scientific dilemma by misleading interpretation of results', illustrating that scientific communication concerning IR was still hampered by an inadequate terminology. More than a decade later, scientific research has led to a better molecular understanding of IR phenotypes and their underlying mechanisms. Unfortunately, this has not been accompanied by a uniform terminology and/or consistently used conceptualizations. Here, by tackling three points of confusion, we aim to disentangle the main concepts within the IR research field and to clarify the IR terminology in the light of recent findings.

Point 1 – A Clarification on Some IR Terms

Ross [10] introduced the term '**systemic acquired resistance**' (SAR) to refer to the reduced susceptibility to viruses in leaves of tobacco (*Nicotiana tabacum*) plants as a result of previous

Highlights

Upon perception of certain stimuli, plants can develop a conditioned state of enhanced defensive capacity against upcoming pathogens and pests, resulting in a phenotype called 'induced resistance' (IR).

Scientific communication in the IR research domain is flawed with inconsistent use of various conceptualizations and terms.

Researchers working on non-model organisms and/or less-studied plant tissues – which often make use of distinct natural defense mechanisms – are struggling to choose the correct term for their observations.

Different biological and chemical IR stimuli tend to induce resistance through various pathways and hence terminology can and should not be linked to underlying mechanisms.

¹Department of Biotechnology, Faculty of Bioscience Engineering, Ghent University, 9000 Ghent, Belgium

²Department of Plant Physiology, Plant Biochemistry and Molecular Biology, RWTH Aachen University, 52056 Aachen, Germany

³Metabolic Integration and Cell Signaling Laboratory, Plant Physiology Section, Unidad Asociada al Consejo Superior de Investigaciones Científicas (EEZ-CSIC), Department of Ciencias Agrarias y del Medio Natural, Universitat Jaume I, 12071 Castellón, Spain

⁴School of Biosciences, University of Birmingham, Edgbaston, Birmingham B15 2TT, UK

⁵Department of Molecular Plant Biology, Norwegian Institute of Bioeconomy Research, 1433 Ås, Norway



viral infections in distant leaves. However, in later studies by Ross and colleagues, several other terms were used to describe the same phenotype: ‘systemic resistance’ [11], ‘systemic induced resistance’ [12], and ‘induced resistance’ [13]. Nowadays, the term SAR is still being used and its definition generally encompasses the following elements: typically induced by a local inoculation with a necrotizing pathogen, predominantly mediated by the phytohormone salicylic acid (SA) and probably by *N*-hydroxypipecolic acid as well, and often associated with at least some accumulation of pathogenesis-related (PR) proteins [14,15]. In the 1990s, non-pathogenic plant growth-promoting rhizobacteria and fungi (PGPR and PGPF, respectively) were found to trigger similar IR phenotypes [16–19]. Pieterse *et al.* [20] demonstrated in the model plant *Arabidopsis thaliana* that this happened independently of SA and PR genes, but rather was based on the jasmonate (JA) and ethylene (ET) pathways. The term ‘**induced systemic resistance**’ (ISR) was adopted to differentiate this IR phenomenon from SAR [21], although the two types of IR were considered as phenotypically similar [22]. Indeed, although the underlying defense mechanisms and spectrum of effectiveness may differ for specific SAR and ISR phenotypes, the observable defensive effects are often similar.

Over time, a dichotomy seems to be introduced with respect to the use of the terms SAR and ISR. Based on their predominant hormonal regulators in *Arabidopsis* and the microorganisms initially found to induce these phenotypes, SAR and ISR are now often considered as fundamentally different IR phenotypes, despite earlier agreements for these terms to be used synonymously [8]. At the same time, ISR is also used as a general term to refer to systemic forms of IR, as initially agreed on [8]. Moreover, chemicals, non-proteinaceous amino acids, physical injury, volatile organic compounds, etc. have been found to elicit IR as well [1,23] but as non-biotic agents or actions, these stimuli do not seem to fit in the strict definitions of ISR and SAR.

To encompass all known IR phenomena in a uniform terminology, we encourage the scientific community to use ‘induced resistance’ (IR) as an umbrella term and ‘**IR stimulus**’ as general reference for the evoking agent or action. ISR should be used when there is convincing evidence that upon local contact with an IR stimulus, endogenous signals are spread systemically to stimulate defense-related processes in essentially all plant parts. In the absence of such evidence, or when the stimulus itself is systemically distributed in the plant – as shown for exogenously applied synthetic β -aminobutyric acid (BABA) [24,25], and as currently being studied for migrating endophytes (V. Pastor, unpublished) – the more general term IR is recommended. Thus, although this is often done in contemporary literature, the term ISR should not be limited only to IR phenotypes triggered by PGPR or PGPF, while phytohormones other than JA and ET can be mediators of ISR. Because the term SAR has strictly been defined over the years (*vide supra*), we propose that these specific ISR phenotypes can still be referred to as ‘SAR phenotypes’.

We make a plea to consider two main aspects of IR phenotypes when selecting an adequate terminology. The first is the predominant occurrence of **local resistance** or **systemic resistance** upon IR establishment. Correspondingly, this trait forms the first axis in our general framework to characterize IR phenotypes (Figure 1, Key Figure). Importantly, a strong local resistance does not exclude systemic effects. Indeed, as shown on the x-axis of Figure 1, we consider nearly all known IR phenotypes to be an outcome of both local and systemic resistance. Nevertheless, only when biologically relevant resistance is observed consistently throughout the entire plant, the terms ISR or SAR are appropriate.

Point 2 – IR: The Sum of Direct and Primed Defense Responses

Over the past decades, scientific progress revealed that the establishment of IR does not always depend on, and sometimes even is not associated with, a strong **direct defense response**

⁶Laboratoire de Biologie Moléculaire et Cellulaire, Institute of Biology, Université de Neuchâtel, CH-2000 Neuchâtel, Switzerland

⁷Department of Soil Microbiology and Symbiotic Systems, Estación Experimental del Zaidín (CSIC), 18008 Granada, Spain

⁸Science for Life, Plant-Microbe Interactions Group, Department of Biology, Utrecht University, 3584 CH Utrecht, The Netherlands

⁹Department of Animal and Plant Sciences, Plant Production and Protection Centre, The University of Sheffield, Sheffield S10 2TN, UK

*Correspondence:
Tina.Kyndt@UGent.Be (T. Kyndt).

Key Figure

A General Framework to Characterize Induced Resistance (IR) Phenotypes in Terms of Local/Systemic Resistance (x-Axis) and Direct/Primed Defense Responses (y-Axis)

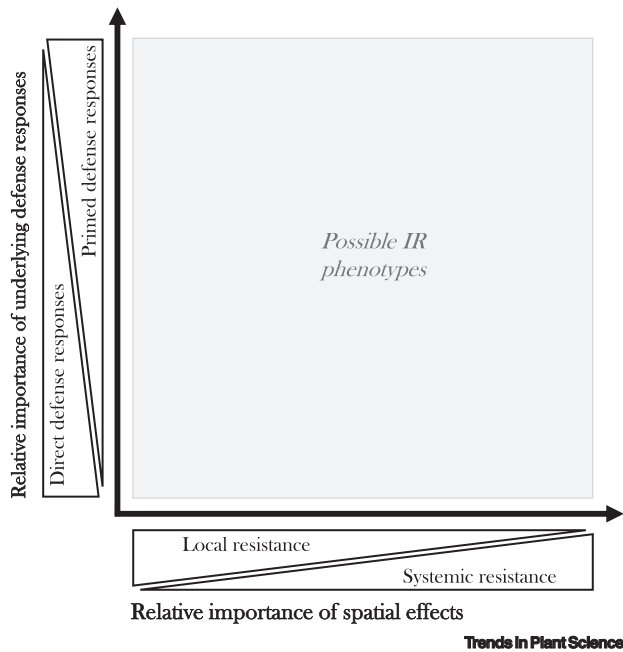


Figure 1. All IR phenotypes can be considered to be the result of both directly induced defense responses and primed defense responses, as well as of local resistance and systemic resistance. Importantly, the ratios of importance for these parameters can vary depending on many parameters such as the IR stimulus, its concentration or intensity, time point of analysis, plant under study, age of the plant, plant tissue under study, pathogen under study, analyzed read-out (see Figure 2).

upon application of an IR stimulus [1,26,27]. While some genes, enzymes, or pathways are not affected directly upon treatment with an IR stimulus, they can get activated earlier, stronger, and/or faster upon later challenges when compared with non-IR plants. Such an enhanced capacity to mobilize infection-induced cellular defense responses is referred to as '(defense) priming' [26]. Important to notice that **primed defense responses** do rely on various mechanisms that are activated directly upon IR stimulation, such as the accumulation of dormant signaling proteins, transcription factors and hormones, epigenetic alterations, and/or increased levels of receptors and coreceptors [1,14,28,29]. Nevertheless, the main outcome of defense priming is a boosted defense response that is only activated upon a later challenge. The relative contribution of direct and primed defense responses is the second main characteristic that we deem essential to describe IR phenotypes. Correspondingly, this facet forms the y-axis in the framework illustrated in Figure 1.

For a while, the primed defense responses were considered as the most relevant aspects of IR phenotypes [14,30], resulting in IR definitions being somehow adapted in various works [27,31–36]. Today it is clear that the IR phenotype is associated with both direct induction of

Glossary

Challenge: inoculation with a pathogen or pest after an IR phenotype has been established by a certain IR stimulus. If on purpose, it is typically to investigate the resulting level of resistance or the molecular effects on the affected defense response.

Direct defense responses: defense responses that are induced, locally or systemically, upon contact with the IR stimulus. Hence, these responses can be detected prior to any challenge.

Induced resistance (IR): reduced disease susceptibility of a plant in response to stimulation by a pathogen, insect herbivore or wounding, beneficial microbe, or chemical agent. IR is often effective against a broad spectrum of pests, pathogens, and sometimes even abiotic stresses.

Induced systemic resistance (ISR): type of IR that leads to resistance in plant tissues distant from those that were initially in contact with the stimulating biological or chemical agent. Should not be limited to IR phenotypes that are systemically triggered by plant growth-promoting rhizobacteria/fungi and are associated with JA/ET.

IR stimulus: any agent or action that leads to the establishment of an IR phenotype in the affected plant.

Local resistance: resistance observed in the plant tissue that was initially in contact to the IR stimulus.

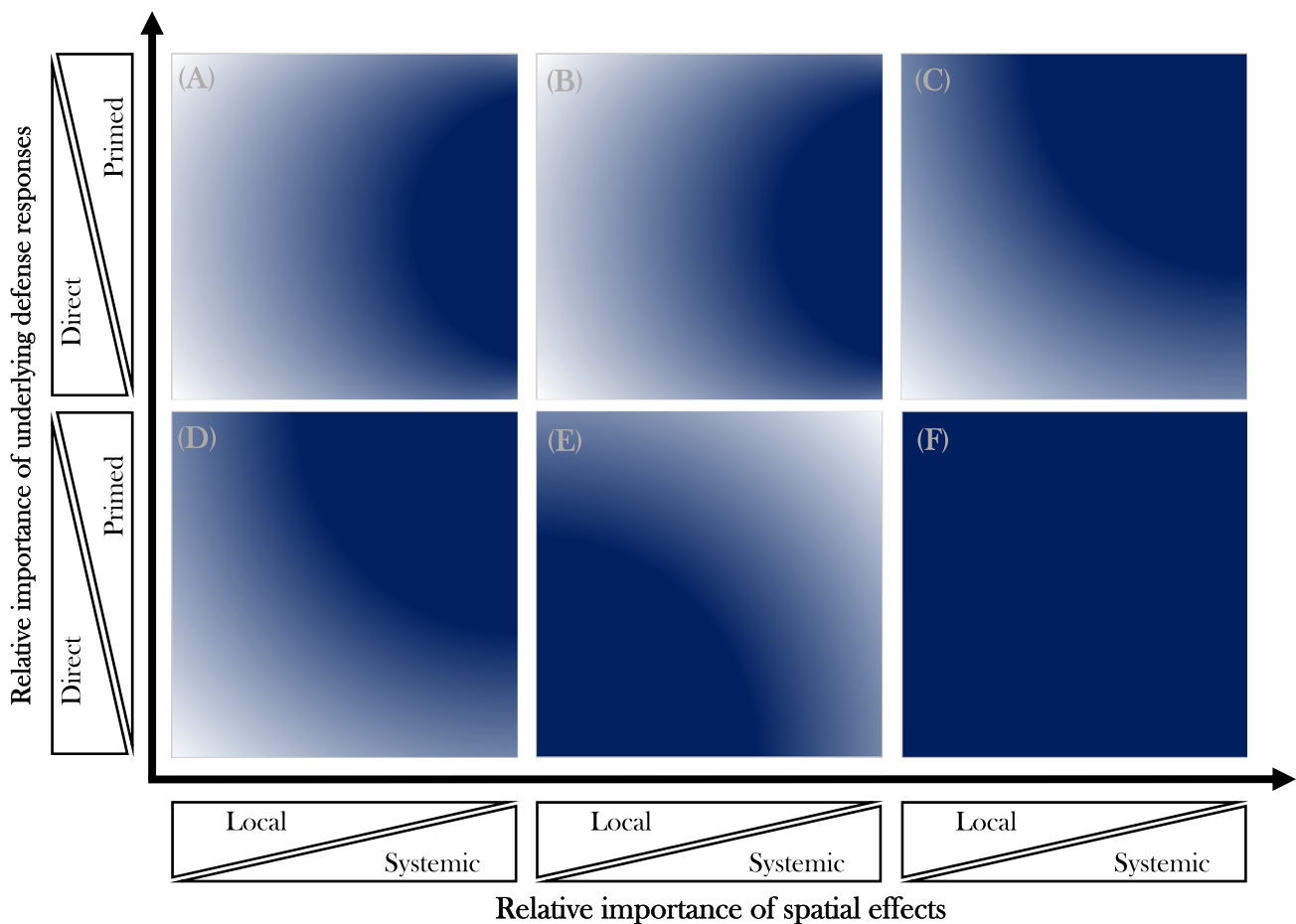
Primed defense responses: defense responses that, because of defense priming, are activated earlier, stronger, and/or faster in IR plants upon subsequent challenge with a pathogen, pest, or insect herbivory, but not in the absence of an attacker. Although these defense response modulations are only observable upon later challenges, defense priming is associated with alterations which take place directly upon IR stimulation, and which make the plant to be primed for enhanced defense (e.g., epigenetic alterations, increased receptor presence, accumulation of dormant proteins and/or hormones, and other, currently unidentified, effects).

Systemic acquired resistance

(SAR): specific type of ISR that classically, but not necessarily (i) leads to resistance in systemic tissues upon local infection by a necrotizing pathogen, (ii) is associated with at least some *PR* gene expression/*PR* protein accumulation, and (iii) is controlled mainly by the action

defense responses – which can be transient or long-lasting – and primed defense responses [1,29,37]. Nevertheless, some stimuli mainly work through direct activation of plant defense genes or metabolites, while others seem to work predominantly via defense priming. Undoubtedly, many other features are also relevant for the description and characterization of IR phenotypes, such as durability, plant species or cultivar, or mode-of-action. However, with respect to the selection of a proper terminology, the axes shown in Figure 1 are considered as the most relevant. As an illustration of how to use and interpret this graphical representation, some well-studied IR phenotypes have been characterized via this framework (Figure 2).

of SA and, probably, by *N*-hydroxyphenylacetic acid.
Systemic resistance: resistance observed in plant parts distant to those that were initially in contact with the IR stimulus, or even in the entire plant.



Trends in Plant Science

Figure 2. An Illustration of a Set of Well-studied Induced Resistance (IR) Phenotypes Using the Framework Presented in Figure 1. Color shading represents the relative importance of local versus systemic resistance, and direct versus primed defense responses for the indicated types of IR. (A) Pathogen-induced systemic acquired resistance (SAR) [15]: this phenotype is characterized – by definition – by a systemic form of IR and includes both direct and primed defense responses. (B) Ascorbate oxidase (AO)-induced systemic resistance (ISR) in rice (*Oryza sativa*) against *Meloidogyne graminicola* [62]: both primed and direct defense responses have been associated with this IR phenotype. Although panels (A) and (B) have a similar shading, AO-ISR is fundamentally different from SAR, as the stimulus is not a necrotizing pathogen, and the phenotype does not depend on salicylic acid but rather on jasmonic acid and ethylene [62]; (C) *Pseudomonas simiae* WCS417-ISR in arabidopsis [63]: for this IR phenotype nearly all observed defense responses have been shown to be primed and systemic; (D) and (E) β -aminobutyric acid (BABA)-IR upon application of low and high doses, respectively [27]: low BABA doses lead to systemically primed defense responses, while high BABA doses lead to directly activated responses mainly in the treated plant parts. These two panels clearly illustrate that for one specific IR stimulus, the underlying mechanisms can be different; (F) methyl-jasmonate (MeJA) IR [64–70]: via direct activation of defense responses [64–69] and via priming [68–70], MeJA has been described to stimulate plant resistance both systemically [64,66] and locally [64,65,69].

Point 3 – IR: What Is in a Name? And What Is Not?

To refer to chemicals that trigger IR, a plethora of terms has been introduced, mostly not accompanied by a clear definition, characterization, or thorough differentiation from others: ‘resistance activator’ [38], ‘plant (defense) activator’ [39–41], ‘synthetic inducer of defense responses’ [42], ‘defense elicitor’ [43], ‘inducer of plant immunity’ [44], ‘(plant) resistance inducer’ [45,46], ‘disease resistance compound’ [47], ‘elicitor’ [48,49], ‘inducer’ [50], ‘SAR inducer’ [21], ‘plant strengthener’ [51,52], ‘priming-inducing chemical’ [31], ‘priming agent’ [53], etc. As these terms seem to be tacitly associated with specific underlying mechanisms – ‘priming agents’ may be thought of as mainly leading to primed defense responses, which can be conceived as opposed to the mode-of-actions of ‘plant defense activators’ – the concurrent use of these terms can be confusing to newcomers in the field and non-experts.

Apart from being confusing, a presumed association of these terms with specific underpinning effects can be considered as incorrect as well. Even for well-studied IR stimuli, the underlying mechanisms can differ depending on multiple parameters. Indeed, how IR is manifested relies on a multidependent and only partially characterized network [54,55]. Whether or not a specific IR phenotype is associated with direct and/or primed defense responses depends not only on the stimulus, but also on multiple experimental, environmental, and spatiotemporal parameters, as well as on the read-out in question (Box 1). Indeed, it has been shown that for BABA, the direct/and or primed activation of certain underlying mechanisms depends on the plant species under study [55], the applied BABA concentration [27], and the necrotrophic or biotrophic lifestyle of the pathogen that is battled [56,57], while indole-3-carboxylic acid (I3CA) has been shown to work differently depending on the age of the plant [58]. Here we propose to use the neutral term ‘IR stimulus’ to refer to any action or agent that induces IR.

Correspondingly, we recommend the use of a general ‘stimulus-phenotype’ terminology (e.g., ‘BABA-IR’, ‘PGPR-ISR’, ‘chemical X-IR’, ‘*Pseudomonas syringae*-SAR’) that should not – and cannot – be associated with any underlying mechanism. Indeed, ‘BABA-IR’ or ‘I3CA-IR’ should merely be used to refer to a phenotype of enhanced defensive capacity, triggered upon treatment with BABA or I3CA. By not referring to underlying mechanisms, the hereby-presented terminology can be easily and correctly applicable for studies executed on non-model organisms or on less-studied tissues, which often employ distinct natural defense mechanisms. For the sake of clarity, the aforementioned terminology can optionally be extended by the pathogen for which an increased resistance is observed, as well as the plant host (e.g., ‘BABA-IR against *Botrytis cinerea* in *Solanum lycopersicum*’ or ‘*Pseudomonas syringae*-SAR against *Hyaloperonospora arabidopsidis* in *Arabidopsis*’). However, adding a pathogen for which an increased resistance is observed is no necessity, as we consider IR

Box 1. Fitness Costs Associated with IR

An important consequence of the assumptions and conceptualizations described in this work lies in the evaluation of the cost-efficiency of IR stimuli. In some reports, predominant induction of direct plant defenses has been linked to evident fitness costs [27], whereas IR phenotypes mainly based on priming have been shown to be associated with lower fitness costs [27,59]. However, this is no ever-valid correlation as there are noticeable exceptions. For instance, the chemical compound diproline was identified as an IR stimulus based on direct induction of defense marker genes in rice (*Oryza sativa*) [60]. Nevertheless, diproline-IR was not associated with any obvious negative effects on rice growth or yield, even when plants were repetitively treated during their lifespan [60]. Indeed, a variety of parameters can influence the relative importance of primed and direct defense responses. Moreover, whether an intense and long-lasting induction of defense pathways, a mainly primed defense response, or an intermediate form is optimal depends on the cost–benefit balance in a given environment [29]. The model in Figure 1 in the main text is consistent with this notion: a specific localization on the y-axis in Figure 1 cannot be interpreted in terms of long-term physiological effects and/or allocation costs. Hereto, IR phenotypes should undergo relevant ecological assessments, as underlined by Martínez-Medina *et al.* [61].

also a proper term to refer to phenotypes in which defense mechanisms are enhanced, regardless of whether effective resistance against a specific pathogen has been experimentally confirmed.

Concluding Remarks

By discussing some potential pitfalls within the IR lexicon and clarifying a consensus point-of-view concerning the current terminology and conceptualizations, we hope to stimulate a more clear, consistent, and unambiguous scientific communication in this field. The aforementioned suggestions, in combination with the general framework presented in [Figure 1](#), may contribute hereto as they can help to characterize and describe scientific observations in a more uniform manner. Although we encourage authors to use the hereby-presented terminology and conceptualization, we realize that because of novel findings (see [Outstanding Questions](#)) or alternative term usage in related scientific fields, specific terms or concepts might remain to be/become used differently. That is why, in general, we strongly encourage a well-evaluated terminology, provided with a thorough elaboration on the intended meaning for the various terms and concepts being used. We believe that our propositions can make the already existing IR vocabulary transparent and easily applicable for contemporary research, in which an expanding range of IR stimuli, plants, tissues, pathogens, and diseases is being studied. As novel introductions in this field can be facilitated by a consistent and widely supported lexicon, we are convinced that the general notion and appreciation for IR will expand, ultimately extending the reach and impact of the reported observations for plant sciences, agriculture, and beyond.

Declaration of Interests

The authors have no interests to declare.

References

- Mauch-Mani, B. *et al.* (2017) Defense priming: an adaptive part of induced resistance. *Annu. Rev. Plant Biol.* 68, 485–512
- Beauverie, J. (1901) Essais d'immunisation des végétaux contre les maladies cryptogamiques. *CR Acad. Sci. Ser. III* 133, 107–110
- Ray, J. (1901) Les maladies cryptogamiques des végétaux. *Rev. Gen. Bot.* 13, 145–151
- Arnaud, C. (1933) On the vaccination of the tobacco plant against *Thielaviopsis basicola*. *Bull. Torrey Bot. Club.* 60, 583–597
- Chester, K.S. (1933) The problem of acquired physiological immunity in plants. *Q. Rev. Biol.* 8, 275–324
- Kuč, J. (1982) Plant immunization-mechanisms and practical implications. In *Active Defense Mechanisms in Plants* (Wood, R.K.S., ed.), pp. 157–178, Springer
- Sequeira, L. (1983) Mechanisms of induced resistance in plants. *Annu. Rev. Microbiol.* 37, 51–79
- Hammerschmidt, R. *et al.* (2000) Inducing resistance: a summary of papers presented at the First International Symposium on Induced Resistance to Plant Diseases, Corfu, May 2000. *Eur. J. Plant Pathol.* 107, 1–6
- Tuzun, S. (2006) Terminology related to induced systemic resistance: incorrect use of synonyms may lead to a scientific dilemma by misleading interpretation of results. In *Multigenic and Induced Systemic Resistance in Plants* (Tuzun, S. and Bent, E., eds), pp. 1–8, Springer
- Ross, F.A. (1961) Systemic acquired resistance induced by localized virus infections in plants. *Virology* 14, 340–358
- Bozarth, R.F. and Ross, A.F. (1964) Systemic resistance induced by localized virus infections: extent of changes in uninfected plant parts. *Virology* 24, 446–455
- Simons, T.J. and Ross, A.F. (1971) Metabolic changes associated with systemic induced resistance. *Pathology* 61, 293–300
- Pritchard, D.W. and Ross, A.F. (1975) The relationship of ethylene to formation of tobacco mosaic virus lesions in hypersensitive responding tobacco leaves with and without induced resistance. *Virology* 64, 295–307
- Conrath, U. *et al.* (2015) Priming for enhanced defense. *Annu. Rev. Phytopathol.* 53, 97–119
- Hartmann, M. and Zeier, J. (2019) *N*-hydroxy-pipecolic acid and salicylic acid: a metabolic duo for systemic acquired resistance. *Curr. Opin. Plant Biol.* 50, 44–57
- Alström, S. (1991) Induction of disease resistance in common bean susceptible to halo blight bacterial pathogen after seed bacterization with rhizosphere pseudomonads. *J. Gen. Appl. Microbiol.* 37, 495–501
- Van Peer, R. *et al.* (1991) Induced resistance and phytoalexin accumulation in biological control of fusarium wilt of carnation by *Pseudomonas* sp. strain WCS 417 r. *Phytopathology* 81, 728–734
- Wei, G. *et al.* (1991) Induction of systemic resistance of cucumber to *Colletotrichum orbiculare* by select strains of plant growth-promoting rhizobacteria. *Phytopathology* 81, 1508–1512
- Meera, M.S. *et al.* (1994) Plant growth promoting fungi from zoysiagrass rhizosphere as potential inducers of systemic resistance in cucumbers. *Phytopathology* 84, 1399–1406
- Pieterse, C.M.J. *et al.* (1996) Systemic resistance in *Arabidopsis* induced by biocontrol bacteria is independent of salicylic acid accumulation and pathogenesis-related gene expression. *Plant Cell* 8, 1225–1237
- Sticher, L. *et al.* (1997) Systemic acquired resistance. *Annu. Rev. Phytopathol.* 35, 235–270
- van Loon, L.C. *et al.* (1998) Systemic resistance induced by rhizosphere bacteria. *Annu. Rev. Phytopathol.* 36, 453–483
- Conrath, U. (2009) Priming of induced plant defense responses. *Adv. Bot. Res.* 51, 361–395
- Cohen, Y. and Gisi, U. (1994) Systemic translocation of 14C-DL-3-aminobutyric acid in tomato plants in relation to induced resistance against *Phytophthora infestans*. *Physiol. Mol. Plant Pathol.* 45, 441–456
- Balmer, A. *et al.* (2019) Accumulation patterns of endogenous β -aminobutyric acid during plant development and defence in *Arabidopsis thaliana*. *Plant Biol.* 21, 318–325

Outstanding Questions

How durable is IR in plants within and over generations?

Will IR be a sustainable and effective strategy for crop protection in future agriculture?

How can IR be integrated within other pest and disease management strategies?

What is the role of epigenetics in the regulation of IR?

How did IR develop in the evolutionary context of the plant immune system?

Are IR phenotypes that act through different molecular mechanisms (e.g., established by different stimuli) additive?

26. Conrath, U. *et al.* (2002) Priming in plant–pathogen interactions. *Trends Plant Sci.* 7, 210–216
27. van Hulten, M. *et al.* (2006) Costs and benefits of priming for defense in *Arabidopsis*. *Proc. Natl. Acad. Sci.* 103, 5602–5607
28. Pastor, V. *et al.* (2014) Preparing to fight back: generation and storage of priming compounds. *Front. Plant Sci.* 5, 295
29. Wilkinson, S.W. *et al.* (2019) Surviving in a hostile world: plant strategies to resist pests and diseases. *Annu. Rev. Phytopathol.* 57, 505–529
30. van der Ent, S. *et al.* (2009) Induced resistance - orchestrating defence mechanisms through crosstalk and priming. In *Annual Plant Reviews, Volume 34: Molecular Aspects of Plant Disease Resistance* (Parker, J., ed.), pp. 334–370, Blackwell
31. Pastor, V. *et al.* (2013) Primed plants do not forget. *Environ. Exp. Bot.* 94, 46–56
32. Conrath, U. *et al.* (2006) Priming: getting ready for battle. *Mol. Plant-Microbe Interact.* 19, 1062–1071
33. van Loon, L.C. (2000) Systemic induced resistance. In *Mechanisms of Resistance to Plant Diseases* (Slusarenko, A.J. *et al.*, eds), pp. 521–574, Springer
34. Vallad, G.E. and Goodman, R.M. (2004) Systemic acquired resistance and induced systemic resistance in conventional agriculture. *Crop Sci.* 44, 1920–1934
35. Beckers, G.J.M. and Conrath, U. (2007) Priming for stress resistance: from the lab to the field. *Curr. Opin. Plant Biol.* 10, 425–431
36. Goellner, K. and Conrath, U. (2008) Priming: it's all the world to induced disease resistance. *Eur. J. Plant Pathol.* 121, 233–242
37. Balmer, A. *et al.* (2015) The 'prime-ome': towards a holistic approach to priming. *Trends Plant Sci.* 20, 443–452
38. Walters, R.D. (2013) Controlling crop diseases using induced resistance: challenges for the future. *J. Exp. Bot.* 64, 1263–1280
39. Du, Q. *et al.* (2012) Novel benzo-1, 2, 3-thiadiazole-7-carboxylate derivatives as plant activators and the development of their agricultural applications. *J. Agric. Food Chem.* 60, 346–353
40. Huang, C.-H. *et al.* (2012) Effect of application frequency and reduced rates of acibenzolar-s-methyl on the field efficacy of induced resistance against bacterial spot on tomato. *Plant Dis.* 96, 221–227
41. Wise, M.L. (2013) Plant defense activators: application and prospects in cereal crops. In *50 Years of Phytochemistry Research* (Gang, D.R., ed.), pp. 55–70, Springer
42. Durner, J. and Klessig, D.F. (1995) Inhibition of ascorbate peroxidase by salicylic acid and 2, 6-dichloroisonicotinic acid, two inducers of plant defense responses. *Proc. Natl. Acad. Sci.* 92, 11312–11316
43. Bruce, T.J.A. (2012) Variation in plant responsiveness to defense elicitors caused by genotype and environment. *Front. Plant Sci.* 5, 349
44. Reimer-Michalski, E.-M. and Conrath, U. (2016) Innate immune memory in plants. *Semin. Immunol.* 28, 319–327
45. Myresiotis, C.K. *et al.* (2012) Evaluation of plant growth-promoting rhizobacteria, acibenzolar-S-methyl and hymexazol for integrated control of *Fusarium* crown and root rot on tomato. *Pest Manag. Sci.* 68, 404–411
46. Sandroni, M. *et al.* (2020) Plant resistance inducers (PRIs): perspectives for future disease management in the field. *CAB Rev.* 15, 1–10
47. Jiang, H.-Y. *et al.* (2018) Transcript profiling and gene identification involved in the ethylene signal transduction pathways of creeping bentgrass (*Agrostis stolonifera*) during ISR response induced by butanediol. *Molecules* 23, 706
48. Porat, R. *et al.* (2002) Effects of various elicitors on the transcription of a β -1,3-endoglucanase gene in citrus fruit. *J. Phytopathol.* 150, 70–75
49. Ren, Y. *et al.* (2012) Postharvest BTH treatment induced disease resistance and enhanced reactive oxygen species metabolism in muskmelon (*Cucumis melo* L.) fruit. *Eur. Food Res. Technol.* 234, 963–971
50. Barilli, E. *et al.* (2010) Benzothiadiazole and BABA improve resistance to *Uromyces pisi* (Pers.) Wint. in *Pisum sativum* L. with an enhancement of enzymatic activities and total phenolic content. *Eur. J. Plant Pathol.* 128, 483–493
51. Newton, A. and Pons-Kühnemann, J. (2007) Induced resistance in natural ecosystems and pathogen population biology: exploiting interactions. In *Induced Resistance for Plant Defence: A Sustainable Approach to Crop Protection* (Walters, D. *et al.*, eds), pp. 133–142, Blackwell Publishing
52. Sobhy, I.S. *et al.* (2012) Less is more: treatment with BTH and laminarin reduces herbivore-induced volatile emissions in maize but increases parasitoid attraction. *J. Chem. Ecol.* 38, 348–360
53. Savvides, A. *et al.* (2016) Chemical priming of plants against multiple abiotic stresses: mission possible? *Trends Plant Sci.* 21, 329–340
54. Mathys, J. *et al.* (2012) Genome-wide characterization of ISR induced in *Arabidopsis thaliana* by *Trichoderma hamatum* t382 against *Botrytis cinerea* infection. *Front. Plant Sci.* 3, 108
55. Cohen, Y. *et al.* (2016) Baba-induced resistance: milestones along a 55-year journey. *Phytoparasitica* 44, 513–538
56. Ton, J. and Mauch-Mani, B. (2004) β -Amino-butyric acid-induced resistance against necrotrophic pathogens is based on aba-dependent priming for callose. *Plant J.* 38, 119–130
57. Ji, H. *et al.* (2015) β -Aminobutyric acid-induced resistance against root-knot nematodes in rice is based on increased basal defense. *Mol. Plant-Microbe Interact.* 28, 519–533
58. Mateu-Garcia, D. *et al.* (2018) The age-dependent priming. *IPOBC-WPRS Bulletin* 135, 65–66
59. Buswell, W. *et al.* (2018) Chemical priming of immunity without costs to plant growth. *New Phytol.* 218, 1205–1216
60. De Kesel, J. *et al.* (2020) The use of PTI-marker genes to identify novel compounds that establish induced resistance in rice. *Int. J. Mol. Sci.* 21, 317
61. Martínez-Medina, A. *et al.* (2016) Recognizing plant defense priming. *Trends Plant Sci.* 21, 818–822
62. Singh, R.R. *et al.* (2020) Ascorbate oxidation activates systemic defence against root-knot nematode *Meloidogyne graminicola* in rice. *J. Exp. Bot.* 71, 4271–4284
63. Pieterse, C.M.J. *et al.* (2020) *Pseudomonas simiae* WCS417: star track of a model beneficial rhizobacterium. *Plant Soil* 1–19
64. Repka, V. *et al.* (2004) Methyl jasmonate is a potent elicitor of multiple defense responses in grapevine leaves and cell-suspension cultures. *Biol. Plantarum* 48, 273–283
65. Kępczyńska, E. and Król, P. (2012) The phytohormone methyl jasmonate as an activator of induced resistance against the necrotroph *Alternaria porri* f. sp. *solani* in tomato plants. *J. Plant Interact.* 7, 307–315
66. Nahar, K. *et al.* (2011) The jasmonate pathway is a key player in systemically induced defense against root knot nematodes in rice. *Plant Physiol.* 157, 305–316
67. Benevenuto, R.F. *et al.* (2019) Transcriptional profiling of methyl jasmonate-induced defense responses in bilberry (*Vaccinium myrtillus* L.). *BMC Plant Biol.* 19, 70
68. Mageroy, M.H. *et al.* (2020) Molecular underpinnings of methyl jasmonate-induced resistance in Norway spruce. *Plant Cell Environ.* 43, 1827–1843
69. Valenzuela-Riffo, F. *et al.* (2020) Priming of defense systems and upregulation of MYC2 and JAZ1 genes after *Botrytis cinerea* inoculation in methyl jasmonate-treated strawberry fruits. *Plants* 4, 447
70. Singh, U.B. *et al.* (2019) *Trichoderma harzianum*-and methyl jasmonate-induced resistance to *Bipolaris sorokiniana* through enhanced phenylpropanoid activities in bread wheat (*Triticum aestivum* L.). *Front. Microbiol.* 10, 1697