

Antibody Responses to *Helicobacter pylori* and Risk of Developing Colorectal Cancer in a European Cohort



Julia Butt¹, Mazda Jenab², Michael Pawlita¹, Anne Tjønneland³, Cecilie Kyrø³, Marie-Christine Boutron-Ruault^{4,5}, Franck Carbonnel^{4,6}, Catherine Dong⁴, Rudolf Kaaks⁷, Tilman Kühn⁷, Heiner Boeing⁸, Matthias B. Schulze^{9,10}, Antonia Trichopoulou¹¹, Anna Karakatsani^{11,12}, Carlo La Vecchia^{11,13}, Domenico Palli¹⁴, Claudia Agnoli¹⁵, Rosario Tumino¹⁶, Carlotta Sacerdote¹⁷, Salvatore Panico¹⁸, Bas Bueno-de-Mesquita^{19,20,21}, Roel Vermeulen²², Inger T. Gram²³, Elisabete Weiderpass², Kristin Benjaminsen Borch²³, Jose Ramón Quirós²⁴, Antonio Agudo²⁵, Miguel Rodríguez-Barranco^{26,27,28}, Carmen Santiuste^{26,29}, Eva Ardanaz^{26,30,31}, Bethany Van Guelpen^{32,33}, Sophia Harlid³², Liher Imaz^{34,35}, Aurora Perez-Cornago³⁶, Marc J. Gunter², Semi Zouïouich², Jin Young Park², Elio Riboli³⁷, Amanda J. Cross³⁷, Alicia K. Heath³⁷, Tim Waterboer¹, and David J. Hughes³⁸

ABSTRACT

Background: While *Helicobacter pylori* (*H. pylori*) is the major cause of gastric cancer, it has also been suggested to be involved in colorectal cancer development. However, prospective studies addressing *H. pylori* and colorectal cancer are sparse and inconclusive. We assessed the association of antibody responses to *H. pylori* proteins with colorectal cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC) cohort.

Methods: We applied *H. pylori* multiplex serology to measure antibody responses to 13 *H. pylori* proteins in prediagnostic serum samples from 485 colorectal cancer cases and 485 matched controls nested within the EPIC study. Odds ratios (OR) and 95% confidence intervals (CI) were calculated using multivariable conditional logistic regression to estimate the association of *H. pylori* overall and protein-specific seropositivity with odds of developing colorectal cancer.

Results: Fifty-one percent of colorectal cancer cases were *H. pylori* seropositive compared with 44% of controls, resulting in an OR of 1.36 (95% CI, 1.00–1.85). Among the 13 individual *H. pylori* proteins, the association was driven mostly by seropositivity to *Helicobacter* cysteine-rich protein C (HcpC; OR: 1.66; 95% CI, 1.19–2.30) and Vacuolating cytotoxin A (VacA) (OR: 1.34; 95% CI, 0.99–1.82), the latter being nonstatistically significant only in the fully adjusted model.

Conclusions: In this prospective multicenter European study, antibody responses to *H. pylori* proteins, specifically HcpC and VacA, were associated with an increased risk of developing colorectal cancer.

Impact: Biological mechanisms for a potential causal role of *H. pylori* in colorectal carcinogenesis need to be elucidated, and subsequently whether *H. pylori* eradication may decrease colorectal cancer incidence.

¹Infections and Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany. ²International Agency for Research on Cancer (IARC-WHO), Lyon, France. ³Diet, Genes and Environment, Danish Cancer Society Research Center, Copenhagen, Denmark. ⁴CESP, Faculté de Médecine, Université Paris-Sud and Faculté de Médecine, UVSQ, INSERM, Université Paris-Saclay, Villejuif, France. ⁵Gustave Roussy, Villejuif, France. ⁶Department of Gastroenterology, Bicêtre University Hospital, Assistance Publique des Hôpitaux de Paris, Le Kremlin-Bicêtre, France. ⁷Division of Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany. ⁸Department of Epidemiology, German Institute of Human Nutrition Potsdam, Rehbruecke, Nuthetal, Germany. ⁹Department of Molecular Epidemiology, German Institute of Human Nutrition Potsdam, Rehbruecke, Nuthetal, Germany. ¹⁰Institute of Nutrition Science, University of Potsdam, Nuthetal, Germany. ¹¹Hellenic Health Foundation, Athens, Greece. ¹²2nd Pulmonary Medicine Department, School of Medicine, National and Kapodistrian University of Athens, "ATTIKON" University Hospital, Haidari, Greece. ¹³Department of Clinical Sciences and Community Health Università degli Studi di Milano, Milan, Italy. ¹⁴Cancer Risk Factors and Life-Style Epidemiology Unit, Institute for Cancer Research, Prevention and Clinical Network-ISPPO, Florence, Italy. ¹⁵Epidemiology and Prevention Unit, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy. ¹⁶Cancer Registry and Histopathology Department, "Civic - M.P. Arezzo" Hospital, ASP Ragusa, Italy. ¹⁷Unit of Cancer Epidemiology, Città della Salute e della Scienza University-Hospital and Center for Cancer Prevention (CPO), Turin, Italy. ¹⁸Dipartimento di Medicina Clinica e Chirurgia, Federico II University, Naples, Italy. ¹⁹Department of Gastroenterology and Hepatology, University

Medical Centre, Utrecht, the Netherlands. ²⁰Department of Epidemiology and Biostatistics, The School of Public Health, Imperial College London, London, United Kingdom. ²¹Department of Social & Preventive Medicine, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia. ²²Julius Center for Health Sciences and Primary Care, Cancer Epidemiology, University Medical Center Utrecht, the Netherlands. ²³Faculty of Health Sciences, Department of Community Medicine, University of Tromsø, The Arctic University of Norway, Tromsø, Norway. ²⁴Public Health Directorate, Asturias, Spain. ²⁵Unit of Nutrition and Cancer, Cancer Epidemiology Research Program, Catalan Institute of Oncology, Barcelona, Spain. ²⁶CIBER of Epidemiology and Public Health (CIBERESP), Madrid, Spain. ²⁷Andalusian School of Public Health (EASP), Granada, Spain. ²⁸Instituto de Investigación Biosanitaria de Granada (ibs.GRANADA), Universidad de Granada, Granada, Spain. ²⁹Department of Epidemiology, Murcia Regional Health Council, IMIB-Arrixaca, Murcia, Spain. ³⁰Navarra Public Health Institute, Pamplona, Spain. ³¹IdiSNA, Navarra Institute for Health Research, Pamplona, Spain. ³²Department of Radiation Sciences, Oncology, Umeå University, Umeå, Sweden. ³³Wallenberg Centre for Molecular Medicine, Umeå University, Umeå, Sweden. ³⁴Ministry of Health of the Basque Government, Public Health Division of Gipuzkoa, Donostia-San Sebastian, Spain. ³⁵Biodonostia Health Research Institute, Donostia-San Sebastian, Spain. ³⁶Cancer Epidemiology Unit, Nuffield Department of Population Health, University of Oxford, Oxford, United Kingdom. ³⁷Department of Epidemiology and Biostatistics, School of Public Health, Imperial College London, London, United Kingdom. ³⁸Cancer Biology and Therapeutics Group, School of Biomolecular and Biomedical Science, UCD Conway Institute, University College Dublin, Dublin, Ireland.

Butt et al.

Introduction

Helicobacter pylori (*H. pylori*) is a well-established cause of non-cardia gastric cancer and has been classified as a group 1 carcinogen by the International Agency for Research on Cancer (IARC; ref. 1). In addition to gastric cancer, *H. pylori* infection has been investigated for an etiologic role in other cancers of the digestive tract, including colorectal cancer (2).

The most recent meta-analysis of 27 published case-control studies reported a pooled odds ratio (OR) of 1.27 [95% confidence interval (CI), 1.17–1.37] for the association of *H. pylori* with colorectal cancer (3). However, only a few prospective studies have assessed the risk of developing colorectal cancer with *H. pylori* seropositivity, which moreover reported inconclusive results: three studies conducted in Germany, Finland, and a Caucasian population in the United States showed null findings (4–6), whereas two other U.S. studies reported a positive association with seropositivity to specific *H. pylori* proteins, including Vacuolating cytotoxin A (VacA), Chaperonin GroEl, *Helicobacter* cysteine-rich protein C (HcpC), and hypothetical protein HP1564 (7, 8).

While a causality and potential biological mechanism for the observed association has not been verified yet, a study by Hu and colleagues further supports the hypothesis of an involvement of *H. pylori* in colorectal cancer development (9). These investigators found that the individuals either successfully eradicated for *H. pylori* infection or uninfected were at significantly lower risk of developing adenomas than those with persistent *H. pylori* infection (9).

To comprehensively assess the association of *H. pylori* infection with risk of developing colorectal cancer, we analyzed antibody responses to 13 *H. pylori* proteins in a case-control study nested within a large European multicenter prospective cohort. We hypothesized that in addition to overall *H. pylori* seropositivity, protein-specific seropositivity, including positivity to *H. pylori* virulence factors and toxins, would be specifically associated with colorectal cancer development.

Materials and Methods

Study population, case ascertainment, and control selection

This colorectal cancer case-control study is nested within the European Prospective Investigation into Nutrition and Cancer (EPIC) cohort study investigating the association between diet, lifestyle, and environmental factors and cancer incidence. The rationale and methods of the EPIC design have been published previously (10). Briefly, more than 520,000 individuals, aged 35 to 70 years, were enrolled from 10 Western European countries between 1992 and 2000. This analysis is based on participant data from 7 of these countries (France, Italy, Spain, United Kingdom, The Netherlands, Greece, and Germany). Dietary and lifestyle data, as well as serum samples, were collected at baseline, with standardized blood collection and processing protocols across the study centers. Serum samples were stored at the International Agency for Research on Cancer (IARC, Lyon, France) at -196°C and shipped on dry ice to the German Cancer Research

Center (DKFZ, Heidelberg, Germany). The study has been approved by the IARC Ethics Committee and the ethics committees of all local EPIC centers.

The nested case-control study included prediagnostic serum samples from 492 colorectal cancer cases (primary tumors coded C18-C20 according to the 10th Revision of the International Statistical Classification of Diseases, Injury and Causes of Death). The median time between blood draw and diagnosis was 3.4 years with a range of 0.4 to 8.5 years. Controls were selected by incidence density sampling from all cohort members alive and free of cancer at the time of matching and matched to cases 1:1 by age at blood collection (± 6 months to ± 2 years), sex, study center, time of the day at blood collection (± 2 to 4 hours interval), fasting status at blood collection ($< 3/3-6$ hours); among women by menopausal status, and among premenopausal women by phase of menstrual cycle and hormone therapy use at time of blood collection. Seven case-control pairs were excluded from the analysis due to technical errors during multiplex serology measurement, resulting in a final sample set of 485 colorectal cancer cases and 485 controls for analysis.

H. pylori multiplex serology

Serum samples were analyzed for antibodies against *H. pylori* proteins in a 1:1,000 dilution using multiplex serology. The methodology has been described in detail elsewhere (11, 12). Briefly, *H. pylori* proteins (Supplementary Table S1) were recombinantly expressed as GST-X-tag fusion proteins and affinity purified on glutathione-casein coated beads, where each bead type has a different internal fluorescent color (SeroMap; Luminex Corp., Austin, TX) to identify the loaded antigen. A mixture of bead sets carrying different antigens was incubated with prediluted serum. A Luminex xMAP analyzer (Luminex Corp.) was used to identify the bead type and simultaneously quantify the amount of bound serum antibody by biotinylated goat anti-human IgA/IgM/IgG antibody and a fluorescent reporter conjugate Streptavidin-R-Phycoerythrin. The level of antibody response is given as median fluorescence intensity (MFI) of at least 100 beads per type measured. Background values against the N-terminal GST, the C-terminal tag, and the bead-surface were subtracted to generate net MFI values.

Antigen-specific cutoffs (Supplementary Table S1) were defined as described previously at the approximate inflection point of frequency distribution curves (8). Overall seropositivity to *H. pylori* was assigned to individuals seropositive for at least 4 of the 13 *H. pylori* proteins included in the multiplex serology panel, as this algorithm had previously been shown to offer the best specificity and sensitivity when the assay was validated against a commercially available ELISA (12).

Statistical analysis

Wilcoxon Mann-Whitney test was applied to compare continuous antibody responses (MFI) to *H. pylori* proteins between controls and cases.

Conditional logistic regression models were applied to compute ORs, and the corresponding 95% CIs, for the association of antibody

Note: Supplementary data for this article are available at Cancer Epidemiology, Biomarkers & Prevention Online (<http://cebp.aacrjournals.org/>).

J. Butt, M. Jenab, T. Waterboer, and D.J. Hughes contributed equally to this article.

Current address for B. Bueno-de-Mesquita: Department for Determinants of Chronic Diseases (DCD), National Institute for Public Health and the Environment (RIVM), Bilthoven, the Netherlands.

Corresponding Author: David J. Hughes, School of Biomolecular and Biomedical Science, Conway Institute, Dublin D04 V1W8, Ireland. Phone: 353-1-716-6988; E-mail: david.hughes@ucd.ie

Cancer Epidemiol Biomarkers Prev 2020;29:1475-81

doi: 10.1158/1055-9965.EPI-19-1545

©2020 American Association for Cancer Research.

responses to *H. pylori* overall seropositivity and seropositivity to individual *H. pylori* proteins with colorectal cancer development. To increase specificity, study participants were only considered *H. pylori* antigen positive to any single antigen when simultaneously classified as overall *H. pylori* seropositive.

The following variables were considered as potential confounders and therefore included in the model for adjustment: body mass index (BMI; <25, 25–29.9, ≥30 kg/m²), smoking status (never, former, current), alcohol consumption (<6, 6–20, ≥20 g/day), and highest education attained at baseline (≤primary school, technical/professional, ≥secondary school; model 1). We also performed an exploratory assessment of the impact of dietary variables and physical activity on the association. Using backward elimination only total daily energy intake (kcal), as well as daily intake of fish (g), and dairy products (g) contributed significantly to the model in our population and were included in a second model in addition to the abovementioned potential confounders (model 2). We furthermore performed a stratified analysis by country since *H. pylori* seroprevalence widely varied geographically (Supplementary Table S2).

All statistical analyses were performed with SAS version 9.4 (SAS Institute, Cary, NC). A *P* value below 0.05 was considered significant.

Data sharing

For information on how to submit an application for gaining access to EPIC data and/or biospecimens, please follow the instructions at <http://epic.iarc.fr/access/index.php>.

Results

Study characteristics and risk factors for *H. pylori* seropositivity

Cases and controls did not differ in any of the baseline socio-demographic characteristics (Table 1). When assessing dietary variables, cases had a higher daily energy intake as well as lower intake of dairy products than controls at baseline (Table 1). Seropositivity to *H. pylori* at baseline was more frequent in individuals with higher BMI (≥30), lower education (≤primary school), and lower physical activity index (≤moderately inactive; Supplementary Table S2). Furthermore, baseline *H. pylori* seroprevalence varied by country with highest rates in Spain (73%) and Italy (54%), followed by Germany (42%), and lowest values in the Netherlands (30%) and the United Kingdom (27%). Sample sizes from France and Greece (each contributing 11 controls) were too small to be considered for comparison (Supplementary Table S2). Regarding dietary variables, *H. pylori* seropositive individuals exhibited a lower daily intake of dairy products at baseline (Supplementary Table S2).

Association of antibody responses to *H. pylori* proteins with colorectal cancer development

We first assessed whether incident colorectal cancer cases and controls differed in their level of antibody response, given by continuous MFI, to *H. pylori* proteins (Fig. 1). MFI level tended to be higher among colorectal cancer cases than among controls to all the antigens, except for NapA. Continuous MFI to GroEl, HcpC, and VacA were significantly higher among colorectal cancer cases than controls (Fig. 1).

Applying cutoffs for seropositivity, overall 51% of cases were *H. pylori* seropositive (positive to ≥4 out of 13 *H. pylori* proteins) as opposed to 44% of controls, which resulted in significantly increased odds of developing colorectal cancer (OR: 1.41; 95% CI, 1.06–1.89; *P* =

Table 1. Baseline characteristics of the colorectal cancer case-control study nested within the EPIC study.

	Total (n = 970)	Controls (n = 485)	Cases (n = 485)
Sex, n (%)			
Female	494 (51)	247 (51)	247 (51)
Male	476 (49)	238 (49)	238 (49)
Age at blood draw (years), mean (SD)	60 (8)	60 (8)	60 (8)
Country, n (%)			
France	22 (2)	11 (2)	11 (2)
Italy	202 (21)	101 (21)	101 (21)
Spain	164 (17)	82 (17)	82 (17)
United Kingdom	268 (28)	134 (28)	134 (28)
The Netherlands	140 (14)	70 (14)	70 (14)
Greece	22 (2)	11 (2)	11 (2)
Germany	152 (16)	76 (16)	76 (16)
Education, n (%)			
≤Primary school	427 (46)	212 (45)	215 (46)
Technical/professional	210 (23)	115 (25)	95 (21)
≥Secondary school	295 (32)	142 (30)	153 (33)
Missing	38	16	22
BMI (kg/m ²), n (%)			
<25	327 (34)	167 (34)	160 (33)
25–29.9	458 (47)	238 (49)	220 (45)
≥30	185 (19)	80 (16)	105 (22)
Smoking status, n (%)			
Never	436 (42)	234 (48)	202 (42)
Former	337 (35)	154 (32)	183 (38)
Current	191 (20)	95 (20)	96 (20)
Missing	6	2	4
Alcohol intake (g/day), n (%)			
<6	442 (46)	229 (47)	213 (44)
6–20	254 (26)	127 (26)	127 (26)
≥20	273 (28)	129 (27)	144 (30)
Missing	1	0	1
Physical activity ^a , n (%)			
Inactive	326 (34)	158 (33)	168 (35)
Moderately inactive	296 (31)	138 (29)	158 (33)
Moderately active	178 (19)	92 (19)	86 (18)
Active	162 (17)	93 (19)	69 (14)
Missing	8	4	4
Dietary variables, intake/day, mean (SD)			
Total energy (kcal)	2,111 (729)	2,062 (615)	2,160 (826)
Total vegetables (g)	198 (128)	200 (123)	196 (132)
Total fruits (g)	258 (192)	255 (181)	261 (203)
Dairy (g)	324 (238)	336 (229)	312 (246)
Cereals (g)	222 (133)	226 (145)	219 (120)
Fish (g)	33 (33)	34 (34)	32 (32)
Red meats (g)	44 (34)	42 (29)	45 (39)
Processed meats (g)	32 (49)	30 (28)	35 (64)

Abbreviations: g, grams; kcal, kilocalories.

^aCambridge physical activity index.

0.02). The association remained significant after adjustment for potential sociodemographic confounders (OR: 1.43; 95% CI, 1.06–1.93; *P* = 0.02) and dietary variables (OR: 1.36; 95% CI, 1.00–1.85; *P* = 0.05; Table 2). For individual *H. pylori* proteins, seropositivity to HcpC (28% of cases vs. 20% of controls) and VacA (36% of cases vs. 30% of controls) was significantly associated with increased odds of developing colorectal cancer in the model adjusting for sociodemographic variables (OR: 1.66; 95% CI, 1.19–2.30; *P* < 0.01 and OR: 1.38; 95% CI, 1.02–1.86; *P* = 0.04, respectively; Table 2). Adjustment for dietary

Butt et al.

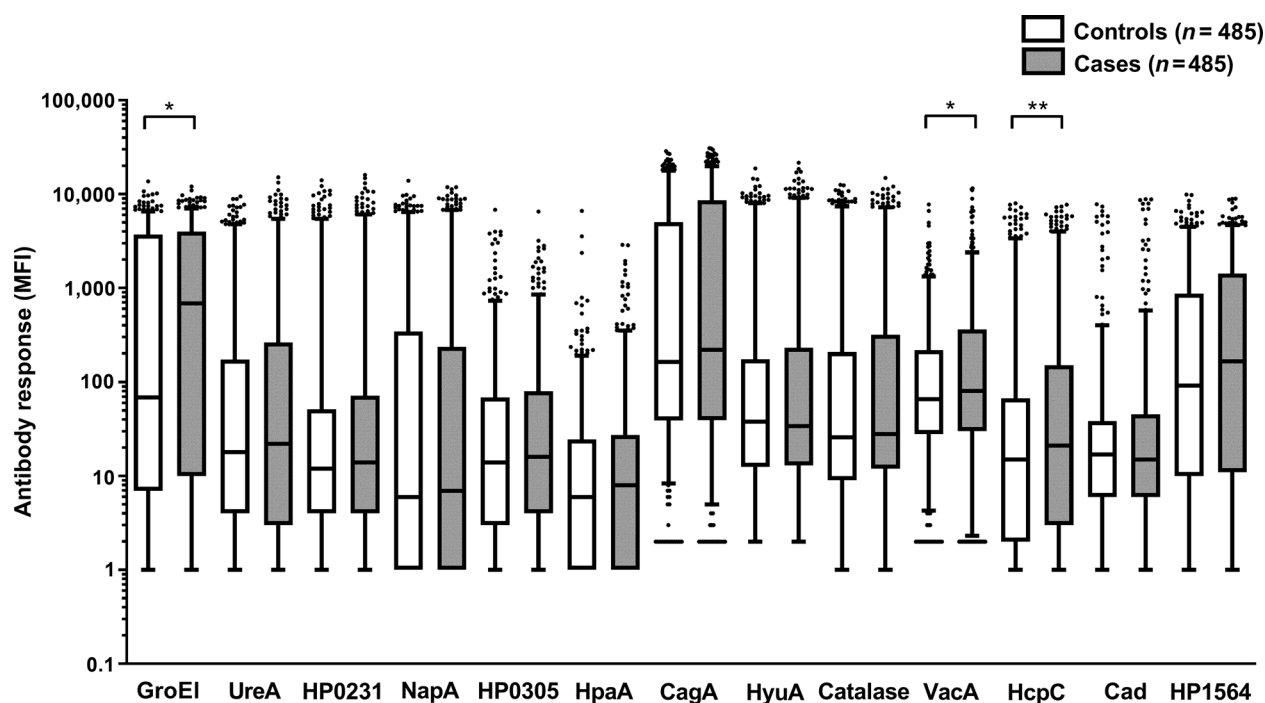


Figure 1.

Antibody responses (MFIs) to *H. pylori* proteins in cases and controls; the EPIC study. Boxes represent 25th to 75th and whiskers the 5th to 95th percentile, solid lines show the median. Dots represent data points lying outside the 5th and 95th percentiles, respectively. Wilcoxon Mann-Whitney test was applied to compare continuous antibody responses (MFI) between controls and cases (*, $P < 0.05$; **, $P < 0.01$).

variables did not change the OR substantially; however, the association of colorectal cancer with VacA seropositivity lost significance (OR: 1.34; 95% CI, 0.99–1.82; $P = 0.06$), while the association with HcpC remained significant (OR: 1.66; 95% CI, 1.19–2.31; $P < 0.01$; Table 2).

Odds for colorectal cancer were increased with seropositivity to most of the other 11 proteins, although these results were not significant, with OR point estimates ranging from 1.39 (HpaA) to 0.93 (NapA; Table 2).

Table 2. Antibody responses to *H. pylori* proteins and odds of developing colorectal cancer; the EPIC study.

Antigen	Positive n (%)		Crude model		Multivariable model 1		Multivariable model 2	
	Controls n = 485	Cases n = 485	OR (95% CI) ^b	P ^b	OR (95% CI) ^c	P ^c	OR (95% CI) ^d	P ^d
<i>H. pylori</i> overall+	214 (44)	247 (51)	1.41 (1.06–1.89)	0.02	1.43 (1.06–1.93)	0.02	1.36 (1.00–1.85)	0.05
HcpC ⁺ a	95 (20)	134 (28)	1.65 (1.20–2.27)	<0.01	1.66 (1.19–2.30)	<0.01	1.66 (1.19–2.31)	<0.01
VacA ⁺ a	145 (30)	174 (36)	1.37 (1.02–1.83)	0.04	1.38 (1.02–1.86)	0.04	1.34 (0.99–1.82)	0.06
HpaA ⁺ a	36 (7)	48 (10)	1.40 (0.88–2.24)	0.16	1.46 (0.90–2.36)	0.13	1.39 (0.85–2.26)	0.19
Catalase ⁺ a	112 (23)	134 (28)	1.31 (0.96–1.78)	0.09	1.32 (0.97–1.82)	0.08	1.24 (0.90–1.72)	0.19
GroEI ⁺ a	208 (43)	231 (48)	1.30 (0.96–1.74)	0.09	1.29 (0.95–1.75)	0.11	1.23 (0.90–1.68)	0.20
CagA ⁺ a	133 (27)	152 (31)	1.24 (0.92–1.68)	0.15	1.28 (0.94–1.74)	0.12	1.24 (0.91–1.70)	0.18
HP1564 ⁺ a	177 (36)	201 (41)	1.28 (0.96–1.69)	0.09	1.27 (0.95–1.69)	0.11	1.26 (0.94–1.69)	0.13
UreA ⁺ a	102 (21)	122 (25)	1.29 (0.94–1.76)	0.12	1.27 (0.92–1.75)	0.14	1.23 (0.89–1.70)	0.22
HP0231 ⁺ a	93 (19)	102 (21)	1.16 (0.84–1.60)	0.37	1.18 (0.85–1.65)	0.32	1.17 (0.83–1.63)	0.37
HP0305 ⁺ a	94 (19)	106 (22)	1.16 (0.85–1.59)	0.34	1.17 (0.85–1.61)	0.35	1.12 (0.80–1.56)	0.50
Cad ⁺ a	67 (14)	77 (16)	1.18 (0.83–1.67)	0.37	1.15 (0.80–1.65)	0.46	1.17 (0.81–1.70)	0.41
HyuA ⁺ a	116 (24)	119 (25)	1.04 (0.76–1.42)	0.81	1.02 (0.74–1.41)	0.89	1.03 (0.74–1.43)	0.86
NapA ⁺ a	128 (26)	127 (26)	0.99 (0.74–1.32)	0.94	0.97 (0.72–1.31)	0.84	0.93 (0.68–1.26)	0.63

Note: Significant associations ($P < 0.05$) are marked in bold font.

^aParticipants were considered antigen-positive only when simultaneously overall *H. pylori* seropositive (i.e., ≥ 4 out of 13 antigens positive).

^bCrude model: Conditional logistic regression model based on the matching factors.

^cMultivariable model 1: crude model plus additional adjustment for BMI (<25 , 25–29.9, ≥ 30 kg/m²), smoking status (never, former, current), alcohol consumption (<6 , 6–20, ≥ 20 g/day), and highest education attained at baseline (\leq primary school, technical/professional, \geq secondary school).

^dMultivariable model 2: multivariable model 1 plus additional adjustment for daily intake levels of dairy products [g], fish [g], and total energy [kcal] (all as continuous variables).

H. pylori seroprevalence is known to vary by European region. In our data, subgroup analyses by country did not differ significantly for *H. pylori* overall or to the two most strongly associated antigens (VacA and HcpC; Supplementary Fig. S1).

Discussion

In this case-control study nested within a large prospective European cohort, we observed that seropositivity to at least 4 out of the 13 assessed *H. pylori* proteins was associated with a significant 36% increased odds of developing colorectal cancer, after adjustments for potential confounding. Among these 13 individual *H. pylori* proteins, higher antibody levels to HcpC and VacA showed the strongest associations with colorectal cancer development.

These results concur with previous findings from two independent populations in the United States, which found significant associations with seropositivity to *H. pylori* HcpC and VacA using the same serologic method (7, 8). However, significant findings in the latter study of diverse populations in the United States were predominantly found with African American colorectal cancer cases ($n = 399$) and not the Caucasian American cases ($n = 3,067$). Two recent reports found an increased risk of developing advanced colorectal neoplasia among individuals with persistent *H. pylori* infection compared with noninfected individuals and/or individuals with successful *H. pylori* eradication (9, 13). In line with this hypothesis, previous studies showed a higher risk of having colorectal adenomas for individuals exhibiting *H. pylori*-related chronic atrophic gastritis or more severe gastric conditions (14, 15). We recently reported that antibody responses to HcpC and VacA are among the four antigens of our 13-plex serology panel that best indicate an active *H. pylori* infection (16). The toxin VacA is an important *H. pylori* virulence factor, which damages host cells by causing vacuolation (17), while the function of HcpC is unknown. Moreover, concordant with the previous U.S. study (8), we here showed that not only seroprevalence but also level of antibody response was higher among incident colorectal cancer cases than controls. These findings suggest that colorectal cancer risk may be increased when *H. pylori* infection is more severe. The severity of *H. pylori* infection is thought to be dependent on the virulence of the bacterial strain, the individual host response to the bacterium, and other environmental factors, including diet (18), potentially explaining the observed difference in association among distinct populations.

The mechanism by which *H. pylori* might contribute to colon carcinogenesis remains unclear. A direct effect would require the presence of *H. pylori* or its toxins in the colon. Presence of *H. pylori* in the feces or tumor tissue of patients with colorectal cancer is rarely reported (19–21). Thus, it needs to be elucidated whether the bacterium is present in and induces a deleterious effect on the gut epithelium. Another possibility is that *H. pylori* could exert indirect effects promoting colonic carcinogenesis, for example, by (i) creating a systemic inflammatory environment beyond the stomach (22), (ii) through inducing gut microbiome dysbiosis that in turn may exert a carcinogenic effect (23), or (iii) by inducing the production of effector molecules in the host like gastrin that could have mitogenic effects in the colon (24–29). These hypotheses, however, need mechanistic studies to show a causal relationship between *H. pylori* infection and colorectal cancer development. The potential involvement of *H. pylori* infection in colorectal cancer etiology may depend on other cofactors and its role may be stronger in populations where infection is more prevalent. We did not

observe significant heterogeneity by region or country, which could be a consequence of our limited number of cases and thus a larger study would be important.

Our study has several limitations and strengths. As mentioned above, we applied a serologic assay to determine antibody responses to *H. pylori*. This is an indirect and systemic measure and does not provide information on whether *H. pylori* may also locally infect the colorectal tissue. Furthermore, we cannot distinguish between current and past infection, also due to the lack of information on antibiotic treatment history. However, referring to the study by Hu and colleagues (9), this would lead to a bias towards the null hypothesis, because the authors of this study reported that only persistent infections are associated with an increased risk of colorectal adenoma. Another limitation is the potential of residual confounding, which cannot completely be ruled out, although most EPIC data has been validated. A strength of the applied multiplex serology assay is its ability to analyze several antigens per infectious agent, allowing assessment of a more detailed host immune response to the bacterium beyond just overall seropositivity. Because of the assessment of multiple antigens, it may be argued that correction of our findings for multiple comparisons is required. We contend against this for two reasons. First, the number of antigens tested was modest and, second, our study was based on a clear a priori hypothesis derived from previous findings for the same antigens (8). Nevertheless, even when applying the conservative Bonferroni correction for the 13 *H. pylori* proteins included (P for significance = 0.004), the association of seropositivity to HcpC with colorectal cancer risk would still retain statistical significance ($P = 0.003$). Multiplex serology furthermore allows quantification of antibody response level to the respective antigens. As reported in a previous observational study, higher antibody levels in addition to seropositivity were associated with increased odds of developing colorectal cancer (8), potentially referring to the severity of *H. pylori* infection and/or strength of the host's response to the infection. A major strength of the study design is that it is based on a large multicenter cohort covering most of Western Europe with detailed prospective data collection and serum samples taken several years prior to diagnosis allowing for a comprehensive adjustment for potential confounders including dietary variables. However, information on medication use, including antibiotics and nonsteroidal anti-inflammatory drugs, was not included in the baseline questionnaire of EPIC and could therefore not be assessed as potential confounders in the current analysis.

In conclusion, antibody responses to *H. pylori*, specifically to proteins HcpC and VacA, in prediagnostic serum samples were significantly associated with increased risk of developing colorectal cancer in the EPIC study. Our observations need verification in different populations, particularly in those with high *H. pylori* prevalence. Furthermore, it remains to be elucidated whether *H. pylori* infection is causally related to colorectal carcinogenesis and what the underlying biological mechanisms are. Direct pathogenic evidence would then advance *H. pylori* eradication or control as a potential beneficial measure for colorectal cancer prevention.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Disclaimer

Where authors are identified as personnel of the International Agency for Research on Cancer/World Health Organization, the authors alone are responsible for the views expressed in this article and they do not necessarily represent the decisions,

Butt et al.

policy or views of the International Agency for Research on Cancer/World Health Organization. The funding sources had no influence on the design of the study; the collection, analysis, and interpretation of data; the writing of the report; or the decision to submit the paper for publication.

Authors' Contributions

Conception and design: J. Butt, M. Jenab, M. Pawlita, E. Weiderpass, D.J. Hughes
Development of methodology: J. Butt, M. Jenab, M. Pawlita, T. Waterboer, D.J. Hughes

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): J. Butt, M. Jenab, M. Pawlita, A. Tjønneland, M.-C. Boutron-Ruault, F. Carbonnel, R. Kaaks, T. Kühn, H. Boeing, M.B. Schulze, A. Trichopoulos, A. Karakatsani, D. Palli, R. Tumino, C. Sacerdote, S. Panico, B. Bueno-de-Mesquita, R. Vermeulen, E. Weiderpass, J.R. Quirós, A. Agudo, M. Rodríguez-Barranco, E. Ardanaz, B. Van Guelpen, L. Imaz, M.J. Gunter, E. Riboli, T. Waterboer, D.J. Hughes

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): J. Butt, M. Jenab, M. Pawlita, M.J. Gunter, D.J. Hughes

Writing, review, and/or revision of the manuscript: J. Butt, M. Jenab, M. Pawlita, A. Tjønneland, C. Kyrø, M.-C. Boutron-Ruault, F. Carbonnel, C. Dong, T. Kühn, H. Boeing, M.B. Schulze, A. Trichopoulos, A. Karakatsani, C. La Vecchia, D. Palli, C. Agnoli, R. Tumino, C. Sacerdote, S. Panico, B. Bueno-de-Mesquita, R. Vermeulen, I.T. Gram, E. Weiderpass, K.B. Borch, J.R. Quirós, A. Agudo, M. Rodríguez-Barranco, C. Santiuste, E. Ardanaz, B. Van Guelpen, S. Harlid, L. Imaz, A. Perez-Cornago, M.J. Gunter, S. Zouiouich, J.Y. Park, E. Riboli, A.J. Cross, A.K. Heath, T. Waterboer, D.J. Hughes

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): M. Jenab, R. Kaaks, H. Boeing, A. Karakatsani, R. Tumino, C. Sacerdote, J.R. Quirós, A. Agudo, E. Ardanaz, D.J. Hughes

Study supervision: J. Butt, M. Jenab, M. Pawlita, D.J. Hughes

Other (overseeing the prospective cohort data collection and follow-up): E. Riboli

Acknowledgments

This work was supported by The Health Research Board of Ireland project grant (grant number HRB-ILP-021, to D.J. Hughes). The authors thank Ute Koch, Monika Oppenländer, and Claudia Brandel for excellent technical assistance with the serologic measurements. The coordination of EPIC is financially supported by the European Commission (DG-SANCO) and the International Agency for Research on Cancer. The national cohorts are supported by the Danish Cancer Society (Denmark); Ligue Contre le Cancer, Institut Gustave Roussy, Mutuelle Générale de l'Éducation Nationale, and Institut National de la Santé et de la Recherche Médicale (INSERM, France); Deutsche Krebshilfe, Deutsches Krebsforschungszentrum, and Federal Ministry of Education and Research (Germany); the Hellenic Health Foundation (Greece); the Sicilian Government, AIRE ONLUS Ragusa, AVIS Ragusa, Associazione Italiana per la Ricerca sul Cancro-AIRC-Italy, and National Research Council (Italy); Dutch Ministry of Public Health, Welfare and Sports (VWS), Netherlands Cancer Registry (NKR), LK Research Funds, Dutch Prevention Funds, Dutch ZON (Zorg Onderzoek Nederland), World Cancer Research Fund (WCRF), and Statistics Netherlands (the Netherlands); Nordic Centre of Excellence programme on Food, Nutrition and Health (Norway); Health Research Fund (FIS), PI13/00061 to Granada, PI13/01162 to EPIC-Murcia, Regional Governments of Andalucía, Asturias, Basque Country, Murcia, Navarra, and the Catalan Institute of Oncology (Barcelona), Spain; Swedish Cancer Society, Swedish Scientific Council, and County Councils of Skåne and Västerbotten (Sweden); and Cancer Research UK (14136 to EPIC-Norfolk; C570/A16491 to EPIC-Oxford) and Medical Research Council [1000143 to EPIC-Norfolk, MR/M012190/1 to EPIC-Oxford (UK)].

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received December 12, 2019; revised February 5, 2020; accepted April 21, 2020; published first April 24, 2020.

References

- IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Biological agents. Volume 100 B. A review of human carcinogens. In: IARC Monographs on the evaluation of carcinogenic risks to humans. Lyon (France): International Agency for Research on Cancer; 2012. p. 1–441.
- Venerito M, Vasapolli R, Rokkas T, Delchier JC, Malfertheiner P. Helicobacter pylori, gastric cancer and other gastrointestinal malignancies. *Helicobacter* 2017; 22 Suppl 1.
- Yang F, Xu YL, Zhu RF. Helicobacter pylori infection and the risk of colorectal carcinoma: a systematic review and meta-analysis. *Minerva Med* 2019;110:464–70.
- Blase JL, Campbell PT, Gapstur SM, Pawlita M, Michel A, Waterboer T, et al. Prediagnostic Helicobacter pylori antibodies and colorectal cancer risk in an elderly, Caucasian population. *Helicobacter* 2016;21:488–92.
- Chen XZ, Schottker B, Castro FA, Chen H, Zhang Y, Hollecsek B, et al. Association of Helicobacter pylori infection and chronic atrophic gastritis with risk of colonic, pancreatic and gastric cancer: a ten-year follow-up of the ESTHER cohort study. *Oncotarget* 2016;7:17182–93.
- Limburg PJ, Stolzenberg-Solomon RZ, Colbert LH, Perez-Perez GI, Blaser MJ, Taylor PR, et al. Helicobacter pylori seropositivity and colorectal cancer risk: a prospective study of male smokers. *Cancer Epidemiol Biomarkers Prev* 2002;11: 1095–9.
- Epplein M, Pawlita M, Michel A, Peek RM Jr, Cai Q, Blot WJ. Helicobacter pylori protein-specific antibodies and risk of colorectal cancer. *Cancer Epidemiol Biomarkers Prev* 2013;22:1964–74.
- Butt J, Varga MG, Blot WJ, Teras L, Visvanathan K, Le Marchand L, et al. Serologic response to Helicobacter pylori proteins associated with risk of colorectal cancer among diverse populations in the United States. *Gastroenterology* 2019;156:175–86.
- Hu KC, Wu MS, Chu CH, Wang HY, Lin SC, Liu CC, et al. Decreased colorectal adenoma risk after Helicobacter pylori eradication: a retrospective cohort study. *Clin Infect Dis* 2019;68:2105–13.
- Riboli E, Hunt KJ, Slimani N, Ferrari P, Norat T, Fahey M, et al. European Prospective Investigation into Cancer and Nutrition (EPIC): study populations and data collection. *Public Health Nutr* 2002;5:1113–24.
- Waterboer T, Sehr P, Michael KM, Franceschi S, Nieland JD, Joos TO, et al. Multiplex human papillomavirus serology based on in situ-purified glutathione s-transferase fusion proteins. *Clin Chem* 2005;51:1845–53.
- Michel A, Waterboer T, Kist M, Pawlita M. Helicobacter pylori multiplex serology. *Helicobacter* 2009;14:525–35.
- Ryoo SK, Kim TJ, Kim ER, Hong SN, Kim YH, Chang DK. Helicobacter pylori infection and the development of advanced colorectal neoplasia. *J Clin Gastroenterol* 2019 Oct 22 [Epub ahead of print].
- Qing Y, Wang M, Lin YM, Wu D, Zhu JY, Gao L, et al. Correlation between Helicobacter pylori-associated gastric diseases and colorectal neoplasia. *World J Gastroenterol* 2016;22:4576–84.
- Lee JY, Park HW, Choi JY, Lee JS, Koo JE, Chung EJ, et al. Helicobacter pylori infection with atrophic gastritis is an independent risk factor for advanced colonic neoplasm. *Gut Liver* 2016;10:902–9.
- Butt J, Blot WJ, Shrubsole MJ, Varga MG, Hendrix LH, Crankshaw S, et al. Performance of multiplex serology in discriminating active vs. past Helicobacter pylori infection in a primarily African American population in the southeastern United States. *Helicobacter* 2020;25:e12671.
- McClain MS, Beckett AC, Cover TL. Helicobacter pylori vacuolating toxin and gastric cancer. *Toxins* 2017;9. pii: E316.
- Correa P, Houghton J. Carcinogenesis of Helicobacter pylori. *Gastroenterology* 2007;133:659–72.
- Grahn N, Hmani-Aifa M, Fransen K, Soderkvist P, Monstein HJ. Molecular identification of Helicobacter DNA present in human colorectal adenocarcinomas by 16S rDNA PCR amplification and pyrosequencing analysis. *J Med Microbiol* 2005;54:1031–5.
- Jones M, Helliwell P, Pritchard C, Tharakan J, Mathew J. Helicobacter pylori in colorectal neoplasms: is there an aetiological relationship? *World J Surg Oncol* 2007;5:51.
- Soylu A, Ozkara S, Alis H, Dolay K, Kalayci M, Yasar N, et al. Immunohistochemical testing for Helicobacter pylori existence in neoplasms of the colon. *BMC Gastroenterol* 2008;8:35.
- Jackson L, Britton J, Lewis SA, McKeever TM, Atherton J, Fullerton D, et al. A population-based epidemiologic study of Helicobacter pylori infection and its association with systemic inflammation. *Helicobacter* 2009;14:108–13.
- Gao JJ, Zhang Y, Gerhard M, Mejias-Luque R, Zhang L, Vieth M, et al. Association between gut microbiota and Helicobacter pylori-related gastric lesions in a high-risk population of gastric cancer. *Front Cell Infect Microbiol* 2018;8:202.

***Helicobacter pylori* Serology and Colorectal Cancer**

24. D'Onghia V, Leoncini R, Carli R, Santoro A, Giglioni S, Sorbellini F, et al. Circulating gastrin and ghrelin levels in patients with colorectal cancer: correlation with tumour stage, *Helicobacter pylori* infection and BMI. *Biomed Pharmacother* 2007;61:137–41.
25. Fireman Z, Trost L, Kopelman Y, Segal A, Sternberg A. *Helicobacter pylori*: seroprevalence and colorectal cancer. *Isr Med Assoc J* 2000;2:6–9.
26. Georgopoulos SD, Polymeros D, Triantafyllou K, Spiliadi C, Mentis A, Karamanolis DG, et al. Hypergastrinemia is associated with increased risk of distal colon adenomas. *Digestion* 2006;74:42–6.
27. Hartwich A, Konturek SJ, Pierzchalski P, Zuchowicz M, Labza H, Konturek PC, et al. *Helicobacter pylori* infection, gastrin, cyclooxygenase-2, and apoptosis in colorectal cancer. *Int J Colorectal Dis* 2001;16:202–10.
28. Machida-Montani A, Sasazuki S, Inoue M, Natsukawa S, Shaura K, Koizumi Y, et al. Atrophic gastritis, *Helicobacter pylori*, and colorectal cancer risk: a case-control study. *Helicobacter* 2007;12:328–32.
29. Strofilas A, Lagoudianakis EE, Seretis C, Pappas A, Koronakis N, Keramidaris D, et al. Association of *Helicobacter pylori* infection and colon cancer. *J Clin Med Res* 2012;4:172–6.

Cancer Epidemiology, Biomarkers & Prevention

Antibody Responses to *Helicobacter pylori* and Risk of Developing Colorectal Cancer in a European Cohort

Julia Butt, Mazda Jenab, Michael Pawlita, et al.

Cancer Epidemiol Biomarkers Prev 2020;29:1475-1481. Published OnlineFirst April 24, 2020.

Updated version Access the most recent version of this article at:
doi:[10.1158/1055-9965.EPI-19-1545](https://doi.org/10.1158/1055-9965.EPI-19-1545)

Supplementary Material Access the most recent supplemental material at:
<http://cebp.aacrjournals.org/content/suppl/2020/04/24/1055-9965.EPI-19-1545.DC1>

Cited articles This article cites 25 articles, 3 of which you can access for free at:
<http://cebp.aacrjournals.org/content/29/7/1475.full#ref-list-1>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link
<http://cebp.aacrjournals.org/content/29/7/1475>.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.