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Research paper

Distribution of organic-walled dinoflagellate cysts in surface sediments of the southern Caribbean and the eastern tropical Pacific and its environmental implications

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Keywords: Eastern tropical Pacific Dinoflagellates Biogeography Southern Caribbean Upwelling Plankton ecology ABSTRACT

Little is known about the marine palynology of the neotropical oceans. Here, we present the first comprehensive study of organic-walled dinoflagellate cyst assemblages in 52 surface marine sediment samples from the southern Caribbean (SC) and the eastern tropical Pacific (ETP) and explain how these assemblages relate to sea-surface parameters. Multivariate analyses show that of the several environmental parameters considered, sea-surface salinity (SSS), phosphate concentrations, and nitrate concentrations best explain the relative abundances and the geographic distribution of dinoflagellate cysts in the studied area. Consistent with regional differences in marine productivity, dinoflagellate cyst concentrations were markedly different in the SC (1979 \pm 1053 cysts/g) and the ETP ($3105 \pm 1956 \text{ cysts/g}$). Sediments of the SC are characterized by high relative abundances of the Spiniferites spp. group, cysts of Protoceratium reticulatum, and the presence of rare dinocyst taxa. The dinocyst assemblages from the ETP are characterized by higher relative and absolute abundances of Bitectatodinium spongium, Brigantedinium spp., and Echinidinium aculeatum. This pattern is explained by the more eutrophic nature of the ETP compared to the highly saline and oligotrophic SC. Average values of the Shannon Index (H') from the ETP are higher compared to the SC, demonstrating that dinocysts respond to the greater climatic variability that characterizes the ETP. Brigantedinium spp. abundances are negatively correlated to H' diversity supporting its dominant character within the dinocyst community. This work fills a gap in our knowledge on modern dinocyst distribution in neotropical oceans and provides a modern reference for interpreting down core dinocyst variations and paleocenographical reconstructions in the region.

1. Introduction

Dinoflagellates are a group of marine and freshwater planktonic eukaryotic organisms (Taylor et al., 2008). As one of the most important primary producers, dinoflagellates are not only responsible for transferring energy from the base of the trophic food web but also for carbon export to the deep ocean (Müller-Karger et al., 2004; Bravo and Figueroa, 2014; Guidi et al., 2016). Their different feeding strategies that

include autotrophy, heterotrophy, and mixotrophy along with their diverse habitats allowed dinoflagellates to thrive in the ocean since the Mesozoic (Schnepf and Elbrächter, 1992; Gaines and Elbraechter, 1987; Dale, 2001; Taylor et al., 2008). Their complex life cycle, which alternates asexual and sexual phases, has further allowed these organisms to successfully endure severe environmental change. Following both sexual (hypnozygote) and asexual reproduction, resting cysts (henceforth referred as dinocyst) can develop (Wall, 1965; Figueroa et al., 2008;

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Fig. 1. Geographic location of surface sediment samples included in the dinocyst database of Marret et al., 2020. The black square highlights the study area and the location of the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Map modified from Marret et al. (2020).

Bravo and Figueroa, 2014). About 15–20% of species are capable of producing viable dinocysts that can be preserved in sediments and rocks over long-term temporal scales (Head, 1996).

Dinocysts have provided paleontologists with a valuable tool based on which to interpret and infer evolutionary trajectories (Wall and Dale, 1968; Evitt, 1985) and past ecological and climatic conditions (Dale, 1983; Dale, 2001; de Vernal et al., 1997). Comprehensive research on dinocyst occurrences in surface sediments from the ocean has also provided a wealth of information on their distributions and the main environmental variables driving their occurrences (Harland, 1983; Rochon et al., 1999; Marret and Zonneveld, 2003; Pospelova et al., 2008; Zonneveld et al., 2013; Marret et al., 2020; Van Nieuwenhove et al., 2020). The most critical environmental factors that control the distribution of dinocysts in the modern ocean seem to be sea-surface temperature (SST), sea-surface salinity (SSS), phosphate concentrations ([PO₄]), nitrate concentrations ([NO₃]), water depth (WD), and distance to the coast (DC) (Wall et al., 1977; Marret and Zonneveld, 2003; Zonneveld et al., 2013).

The diversity and abundance of dinocysts is seemingly higher in temperate and tropical latitudes, but endemic polar and sub-polar dinocyst species have also been recorded (Mudie, 1992; Montresor et al., 2003; Limoges et al., 2018; Marret et al., 2020). Despite this, the majority of studies on dinocyst morphology and ecology have focused on the northern temperate and high latitudes (de Vernal et al., 1997, 2001, 2020; Rochon et al., 1999; Orlova et al., 2004; Marret et al., 2001; Esper and Zonneveld, 2002; Matthiessen et al., 2005; Crouch et al., 2010; Prebble et al., 2013; Zonneveld et al., 2013; Price et al., 2016; Mudie et al., 2017; Allan et al., 2020; Marret et al., 2020) (Fig. 1, blue dots), whereas dinocyst distribution in tropical areas such as the southern Caribbean Sea and the eastern tropical Pacific Ocean remains



Fig. 2. a- Oceanic currents, upwelling cells, Intertropical Convergence Zone (ITCZ) seasonal position, and drainage system in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Oceanic currents: Caribbean Current (CC), Pacific Current (PC), North Equatorial Counter Current (NECC), South Equatorial Current (SECC). Hatched pattern: Upwelling zones. The dotted lines delimit the position of the ITCZ in June–July–August (JJA) and January–February–March (JFM). Also shown are the Central American isthmus (CAI) and the Andes Cordillera. The relevant rivers for this study are shown in white; note how the Magdalena, Atrato, and Orinoco rivers flow into the SC, and the San Juan and Patia rivers flow into the ETP (Restrepo and Kjerfve, 2004; Poveda et al., 2006, 2014; González et al., 2008). b-Location of studied samples (the red dots show the location of samples from the SC and the black dots the location of samples from the ETP).

Locality (light typeface = southern Caribbean (SC), bold typeface = eastern tropical Pacific (ETP)); longitude (Long); latitude (Lat); sea-surface temperature (SST) in °C, Annual: SST_A; June–July–August: SST_JJA; January–February–March: SST_JFM; the sea-surface cancentration of phosphates [PO4] in µmol/L, Annual: [PO4]_A; June–July–August: [PO4]_JJA; January–February–March: [NO3]_in µmol/L, Annual: [NO3]_A; June–July–August: [NO3]_JJA; January–February–March: [NO3]_JFM. Water depth in m (WD); Distance to the coast in m (DC). All SSTs, SSSs, and nutrient concentrations were downloaded from the World Ocean Atlas (Boyer et al., 2013). Repositories (R): Laboratorio de Biología Molecular Marina (BIO); Universidad EAFIT (EAF); Lamont–Doherty Earth Observatory (LDE); Laboratorio de Palinología y Paleoecología tropical (PPT); Ocean Drilling Program (ODP), Instituto Colombiano del Petróleo (ICP). Device: Sampling method/tool.

Locality	Long	Lat	SST_A	SST_JJA	SST_JFM	SSS_A	SSS_JJA	SSS_JFM	[PO4] _A	[P] O4_JJA	[P O4] _ JFM	[NO3]_A	[NO3] _JJA	[NO3]_JFM	WD	DC	R	Device
CUEVAS	-75.6	10	28.60	28.7	27.2	34 37	35.7	36.8	0.02	0.01	0.47	0.1	0	0	4	2	BIO	
L VARADERO	-75.6	10	28.69	28.7	27.2	34.37	35.7	36.8	0.02	0.01	0.47	0.1	0	0	4	2	BIO	Diving
BC13-100	-81.2	16	28.11	27.7	26.7	36.14	36.2	36.1	0.042	0.03	0.05	0.96	0	0	4570	244.9	EAF	Piston Core
VM28-115	-72.4	14	28.29	27.3	26.4	32.18	35.9	35.7	0.056	0.26	0.06	0.3	4.578	0	2109	178	LDE	Piston Core
TRI 49–19 I	-83	17	28.11	28	26.1	36.04	36.1	35.9	0.04	0.03	0.05	1.21	0	0	1830	177.5	EAF	Piston Core
NECOCLÍ	-76.8	8.4	28.5	28.9	27	33.69	36.2	36.8	0.01	0.01	0.47	0.1	0.1	3.4	15	1.56	PPT	Diving
ZAPSURRO	-77.5	8.8	27.27	28.9	27	35.66	36.2	36.8	0.014	0.01	0.47	0.13	0	3.4	30	8.63	PPT	Diving
RC13–169	-65	13	27.51	28.3	26	35.72	36.5	35.8	0.065	0.06	0.03	0.49	2.2	0.867	4649	177.5	LDE	Piston Core
VM12-110	-68.7	14	27.52	27.7	26.3	35.72	35	35.7	0.108	0.03	0.24	0.61	4.578	0	5040	159.6	LDE	Piston Core
RC09-052	-69.8	14	27.41	27.4	26.2	35.72	36.5	35.9	0.081	0.03	0.105	0	4.578	0	4599	174.9	LDE	Piston Core
VM20-012	-77.2	11	27.86	27.3	26.3	35.92	36.2	35.1	0.027	0	0.05	0.1	4.578	0	3396	177	LDE	Piston Core
RC11-239	-75.5	12	27.23	27.4	26.2	35.62	36.7	36.3	0.032	0.01	0.05	0	4.578	0	3301	122.3	LDE	Piston Core
PALOMINO4	-73.6	11	26.4	26.9	25.5	36.62	36.3	37.1	0.015	0.01	0.05	0.71	4.578	0	25	12.05	PPT	Dredging
POPOYA13	-72.8	12	27.25	25.7	23.8	35.98	36.3	36.5	0.112	0.01	0.05	0.27	4.578	0	1	24.95	PPT	Dredging
VM28-07	-75.8	12	27.08	27.7	26.2	35.9	36.3	36.3	0.01	0.01	0.05	0	4.578	0	2869	99.21	ODP	Piston Core
RC09–045	-65.5	14	27.5	28.3	26.2	35.72	35	35.9	0.037	0.075	0.02	0	0	0	4674	436.4	LDE	Piston Core
VM24-031	-76.6	12	27.56	27.7	26.2	36.29	36.2	36.3	0.06	0	0.05	0.1	4.578	0	3623	218.6	LDE	Piston Core
VM24-032	-76.1	11	28.06	27.8	26.9	35.97	36.3	36.8	0	0	0.05	0.1	0	0	2704	94.8	LDE	Piston Core
VM19-017	-74	13	27.52	26.5	25.1	35.92	36.3	36.7	0.026	0.01	0.05	0.03	4.578	0	3872	151.8	LDE	Piston Core
POPOYA9	-73.1	12	28.62	25.7	23.8	36.02	36.3	37.1	0.015	0.01	0.05	0.71	4.578	0	1	17.13	PPT	Dredging
RINCÓN MAR3	-75.8	9.8	28.86	28.8	27.2	34.31	35.7	36.8	0.01	0.01	0.47	0.1	0	0	36	2.61	PPT	Dredging
RC13–148	-74.3	12	27.08	26.9	25.5	35.7	36.3	37.1	0.015	0.01	0.05	0.71	4.578	0	1257	45.07	LDE	Piston Core
C.DIQUE	-75.5	10	29.2	28.7	27.2	34.37	35.7	36.8	0.02	0.01	0.47	0.1	0	0	4	2	BIO	Diving
BERRUGAS16	-76	9.7	28.86	28.8	27.2	34.31	35.7	36.8	0.01	0.01	0.47	0.1	0	0	26	20.04	PPT	Dredging
EGORO1	-78.6	2.5	26.75	26.8	26.2	29.99	32.9	31.8	0.382	0.35	0.15	1.89	0	0.1	76	6.47	ICP	Piston Core
EGORO5	-79	2.5	26.77	26.8	26.2	32.36	32.9	31.8	0.382	0.35	0.15	3.85	0	0.1	210	47.17	ICP	Piston Core
EGORO7	-78.9	2.3	26.47	26.8	26.2	32.36	32.9	31.8	0.382	0.35	0.15	3.85	0	0.1	768	18.23	ICP	Piston Core
EGORO12	-79.3	2.2	26	26.8	26.2	32.71	32.9	31.8	0.382	0.35	0.15	1.89	0	0.1	591	60.97	ICP	Piston Core
EGORO15	-79.3	1.7	26.47	26.6	26	32.71	32.1	32.2	0.382	0.35	0.215	1.89	0	0.13	839	33.69	ICP	Piston Core
EGORO14	-79.3	1.9	20.47	20.0	20	32.71	32.1	32.2	0.382	0.35	0.215	1.89	0	0.13	/9/	38.5	ICP	Piston Core
EGORO10	-/9.2	1.9	20.23	20.0	20	32.39 23.71	32.1 22.0	32.2 21.9	0.382	0.35	0.215	1.89	0	0.15	570	39.34	ICP	Piston Core
EGOROI7	-/9.1	2.2	20.47	20.0	20.2	20.42	32.9	21.0	0.382	0.35	0.15	1.09	0	0.1	653	2.77	ICP	Piston Core
EGORO20	-798	2.5	26.47	20.8	20.2	32.36	32.1	31.0	0.382	0.33	0.13	3.85	0	0.1	664	85	ICP	Piston Core
MFRA2	-77.5	46	20.47	20.4	26.1	26.65	32.3	30.3	0.355	0.185	0.125	1 16	0	0.1	94	15.97	ICP	Piston Core
MERA3	-77.5	5.1	28.1	26.4	20.1	26.65	32.3	31.1	0.355	0.185	0.125	1.16	0	0.1	66	9.16	ICP	Piston Core
MERA5	-77.9	4.6	26.6	26.4	27	29.99	32.3	31.1	0.355	0.185	0.125	1.16	0	0.1	1534	61.4	ICP	Piston Core
MERA8	-77.7	4.8	26.97	27.8	26.1	27.6	32.3	30.3	0.24	0.185	0.125	6.03	0	0.1	1519	33.12	ICP	Piston Core
MERA13	-77.5	5.2	28.69	27.8	27	28.23	32.6	32.3	0.24	0.45	0.22	6.03	0	0.5	66	9.36	ICP	Piston Core
VM20-015	-84.3	7	27.63	28.1	25.8	32.77	31.3	32.7	0.16	0.19	0.47	0.5	0	3.4	1736	191.2	LDE	Piston Core
VM15-011	-79.1	7.8	26.05	27.5	25.6	33.22	32.5	32.1	0.23	0.185	0.405	0.41	0	6	249	74.42	LDE	Piston Core
RC10-253	-79.4	6.5	27.59	26.8	26.2	32.06	32.9	31.8	0.303	0.35	0.15	0.93	0	0.1	3219	202.9	LDE	Piston Core
RC9-065	-79.3	2.5	26.45	27.8	26.1	32.36	32.3	30.3	0.382	0.185	0.125	1.89	0	0.1	2074	70.03	LDE	Piston Core
VM21-216	-77.6	5.3	27.4	26	25.4	28.39	33.5	33.4	0.24	0.66	0.57	6.03	0.2	5.7	761	22.8	LDE	Piston Core
KAMA3	-81	0.5	25.84	26.1	26	33.17	32.1	32.2	0.236	0.35	0.193	0	0	0.13	3797	106	EAF	Piston Core
KAMA14	-79.2	1.6	26.39	26.6	26.4	32.64	32.8	32.6	0.313	0.35	0.215	3.85	0.2	0.15	784	24.93	EAF	Piston Core
KAMA24	-79.9	1.4	26.27	26	26.9	34.57	33.3	32.7	0.313	0.66	0.57	3.85	0	5.7	1623	47.2	EAF	Piston Core
KAMA1	-80.7	0.3	25.7	26	26.9	33.36	33.3	32.7	0.375	0.66	0.57	0	0	5.7	3046	74.79	EAF	Piston Core
KAMA2	-80.8	0.5	25.97	27.4	26.5	26.65	32.5	31.7	0.375	0.2	0.47	0	0	0.3	1315	78	EAF	Piston Core
KNR176–2 JPC9	-77.9	6.8	26.92	26.4	27	27.02	32.3	31.1	0.268	0.185	0.125	1.15	0	0.1	288	20.92	LDE	Piston Core
KNR176-2-	-78	4.7	23.17	27.5	26.6	30.74	32.5	31.7	0.355	0.112	0.47	1.16	0	3.4	2195	68.21	LDE	Piston Core
JPC32																		
KNR176-2-MC4	-78.2	7.3	28.65	28.5	27.8	30.22	32.3	31.9	0.162	0.45	0.39	0.24	6	3.2	2121	25.09	LDE	Piston Core

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Fig. 3. Surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), b-June–July–August (SST_JJA), c- January–February–March (SST_JFM). Sea-surface salinity (SSS) in psu: d- Annual (SSS_A), e- June–July –August (SSS_JJA), f-January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: g- Annual ([PO4]_A), h- June–July–August ([PO4]_JJA), i-January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: j- Annual ([NO3]_A), k- June–July–August ([NO3]_JJA), l-January–February–March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).

underexplored (Fig. 1, black square). While the range of climatic variability in these areas is smaller than that which is observed in temperate and high latitudes (Boyer et al., 2013), the few studies available suggest that the composition and distribution of dinocysts vary in space and time, in response to some of the above-mentioned environmental parameters (Wall et al., 1977; Lewis et al., 1990; Vink et al., 2000; Vásquez-Bedoya et al., 2008; González et al., 2008; Limoges et al., 2010, 2013; Bringué et al., 2019).

The purpose of this study was to examine the spatial distribution of dinocysts in marine surface sediments from the southern Caribbean (SC) and the eastern tropical Pacific (ETP) in order to: 1) document the dinoflagellate assemblages on a regional scale; 2) assess the main environmental parameters that drive the distribution of dinocysts and how these variables shape the abundance and diversity in both areas; and 3) set a modern baseline for dinocyst assemblages based on which paleoenvironmental reconstructions can be assessed.

2. Oceanography of the southern Caribbean (SC) and eastern tropical Pacific (ETP) areas

The study area is located between latitudes 0° N to16° N and longitudes 65° W to 84° W (Fig. 2). The modern oceanography of the SC and ETP is the result of a complex geologic history in which the collision between the western margin of the South American Plate and the Caribbean Plate gave rise 13–4 Ma to the Central American Isthmus (CAI) and Colombia's eastern Cordillera (Fig. 2a) (Mora et al., 2006; Montes et al., 2015; O'Dea et al., 2016). This geological process brought a long-established connection between the SC and the ETP to an end, and, in turn, changed the hydrographic and atmospheric conditions in these basins (O'Dea et al., 2016). The closure of the CAI caused a major reorganization of the ocean circulation. As a result, the waters of the Caribbean became highly saline (36 psu) and over saturated in CaCO₃, contrasting with the low saline (33 psu) and more corrosive waters characteristic of the Pacific (Haug and Tiedemann, 1998). Also, the SC is characterized by lower mean primary-productivity values (71–141 g C



Fig. 4. Distribution maps of sedimentation rates (cm/yr.) used for the age estimation of samples in a- the southern Caribbean (SC), b- the eastern tropical Pacific (ETP).

 $m^{-2} y^{-1}$) than the mean value of 120–363 g C $m^{-2} y^{-1}$ estimated for the ETP (Longhurst et al., 1995; Boyer et al., 2013). The hydroclimatology of the SC and ETP is dominated by seasonal latitudinal shifts of the Intertropical Convergence Zone (ITCZ) from its northernmost position in the months of June–July–August (JJA) to its southernmost position in the months of January–February–March (JFM) (Fig. 2a).

2.1. The southern Caribbean (SC)

The SC is located in the western margin of the Atlantic Ocean on the continental platforms of Panama, Colombia, Nicaragua, and Venezuela (Fig. 2a). In general, nutrient-depleted surface waters characterize the Caribbean Sea except for areas adjacent to river mouths and local up-welling zones (Longhurst et al., 1995; Boyer et al., 2013).

The primary oceanic current in the SC is the Caribbean Current (CC) (Fig. 2a) that feeds the Gulf Current, a surficial current that in turn regulates the global climate by transporting warm, humid and saline waters to the North Atlantic (Haug and Tiedemann, 1998).

In the SC (Fig. 2a), the Magdalena, Atrato, and Orinoco rivers discharge 666 t km⁻² yr⁻¹ of sediments and approximately 47,574 m³ s⁻¹ of freshwater into their adjacent coastal areas (Hernandez-Guerra and Joyce, 2000; Restrepo and Kjerfve, 2004; Restrepo and López, 2008; Mora et al., 2020). In addition to river discharge, another mechanism that explains the localized high levels of primary productivity in the SC consists in the trade winds that impinge on the coastal areas of the Cariaco Basin and the Guajira Peninsula from January to March allowing nutrient-rich cold waters to upwell (Milliman, 1990; Müller-Karger et al., 2004).

The rainy season in SC begins during the late boreal summer when the ITCZ is directly over the southern Caribbean (Fig. 2a; Hu et al., 2004; Müller-Karger et al., 2004). A permanent Caribbean low-level jet, which enhances evaporation and carries humidity across Central America is, to some extent, responsible for a greater salinity in the SC compared to the ETP (Wang, 2007; Poveda et al., 2014).

2.2. The Eastern Tropical Pacific (ETP)

The ETP is located at the eastern end of the equatorial Pacific Ocean, off northwestern South America, on the Panama Basin (Fig. 2a). The western margin and the central part of the ETP is an area of high primary productivity with high nutrient levels (Restrepo and López, 2008; Cabarcos et al., 2014).

The main oceanic currents in the ETP are the Pacific Current (PC), North Equatorial Counter Current (NECC), and South Equatorial Current (SECC) (Fig. 2a). The current system of the ETP is influenced by the Peru–Chile Current System and represents the main route for the exchange of nutrients from high to low latitudes in the South Pacific (Cabarcos et al., 2014).

The ETP is characterized by having river discharges greater than 4709 m³ s⁻¹ that deliver an excess of nutrients into the ocean, hence enhancing primary productivity (Poveda et al., 2006, 2014; Restrepo and López, 2008; Hidalgo et al., 2015). The San Juan River and the Patia River are the main Colombian rivers feeding the ETP (Fig. 2a). Despite these rivers being much shorter, the San Juan and Patia rivers deliver ~8 times the amount of water delivered into the ocean by the rivers that drain into the SC (Restrepo and Kjerfve, 2004). Similarly, the estimated sediment load that drains into the Pacific Ocean from Colombia is ca. 1260 t km⁻² yr⁻¹ (Restrepo and Kjerfve, 2004).

Trade winds drive the surface oceanic currents that include the North Equatorial Counter Current (NECC), the South Equatorial Currents (SEC), and the Pacific current (PC) (Fig. 2a; Fiedler and Talley, 2006).

The Colombian Pacific coast experiences an annual average precipitation of 12,500 mm/year that helps to maintain the ETP's low salinity compared to the SC (Poveda and Mesa, 1997; Restrepo and López, 2008; Durán-Quesada et al., 2012; Boyer et al., 2013).

3. Material and methods

3.1. Sediment samples

Samples were collected over the 1958–2014 period. Twenty-four core-top samples (\sim 0–1 cm) from the SC and twenty-eight core-top samples (\sim 0–1 cm) from the ETP were studied. The samples represent a gradient of coastal (neritic) to oceanic (pelagic) depths on the continental margins and abyssal plain of northwestern South America (Fig. 2b). Details of the geographic location, distance to the coast, water depth, and sampling devices are provided in Table 1.

3.2. Oceanographic parameters

Annual and seasonal sea-surface temperature (SST), sea-surface salinity (SSS), and values of concentrations of sea-surface phosphates [PO₄] and nitrates [NO₃] were downloaded from the World Ocean Atlas and averaged through the 1954 to 2013 period at a resolution of 0.25 degrees (Boyer et al., 2013). A shapefile that contained the values for the environmental parameters was then loaded into QGIS (QGIS Development Team, 2014). The yearly and seasonal average values for each environmental variable were extrapolated and plotted on a map using

Sedimentation rates (cm/kyr, cm/yr) based on biostratigraphy, magnetostratigraphy, ¹⁴C chronology and, chemostratigraphy for the southern Caribbean (SC) and eastern tropical Pacific (ETP). Long: longitude, Lat: latitude, and SR: sedimentary rate (cm/kyr, cm/yr).

Label	Basin	Locality	Long	Lat	SR(cm/	SR(cm/	Method	Reference
					kyr)	yr)		
1	SC	998	-82.93	19	1.9	0.0019	Nannoplankton	Kameo and Bralower, 2000
2	SC	1000	-80	17	2.73	0.0027	Nannoplankton	Kameo and Bralower, 2000
3	SC	PC-117	-78.6	18.15	0.7	0.0007	Oxygen isotopes	Sullivan, 1992
4	SC	PC-108	-78.8	17.86	0.26	0.00026	Oxygen isotopes	Sullivan, 1992
5	SC	PC-11	-77.64	17.46	10.2	0.0102	Oxygen isotopes	Sullivan, 1992
6	SC	PC-21	-77.48	17.6	0.48	0.00048	Oxygen isotopes	Sullivan, 1992
7	SC	PC-37	-78.35	17.51	0.31	0.00031	Oxygen isotopes	Sullivan, 1992
8	SC	PC-54	-77.1	17.14	0.129	0.00012	Oxygen isotopes	Sullivan, 1992
9	SC	PC-46	-77.71	17.16	3.68	0.0036	Oxygen isotopes	Sullivan, 1992
10	SC	PC-65	-76.31	17	0.31	0.00031	Oxygen isotopes	Sullivan, 1992
11	SC	PC-72	-76.27	16.25	1.93	0.0019	Oxygen isotopes	Sullivan, 1992
12	SC	V28-119	-75.7	16.6	0.3	0.0003	Oxygen isotopes	Sullivan, 1992
13	SC	PC-89	-78.03	17	0.34	0.00034	Oxygen isotopes	Sullivan, 1992
14	SC	PC-73	-76.37	16.41	0.27	0.00027	Oxygen isotopes	Sullivan, 1992
15	SC	PC-75	-76.56	16.6	0.48	0.00048	Oxygen isotopes	Sullivan, 1992
16	SC	999	-79	13	3.36	0.0033	Nannoplankton	Kameo and Bralower, 2000
17	SC	502	-79.4	11.49	7.5	0.0075	Paleomag	Gardner, 1982
18	SC	1002	-65	10.42	35	0.035	Oxygen isotopes	Peterson et al., 2000
19	ETP	852	-110.07	5.29	1.2	0.012	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
20	ETP	851	-110.57	2.77	2.5	0.0025	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
21	ETP	850	-110.52	1.29	2.5	0.0025	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
22	ETP	849	-110.51	0.18	2.5	0.0025	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
23	ETP	848	-110.47	29.99	5	0.0005	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
24	ETP	845	-94.59	9.58	1.2	0.013	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
25	ETP	844	-90.48	7.92	2.5	0.025	Magnetostratigraphy/	Kemp, 1995
							Nannoplankton	
26	ETP	TR163-22	-92.23	3.99	10	0.01	Biostratigraphy	Lea et al., 2000
27	ETP	TR163–19	-90.95	2.26	3	0.003	Biostratigraphy	Lea et al., 2000
28	ETP	847	-95.32	0.193	4	0.004	Biostratigraphy	McCartney et al., 1995
29	ETP	1226B	-90.81	-3.09	2.4	0.0024	¹⁴ C geochronology	http://www-odp.tamu.edu/publications/201_SR/108/
								108_5.htm
30	ETP	TR163-31	-83.95	3.6	2	0.002	Biostratigraphy	Kienast et al., 2007
31	ETP	1239	-82.08	-0.67	3.5	0.0035	Bioestratigraphy	Rincón-Martínez et al., 2010
32	ETP	1230D	-80.58	-9.11	2	0.002	¹⁴ C geochronology	http://www-odp.tamu.edu/publications/201_SR/108/
								108_5.htm
33	ETP	1227B	-79.95	-8.99	116.7	0.1167	¹⁴ C geochronology	http://www-odp.tamu.
								edu/publications/201_SR/108/108_t2.htm#1003206
34	ETP	1229E	-77.95	-10.97	32.3	0.0323	¹⁴ C geochronology	http://www-odp.tamu.
								edu/publications/201_SR/108/108_t2.htm#1003206
35	ETP	1231C	-81.94	-12.02	1.5	0.0015	¹⁴ C geochronology	http://www-odp.tamu.edu/publications/201_SR/108/
								108_5.htm
36	ETP	1228B	-78.07	-11.06	40.4	0.0404	¹⁴ C geochronology	http://www-odp.tamu.
								edu/publications/201_SR/108/108_t2.htm#1003206

the freeware program Ocean Data View (Schlitzer, 2017) (Fig. 3, Table 1). Seasonal values of environmental parameters for each locality were plotted for the periods when the ITCZ is at its northernmost and southernmost position in June–July–August (JJA) and Januar-y–February–March (JFM), respectively (Figs. 2a, 3). Water depth values and the distances from the coast for each locality were measured and recorded during each oceanographic expedition.

3.3. Palynological analyses

Sediment samples were dried overnight at 60 °C. Between one and two grams of sediment were processed for palynological analyses. Samples were treated with 10% hydrochloric acid (HCl) and 40% cold hydrofluoric acid (HF) to dissolve carbonates and silicates, respectively (de Vernal et al., 1999) and no oxidation was performed. Tablets of *Lycopodium clavatum* (batch number 1031) containing 20,848 \pm 691 spores were added during the decalcification process to achieve a quantitative (palynomorphs/g) analysis of the data (Wood et al., 1996). The demineralized and spiked residue was processed in an ultrasonic bath for no longer than 30 s and subsequently sieved through a 10 µm nylon mesh. Aliquots were strewn onto microscope slides and mounted using glycerin jelly. Dinocysts were counted using a Zeiss Imager A.2 optical microscope under $40 \times$ and $100 \times$ objectives and photographed using an Olympus FV1000 laser scanning confocal microscope with a 60×/1.42 NA oil immersion objective. Generic and specific dinocyst classification followed the taxonomic nomenclature of Williams et al. (2017). Zonneveld and Pospelova (2015) was used as determination key for modern dinocysts. Brigantedinum cariacoense, Brigantedinium simplex, and other round brown cysts were grouped as Brigantedinium spp. when the orientation of the cysts precluded determination at the species level. Most of the species belonging to the genus Spiniferites found in the SC and the ETP were assigned to the Spiniferites ramosus that has a cosmopolitan distribution. Very few specimens of S. bentorii and S. mirabilis were found. We grouped all Spiniferites species under the category Spiniferites spp.

In the case of "Operculodinium centrocarpum sensu Wall and Dale

Nearest locality from which age estimates were calculated (NLA), estimated age (yr) for top sediments according to the sedimentation rate framework of reference for all studied sites.

Locality	NLA	Estimated age first cm of sediment (yr)
CUEVAS	502	133
L.VARADERO	502	133
RC13–100	998	526
VM28-115	1002	29
TRI 49–19 I	998	526
NECOCLÍ	502	133
ZAPSURRO	502	133
RC13–169	1002	29
VM12-110	1002	29
RC09-052	1002	29
VM20-012	502	133
RC11-239	502	133
PALOMINO4	1002	29
POPOYA13	1002	29
VM28-07	502	133
RC09-045	1002	29
VM24-031	502	133
VM24-032	502	133
VM19-017	1002	29
POPOYA9	1002	29
RINCÓN MAR3	502	133
RC13–148	502	133
C.DIQUE	502	133
BERRUGAS16	502	133
EGORO1	1239	286
EGORO5	1239	286
EGORO7	1239	286
EGORO12	1239	286
EGORO15	1239	286
EGORO14	1239	286
EGORO16	1239	286
EGORO17	1239	286
EGORO9	1239	286
EGORO20	1239	286
MERA2	TTRI163-31	500
MERA3	TTRI163-31	500
MERA5	TTRI163-31	500
MERA8	TTRI163–31	500
MERA13	TTRI163-31	500
VM20-015	844	40
VM15-011	TTRI163-31	500
RC10–253	TTRI163-31	500
RC9–065	TTRI163-31	500
VM21-216	TTRI163–31	500
KAMA3	TTRI163–31	500
KAMA14	TTRI163-31	500
KAMA24	TTRI163-31	500
KAMA1	TTRI163-31	500
KAMA2	TTRI163-31	500
KNR176–2 JPC9	TTRI163-31	500
KNR176–2-JPC32	TTRI163-31	500
KNR176-2-MC4	TTRI163-31	500

(1966)", we follow Paez-Reyes and Head (2013) in that cysts circumscribed by the morphology described by Wall and Dale (1966) are referred to as cysts of Protoceratium reticulatum. We justify our choice based on several lines of evidence: 1) Mertens et al. (2018) described the theca-based dinoflagellate species Pentaplacodinium saltonense. According to these authors, the cysts produced by P. saltonense more closely resemble to the cyst-based species Operculodinium israelianum and Operculodinium psilatum; 2) given the tabulation similarities between P. saltonense, its cysts, and Operculodinium bahamense, it is likely that all Operculodinium species are (were) produced by a species related to P. saltonense; 3) molecular evidence also suggests that the genera Pentaplacodinium and Protoceratium are not related at the phylogenetic level and that they form two distinctive clades (Mertens et al., 2018; Wang et al., 2019); 4) the cysts of Protoceratium reticulatum were initially described to be from the North Sea where cysts of O. centrocarpum sensu Wall and Dale occur abundantly (de Vernal et al., 2020), whereas warmwater *O. israelianum*, and *Pentaplacodinium* have not been recorded. All this evidence strongly supports our argument that cysts of *Protoceratium reticulatum* are the same as what is referred to as *O. centrocarpum* sensu Wall and Dale (1966), and that the use of the latter name only generates confusion.

3.4. Age of the sediments in the southern Caribbean and eastern tropical Pacific

The age of the surface sediments (top 0–1 cm) is unknown, as is the case in virtually all dinocyst studies performed over the past forty years. Indeed, when using surface sediments from piston cores (as we did here for 90% of the samples presented), the uppermost sediments could be missing and assuming a present-day age for the core-top samples is not necessarily accurate. We argue, however, that the top sediment represents the last millennium or less. To prove our point, we calculated the sediment accumulation rates and the estimated age of the samples using the sedimentation rates from nearby localities that have biostratigraphy (McCartney et al., 1995; Kameo and Bralower, 2000; Rincón-Martínez et al., 2010; Kemp, 1995), magnetostratigraphy (Kent and Spariosu, 1982), ¹⁴C geochronology (ODP samples) or chemostratigraphy (Gardner, 1982; Sullivan, 1992; Peterson et al., 2000; Lea et al., 2000; Kienast et al., 2007) (Fig. 4; Tables 2, 3).

3.5. Assemblage data and statistical analyses

Two different metrics to express palynomorph representation were used throughout this paper: 1) the concentration of palynomorphs (dinocyst, pollen, and spores) per gram of dry sediment (Table 5) (Stockmarr, 1971; Maher Jr, 1981), and 2) the proportion of each dinocyst taxon or group relative to the total dinocyst counts in a given assemblage.

To represent the dinocyst assemblages, we used a diagram of relative abundances by region and a shaded plot. The shaded plot input data corresponded to the square root of dinocyst relative abundances to minimize the effects of dominant species in each sample following Mudie et al. (2017).

We counted the total pollen and spores in the samples as one group. Statistical analyses were performed on fifty-two samples, and the Shannon–Wiener index (H') was used to estimate the diversity and evenness of taxa across space (Table 5). Similar to Marret et al. (2001), we used a minimum of 50 specimens as a cut-off value to include a sample in the statistical analyses. We acknowledge that samples with low counts (less than 100 dinocysts) might underestimate the diversity and artificially over or underrepresent some species in a particular sample (Table 4) and thus bias the diversity index estimation. Nevertheless, a 50 dinocyst cut-off value has proved to represent the main dinocyst species in areas with relative low cyst densities (Marret et al., 2001).

To evaluate environmental heterogeneities within SC and ETP localities, a non-metric multidimensional scaling (NMDS) was applied to the SST, SSS, and [PO₄] and [NO₃] data using the Bray-Curtis index as the comparison metric (Legendre and Gallagher, 2001). To assess for relationships between environmental parameters and dinocyst taxa with abundances higher than 1%, a redundancy analysis was performed (RDA; Legendre and Gallagher, 2001). Cyst raw counts were transformed to Hellinger distances to enable common taxa to be weighted equally to rare species in the statistical analysis (Legendre and Gallagher, 2001). Scaling and transformation of environmental parameters was performed prior to RDA (Legendre and Gallagher, 2001). The relative abundances of the six more dominant dinocyst taxa were overlaid on the environmental NMDS values using bubble plots to show the geographic distribution and relative abundance of these taxa in the multivariate space (Mudie et al., 2017).

Dinocysts were grouped according to nutritional strategies into heterotrophs and autotrophs following Schnepf and Elbrächter (1992)

8

Raw counts of organic-walled dinoflagellate cysts reported from surface samples from the southern Caribbean (SC) and the eastern tropical Pacific (ETP). *Bitectatodinium spongioum*. (BSPO), *Brigantedinium spp*. (BSPP), Cysts of *Archaeperidinium bailongense* (ABAI), Cysts of *Archaeperidinium constrictum* (ACON), Cysts of *Polykrikos schwartzii* (PSCH), cysts of *Protoceratium reticulatum* (PRET), *Echinidinium aculeatum* (EACU), *Echinidinium granulatum* (EGRA), *Echinidinium spp*. (ESPP), *Impagidinium aculeatum* (IACU), *Lingulodinium machaerophorum* (LMAC), *Lejeunecysta* sp. (LESP), *Nematosphaeropsis lemniscata* (NLEM), *Operculodinium israelianum* (OISR), *Selenopemphix nephroides* (SNEP), *Selenopemphix quanta* (SQUA), *Spiniferites spp*. (SSPP), *Stelladinium bifurcatum* (SBIF), *Tuberculodinium vancampoae* (TVAN), *Votadinium spinosum* (VSPI), *Votadinium sp*. (VSPP).

CUEVAS 1 120 0 0 2 0 0 0 0 0 0 0 0 6 0 1 0 0 0 130 L.VARADERO 0 38 1 0 0 4 0 1 1 0 0 0 0 1 5 1 0 0 1 4 58 RC13-100 148 0 0 1 0 0 0 0 0 0 27 0 3 1 0 0 0 180 VM28-115 2 65 0 0 7 3 0 0 0 0 1 0
LVARADERO 0 38 1 0 0 4 0 1 1 0 0 0 1 5 1 0 0 1 4 58 RC13-100 148 0 0 1 0 0 0 0 0 0 0 27 0 3 1 0 0 180 VM28-115 2 65 0 0 0 7 3 0 0 0 0 0 0 0 1 0 0 0 0 7 3 0
RC13-100 148 0 0 1 0
VM28-115 2 65 0 0 7 3 0 0 0 1 0 0 1 0 0 0 7 7 TRI 49-19 I 17 69 0 2 0 6 10 2 0 0 0 0 0 4 4 2 0 0 0 116 NECOCLÍ 10 71 0 1 0 8 11 0 0 0 0 0 1 3 2 0 0 0 107 ZAPSURRO 10 34 0 0 0 0 0 0 0 0 0 10 1 0 0 0 0 1 3 2 0
TRI 49-19 I 17 69 0 2 0 6 10 2 0 0 0 0 4 4 2 0 0 0 16 NECOCLÍ 10 71 0 1 0 8 11 0 0 0 0 0 1 3 2 0 0 0 107 ZAPSURRO 10 34 0 0 7 2 0 0 0 0 0 19 0 4 0 0 0 77 RC13-169 0 49 0 0 0 0 0 0 0 0 10 1 0 0 0 77 W12-110 0 0 0 0 0 0 0 0 0 0 0 0 0 0 77 W12-110 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
NECOCLÍ 10 71 0 1 0 8 11 0 0 0 0 0 1 3 2 0 0 0 107 ZAPSURRO 10 34 0 0 7 2 0 0 1 0 0 1 3 2 0 0 0 107 ZAPSURRO 10 34 0 0 7 2 0 0 1 0 0 0 19 0 4 0 0 0 77 RC13-169 0 49 0 0 0 1 5 0 0 0 0 0 10 1 0 0 0 67 VM12-110 0<
ZAPSURRO 10 34 0 0 7 2 0 0 1 0 0 0 19 0 4 0 0 0 77 RC13-169 0 49 0 0 0 1 0 0 0 0 19 0 4 0 0 0 77 VM12-110 0 49 0 0 0 1 0 0 0 0 0 10 1 0 0 0 67 VM12-110 0 <th< td=""></th<>
RC13-169 0 49 0 0 1 5 0 0 0 0 0 1 0 1 0 0 67 VM12-110 0
VM12-110 73
RCU9-U52 U 54 O O O 2 O O O O O O O O O O O O O O O
VM20-012 1 58 0 0 0 1 0 0 0 0 0 0 0 0 0 2 0 0 0 0 0 0
RC11-239 0 46 0 0 0 1 0 0 0 0 1 0 0 0 2 0 0 0 0 0 0 50
PALOMINO4 3 74 0 0 0 10 0 1 0 0 1 0 0 2 0 11 0 1 0 0 103
POPOYA13 2 187 0 0 0 1 0 0 0 0 0 0 0 0 0 4 0 0 0 0 0 0
VM28-07 0 38 0 0 0 6 0 0 4 0 0 0 0 2 0 0 1 0 0 0 5 5
RC09-045 0 59 0 0 0 3 0 0 0 0 0 0 0 0 0 0 0 0 4 0 0 0 0
$\begin{array}{cccccccccccccccccccccccccccccccccccc$
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FORMS 30 104 1 0 2 0 11 4 8 0 0 0 2 0 11 0 6 0 1 0 1 108
FORO7 17 58 1 0 3 0 10 1 0 0 0 0 3 6 0 2 0 1 0 1 0 0 111
FOROI2 3 68 1 0 0 0 1 1 0 0 0 0 0 0 4 1 1 0 0 0 0 80
FORM12 5 6 1 0 1 0 7 2 0 0 1 0 0 0 2 0 0 2 0 0 138
FGORDIA 4 42 0 0 0 0 4 1 0 0 0 0 1 0 2 0 0 0 0 0 54
EGORO16 30 90 0 0 1 0 13 2 0 0 2 0 0 0 2 1 0 0 0 0 1 142
EGOR017 17 91 0 0 2 6 9 5 0 0 1 0 0 4 2 0 0 0 0 1 0 138
EGOR09 57 133 1 1 3 0 8 3 0 0 0 1 0 1 2 0 0 0 0 0 210
EGORO20 57 269 2 0 4 2 12 3 2 0 0 0 0 1 2 0 1 0 0 0 2 359
MERA2 16 175 1 0 2 0 8 3 0 0 0 0 0 1 3 0 0 0 0 0 209
MERA3 35 120 1 0 2 0 6 5 0 0 1 0 0 2 3 0 0 0 0 0 175
MERA5 26 89 0 0 2 1 2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
MERA8 10 178 2 0 1 0 2 1 0 0 0 0 0 0 4 1 0 0 0 0 199
MERA13 11 63 3 0 2 0 4 5 0 0 0 0 0 0 3 0 1 0 0 0 92
VM20-015 12 104 1 0 1 0 8 1 0 0 0 0 0 0 3 0 2 0 0 0 132
VM15-011 11 80 1 0 2 2 5 0 0 0 0 0 0 3 6 0 2 0 0 0 112
RC10-253 43 91 0 0 1 1 20 0 0 0 1 0 2 0 3 0 5 0 0 0 167
RC9-065 38 172 0 0 7 0 21 0 1 0 0 0 1 0 4 0 4 0 1 0 250
VM21-216 62 70 0 0 2 0 2 1 0 0 1 0 1 1 0 0 2 0 0 0 142
KAMA3 27 72 0 1 1 5 6 1 0 1 0 0 0 0 2 7 2 0 0 1 0 126
KAMA14 27 271 0 3 2 23 1 0 1 0 10 3 6 0 0 3 348
KAMA24 20 240 0 1 15 17 2 1 1 0 0 9 3 5 0 0 2 0 319
KAMA1 9 90 0 5 15 8 0 0 0 0 1 0 2 0 0 130
KAMA2 28 126 0 0 2 16 3 2 2 0 1 0 4 1 2 0 0 189
KNR176-2 JPC9 32 185 0 2 1 3 17 4 0 0 0 0 0 0 4 1 4 0 0 1 0 254
KNR176-2-JPC32 22 168 1 2 1 2 10 1 0 1 0 0 2 0 3 1 10 0 0 0 224
KNR176-2-MC4 41 98 0 0 0 0 5 1 0 0 0 0 0 0 0 0 1 0 0 0 0 146

Pollen and spores count (p + s count), dinocyst count, *Lycopodium* sp. count (*Lycopodium* count), concentration of pollen and spores per gram of dry sediment (p + s/g), pollen and spores per cm² per year ($p + s/cm^2/yr$), pollen and spores per m² per year ($p + s/m^2/yr$), organic-walled dinoflagellate cyst concentration per gram of dry sediment (dinocysts/g), dinoflagellate cyst per cm² per year (dinocysts/cm²/yr), dinoflagellate cyst per m² per year (dinocysts/cm²/yr), Shannon–Wiener index (*H*') and Heterotrophic/Autotrophic ratio (H/A) for all studied samples.

Locality	$\mathbf{p} + \mathbf{s}$	dinocyst	Lycopodium	p + s/g	$p + s/cm^2/$	$p + s/m^2/$	dinocyst/	dinocyst/cm ² /	dinocyst/m²/	H′	H/
	count	count	count		yr	yr	g	yr	yr		A
CUEVAS	259	130	1059	4012	2.9	28,554	3253	1.4	14,332	0.35	97
L.VARADERO	312	58	1120	4570	2.3	23,400	1585	0.4	4350	1.21	90
RC13-100	129	180	2705	1565	0.3	3235	3526	0.5	4514	0.76	16
VM28-115	60	79	2140	920	2.6	26,250	1956	3.5	34,562	0.49	86
TRI 49–19 I	95	116	1636	1905	0.2	2382	3757	0.3	2909	1.3	78
NECOCLÍ	232	107	2090	2958	2.2	21,750	3341	1.0	10,031	1.17	81
ZAPSURRO	200	77	2165	2461	2.8	28,350	2321	1.1	10,914	1.64	71
RC13-169	42	67	2020	682	1.9	19,404	1758	3.1	30,954	1.07	82
VM12-110	175	73	2125	2702	8.1	80,850	1820	3.4	33,726	0.19	86
RC09-052	25	57	2075	395	1.2	11,550	1456	2.6	26,334	0.43	95
VM20-012	62	62	2558	795	0.6	6138	1284	0.6	6138	0.35	97
RC11-239	66	50	2635	822	0.7	6534	1006	0.5	4950	0.36	96
PALOMINO4	239	103	2041	4009	11.0	11,0418	2563	4.8	47,586	0.84	75
POPOYA13	102	194	2132	1638	4.7	47,124	4621	9.0	89,628	0.32	98
VM28-07	55	55	4471	404	0.5	5156	604	0.5	5156	0.7	80
RC09-045	31	66	2135	476	1.4	14,322	1638	3.0	30,492	0.49	89
VM24-031	17	58	2232	250	0.2	1683	1377	0.6	5742	0.55	100
VM24-032	63	71	2751	751	0.6	5906	1368	0.7	6656	0.26	93
VM19-017	46	74	2917	517	2.1	21,252	1235	3.4	34,188	0.36	92
POPOYA9	22	70	1255	600	1.0	10,164	2590	3.2	32,340	0.72	91
RINCÓN MAR3	147	54	2550	1529	1.5	14,553	1388	0.5	5346	0.59	91
RC13-148	61	51	3910	512	0.6	5718	691	0.5	4781	0.48	92
C.DIQUE	374	57	3415	3593	3.7	37,026	869	0.6	5643	0.88	56
BERRUGAS16	104	54	2335	1187	1.0	10,296	1509	0.5	5346	1.23	80
EGORO1	393	111	585	22,766	1.4	13,755	2192	0.4	3535	1.46	78
EGORO5	454	80	1268	12,134	1.7	16,684	1942	0.7	7276	1.56	72
EGORO7	297	138	1045	9631	1.1	10,914	1510	0.4	4079	1.5	79
EGORO12	682	54	902	25,623	2.4	23,870	1166	0.3	2800	0.64	95
EGORO15	507	142	2915	5894	1.9	18,632	651	0.5	5071	1.14	79
EGORO14	240	138	1032	7881	0.9	8820	721	0.2	1984	0.94	91
EGORO16	304	210	1126	9149	1.4	14,470	1754	0.7	6759	1.19	77
EGORO17	380	359	1720	7487	1.4	13,965	1007	0.5	5071	1.06	80
EGORO9	450	209	1012	15,069	1.6	15,750	2699	0.7	7350	1.15	72
EGORO20	290	175	902	10,895	1.1	10,657	5162	1.3	13,193	0.88	82
MERA2	534	120	1165	7138	1.1	10,680	5055	0.4	4180	0.75	92
MERA3	420	199	1140	5738	0.9	8820	4235	0.4	3675	0.98	78
MERA5	333	92	1280	8103	0.7	6660	5277	0.2	2400	0.69	78
MERA8	453	132	2960	4767	0.9	9060	3790	0.4	3980	0.51	95
MERA13	279	112	4172	2083	0.6	5580	1231	0.2	1840	1.21	87
VM20-015	87	167	2380	1280	2.3	22,837	2836	3.5	34,650	0.84	89
VM15-011	194	250	25/5	2639	0.4	4074	2101	0.2	2352	0.95	84
RC10-253	189	142	2085	3175	0.4	3969	4000	0.4	3507	1.28	69
RC9-065	241	126	2395	3524	0.5	5061	5161	0.5	5250	1.07	82
VM21-216	474	348	3800	4369	1.0	9954	1842	0.3	2982	1	53
KAMA3	191	319	2470	2708	0.4	3820	2592	0.3	2520	1.18	72
KAMA14	292	130	2225	4596	0.6	6132	/98/	0.7	/308	1.07	89
KAWA24 VAMA1	228	109	2097	3808 2025	0.5	4300	7740	0.0	2250	1	8/ 70
KAWAD	200	204 224	2329	2020	0.0	4700	2/49	0.3	323U 479E	1.14	/9 00
KANDIAZ	100	224 146	2100	3020 1720	0.5	3760	3490	0.5	5080	1.33	00 Q⊏
KNR170-2 JPC9	340	120	2612	1/34	0.4	6800	3421 2242	0.3	4480	1.05	63
IDC22	340	130	2012	2200	0.7	0000	2242	0.4	140V	1.15	03
JPG32 VND176-0-MC4	410	EQ	2406	2001	0.8	9240	1450	0.2	2020	0.97	70
KINK1/0-2-WIC4	412	30	2490	2091	0.0	0240	1432	0.5	2920	0.8/	12

and Limoges et al. (2013) (Tables 5 and 6). The heterotrophic/auto-trophic (H/A) % was calculated as the number of heterotrophic dinocysts found in the sample divided by the total number of dinocysts in a sample multiplied by 100.

All statistical analyses were performed using the vegan R package (Oksanen et al., 2014; R core Team, 2015).

4. Results and discussion

4.1. Dinocyst, pollen, and spore concentrations and accumulation rates

Dinocyst concentrations in the SC range from 604 to 4621 cysts per gram of dry sediment (mean = 1979 \pm 1688), whereas in the ETP, the

absolute concentration of cysts per gram of dry sediment ranges from 651 to 7987 (mean = 3105 ± 1956) (Table 5, Figs. 5a, 6). The lower concentration of dinocysts in the SC compared to the ETP can be explained by: 1) the predominance of silt and sand grains in the SC (i.e., coarser grain size) compared to the more clayey substrate of the ETP (Narale et al., 2013); 2) differences in primary productivity, which are considerably higher in the ETP (e.g., Lewis et al., 1990; Dale, 2001; Radi and de Vernal, 2004; Sangiorgi and Donders, 2004; Zonneveld et al., 2010; Hardy et al., 2016; Bringué et al., 2019; de Vernal et al., 2020); and/or 3) differences in the accumulation rate of sediments between the SC and the ETP.

Pollen and spores are very abundant in the studied samples, especially in the ETP samples (Fig. 5b, 6). Pollen and spore concentrations in

Dinocyst species recorded in the southern Caribbean (SC) Sea and the eastern tropical Pacific Ocean (ETP), their corresponding plate and map numbers, trophic habit of the corresponding motile stage. A = Autotrophic, H = heterotrophic as determined by Schnepf and Elbrächter (1992) and Limoges et al., (2010), (%): Relative abundance of dinocyst taxa for the fifty-two samples analyzed in this work (Code: Abbreviation name, Fig. 9). Species in bold correspond to the most representative taxa (relative abundance >1%). * = taxon is reported but not mapped.

Cyst name	Plate	Мар	Trophism	Motile stage name (Head, 1996)	%	Code
Bitectatodinium spongium	I	Fig. 10	Autotrophic	Unknown	14	BSPO
Brigantedinium spp.	I	Fig. 11	Heterotrophic	Protoperidinium sp.	70	BSPP
Brigantedinium cariacoense*	I	No map	Heterotrophic	Protoperidinium avellanum		BCAR
Brigantedinium simplex*	I	No map	Heterotrophic	Protoperidinium conicoides		BSIM
Cyst of Archaeperidinium bailongense*	I	No map	Heterotrophic	Archaeperidinium bailongense	0.3	ABAI
Cyst of Archaeperidinium constrictum*	I	No map	Heterotrophic	Archaeperidinium constrictum	0.2	ACON
Cyst of Polykrikos schwartzii*	I	No map	Heterotrophic	Polykrikos kofoidii	0.7	PSCH
Cyst of Protoceratium reticulatum	I	Fig. 12	Autotrophic	Protoceratium reticulatum	2	PRET
Echinidinium aculeatum	I	Fig. 13	Heterotrophic	Unknown	5	EACU
Echinidinium granulatum*	I	No map	Heterotrophic	Unknown	0.9	EGRA
Echinidinium spp.*	I	No map	Heterotrophic	Unknown	0.5	ESPP
Impagidinium aculeatum*	I	No map	Autotrophic	Gonyaulax sp. indet	0.1	IACU
Lingulodinium machaerophorum*	II	No map	Autotrophic	Lingulodinium polyedra	0.2	LMAC
Lejeunecysta sp.*	II	No map	Heterotrophic	Unknown	0.01	LESP
Nematosphaeropsis lemniscata *	II	No map	Autotrophic	Gonyaulax spinifera complex	0.3	NLEM
Operculodinium israelianum*	II	No map	Autotrophic	Protoceratium sp.	0.3	OISR
Selenopemphix nephroides	II	Fig. 14	Heterotrophic	Protoperidinium subinerme	4	SNEP
Selenopemphix quanta*	II	No map	Heterotrophic	Protoperidinium conicum	0.5	SQUA
Spiniferites spp.	II	Fig. 15	Autotrophic	Gonayaulax spp.	2	SSPP
Spiniferites bentorii*	II	No map	Autotrophic	Gonyaulax digitale		SBEN
Spiniferites mirabilis*	II	No map	Autotrophic	Gonyaulax spinifera complex		SMIR
Spiniferites pachydermus*	II	No map	Autotrophic	Gonyaulax spinifera complex.		SPAC
Spiniferites ramosus*	II	No map	Autotrophic	Gonyaulax spinifera complex.		SRAM
Stelladinium bifurcatum*	II	No map	Heterotrophic	Unknown	0.06	SBIF
Tuberculodinium vancampoae*	II	No map	Autotrophic	Pyrophacus steinii	0.08	TVAN
Votadinium spinosum*	II	No map	Heterotrophic	Protoperidinium claudicans	0.1	VSPI
Votadinium sp.*	-	No map	Heterotrophic	Protoperidinium sp.	0.09	VSPP

the SC range from 250 to 4570 palynomorphs per gram of dry sediment (mean = 1635 ± 5172). In the ETP, pollen and spore concentrations range from 1280 to 25,623 (mean = 6936 ± 5963) (Table 5, Figs. 5b, 6). The higher concentration of terrestrial palynomorphs in the ETP can be explained by: 1) the proximity of many samples to the coast; and 2) the general morphology of the rivers that flow into the ETP that are characterized by their shortness, steepness, and the absence of alluvial floodplains. All together the fluvial morphology of rivers draining the ETP makes it easier for terrestrial palynomorphs to be readily deposited in the basin without being entrapped in proximal areas before reaching the ocean (Restrepo and Kjerfve, 2004).

4.2. Dinocyst assemblages

A total of twenty dinocyst taxa were found in the SC and the ETP (total dinocyst diversity is of 24, but we expose 20 taxa since some groups have been made) (Table 6; Plates I-II). The assemblages consisted of the autotrophic species *Bitectatodinium spongium* and *Spiniferites* spp., and the heterotrophic *Brigantedinium* spp. (the most dominant cyst), and the cysts of *Protoceratium reticulatum, Echinidinium aculeatum,* and *Selenopemphix nephroides*. These species represented 94% of the total dinocyst assemblages (Table 6; see Section 5 for a detailed discussion of these most abundant dinocysts and their environmental preferences). The dinocyst species *Archaeperidinium constrictum, Echinidinium* spp. (except for *E. aculeatum*), *Stelladinium bifurcatum, Votadinium* sp., *Votadinium spinosum*, and *Lejeunecysta* sp. were found in very low numbers or recorded as single occurrences (Fig. 6; Table 4). The relative abundances of these taxa are presented in Figs. 6 and 7, but they will not be further discussed in this paper due to their scarce presence in the assemblages.

4.3. Heterotrophic-autotrophic ratio as an indication of productivity levels

A high heterotrophic/autotrophic (H/A) ratio value has been considered as an indication of high productivity possibly due to high nutrient availability in marine waters (Lewis et al., 1990; Radi and de

Vernal, 2004; Sangiorgi and Donders, 2004; Verleye and Louwye, 2010; Zonneveld et al., 2010; Hardy et al., 2016). Based on the modern environmental conditions of the study area, a higher H/A ratio is expected for the ETP (Fig. 6). However, no significant differences in the H/A ratio were observed for the two basins. Conversely and, consistent with the contrasting environmental patterns observed in the ETP and the SC, Huguet et al. (2019) studied the biomarker content of the same surface sediments used in this study and found that the ETP has much higher nutrient levels compared to the more oligotrophic SC. These findings imply that using the H/A ratio in the study area without other proxies can lead to misleading conclusions regarding the nutrient levels. Moreover, if oxygen conditions in bottom water have selectively preserved dinocysts in the samples, the use of the H/A ratio to infer productivity is biased. It is well known that Protoperidiniod cysts (mostly heterotrophic) are more sensitive than G cysts (mostly autotrophic) to oxidation (e.g., Zonneveld et al., 2007, 2010). In locations and/or in down-core records where bottom water oxygen is known to have varied, the use of such a ratio is far from ideal. Such bias can be excluded in our samples (see paragraph 4.4).

4.4. Diversity of dinocyst assemblages

The Shannon-Wiener index (H') was higher in the ETP (mean $H' = 1.05 \pm 0.24$) than in the SC (mean $H' = 0.69 \pm 0.39$) (Fig. 8a; Table 5). Dinocysts, in general, are more abundant and diverse in areas where seasonality and environmental conditions in the water column strongly vary (Dale, 1983). Although distance to the shoreline has an important effect on the dinocyst diversity (Sluijs et al., 2008), the lack of correlation between this parameter and diversity in the study areas is consistent with the higher climatic variability present in the ETP as the main driver of a higher dinocyst diversity compared to the SC.

When samples RC13–100 and VM12–110 from the SC are excluded (*Brigantedinum* spp. concentrations are zero), a significant negative correlation ($R^2 = -0.836$; p < 0.001; Fig. 8b) was found between H' and the relative abundance of *Brigantedinim* spp. A preservation bias can be



Fig. 5. Geographic distribution of: a- Dinocyst concentration (dinocyst/g), b- concentration of terrestrial palynomorphs (pollen + spores/g), c- flux of dinocysts (Log (dinocyst/ m^2 /yr.)), d- Log of terrestrial palynomorphs accumulation (Log (pollen + spores/ m^2 /yr.)) in the southern Caribbean (SC), and eastern tropical Pacific (ETP).

ruled out as *Brigantedinim* spp. is more prone to be oxidized in well-oxygenated environments and therefore be underrepresented in surface sediments when assemblages have been affected by post-depositional processes (Zonneveld et al., 2007). This negative relation-ship can therefore be explained by the dominance of the *Brigantedinium* group since the H' index is sensitive not only to the species richness but also to the evenness of the dinocyst assemblage.

4.5. Environmental parameters related to dinocyst assemblages

Environmental differences between the SC and ETP areas are evident from the different values of the annual mean SST (27.88 °C in the SC and 26.64 °C in the ETP), mean yearly SSS (35.34 psu in the SC and 30.92 psu in the ETP), annual mean [PO₄] (0.035 μ mol/L in the SC and 0.032 μ mol/L in the ETP), and seasonal [NO₃]_JFM (0.1 μ mol/L in the SC and 1.28 μ mol/L in the ETP) (Fig. 3).

The environmental parameters associated with NMDS Axis 1 were $[NO_3]_JJA$, $[NO_3]_JFM$, and $[PO_4]_JJA$. For Axis 2, $[NO_3]_JJA$ and WD drive most of the variation (Table 7). The NMDS plot grouped sample points into two clusters that are clearly related to the geographic location of the samples (Fig. 9a). SC data points are plotted on the left-hand side of the first NMDS axis, whereas all the ETP sediment samples are plotted on the right-hand side of the first axis (Fig. 9a, Table 7).

The SC samples are scattered along axis 2 and can be further subdivided into two subgroups (SC-1 and SC-2) in the NMDS plot (Fig. 9a). The first subgroup (SC-1) is represented by seventeen data points containing the samples collected from the deepest parts of the SC region and therefore these samples have very limited riverine influence. The SC-2 subgroup, on the other hand, included seven samples collected in areas under the influence of the Atrato and Magdalena Rivers or in their embayments (i.e. Berrugas 16, Rincon del Mar, Necocli, and Zapsurro).

4.6. Environmental data as drivers of the most common species

Seventy-seven per cent of the total variance in the data is explained by the environmental data and the relative abundance of dinocysts (Fig. 9; Axis 1 = 57% and Axis 2 = 20%). Based on the RDA, seasonal and annual SSS [PO₄], and [NO₃] are the most important variables controlling the differences in the dinocyst abundances in the SC and the ETP (Fig. 9b, Table 8). SSS Annual and in JJA were negatively related to RDA axis 1. Brigantedinium spp., the cysts of P. reticulatum, S. nephroides, and Spiniferites spp. plot towards the center of the first axis, while B. spongium and E. aculeatum plotted on the right-hand side of the RDA reflecting a first order control of the SSS values on the abundance of these species (Fig. 9b). SSS Annual and in JJA are negatively related to RDA Axis 1. Axis 1 is defined by negative RDA scores for Brigantedinium spp., the cysts of P. reticulatum, S. nephroides, and Spiniferites spp., and by positive scores for B. spongium and E. aculeatum (Fig. 9b, Table 8). In contrast, Brigantedinium spp. is positively related to RDA Axis 2, plotted in a more negative position compared to the other species that dominate the assemblages (Fig. 9b, Table 8).



Fig. 6. From left to right: Pollen grains and spore concentrations per gram of dry sediment (pollen +spores/g). Dinocyst concentration per gram dry sediment (cyst/g). Heterotrophic/Autotrophic ratio (H/A). Relative abundances of dominant and rare dinocyst taxa and Shannon–Wiener index (H'). The grey rectangle highlights the southern Caribbean; the white background represents the eastern tropical Pacific.

5. Occurrences and environmental requirements of the most abundant dinocyst taxa

dominated by *B. spongium* were similarly associated with upwelling zones.

The following section presents a discussion of the distribution and abundance patterns of the dominant taxa in the dinocyst assemblages and their preferred environmental conditions in the SC and ETP zones.

5.1. Bitectatodinium spongium Zonneveld, KAF and Jurkschat et al., (1999), Figs. 10, 16a, Plate I, 1–2

Distribution: *Bitectatodiniumspongium* was found to be a common cyst in the ETP and a minor component of the dinocyst assemblages in the SC. This species was the second most abundant taxon in the studied samples representing 14% of the SC and ETP cyst assemblage. The highest abundances of the cyst were found in the ETP associated with upwelling zones (Figs. 10a). The low abundance of cysts found in modern sediments of the SC zone are at odds with high-production rates of the cyst that have been reported from sediment traps in the Cariaco Basin (Bringué et al., 2019). The highest abundances of *B. spongium* in the study area were observed at annual temperatures above 27 °C and salinity of 28 psu and 36 psu. The highest values of the cyst in the study area are positively correlated with [PO₄] and [NO₃]. The preferred SST, SSS, [PO₄], and [NO₃] values reported here for *B. spongium* are in good agreement with previous environmental ranges reported for the species (Zonneveld et al., 2013).

On a global scale, the highest abundances of *B. spongium* have been found in nutrient-rich waters and upwelling zones (Zonneveld and Jurkschat, 1999; Zonneveld and Pospelova, 2015). For the tropics, Bringué et al. (2019) found the highest abundances of *B. spongium* following seasonal episodes of intense upwelling in the Cariaco Basin. Also, Limoges et al. (2010) and Vásquez-Bedoya et al. (2008) found *B. spongium* along the southwestern coast of Mexico and in the Gulf of Tehuantepec areas with seasonal upwelling. In the SC and the ETP, areas

5.2. Brigantedinium spp. Figs. 11, 16b; Plate I, 3-4

Morphological remark: *Brigantedinium simplex*, *Brigantedinium cariacoense*, and other undetermined species of the genus *Brigantedinium* were grouped as *Brigantedinium* spp.

Distribution: *Brigantedinium* spp. were the most abundant species of cysts in both the ETP and the SC. The abundances of *Brigantedinium* spp. increased when [NO₃] were at the lowest annual average (0.1 µmol/L). This group of species represents 70% of the total number of cysts found in the ETP and the SC. *Brigantedinium* spp. were found through a wide range of water depths (mean = 1326 m, sd = 1319, min = 1 m, max = 4649 m). The highest quantities of *Brigantedinium* spp. were found when annual SST > 26 °C. In sediment traps of the Cariaco Basin, *Brigantedinium* spp. is very abundant (ca. 59%) in modern assemblages with a maximal annual cyst production from December to May coinciding with active upwelling (Bringué et al., 2019).

On a global scale, the highest abundances of *Brigantedinium* spp. have been found in upwelling zones, with increasing abundance when upwelling is active and decreasing abundance during upwelling relaxation (de Vernal et al., 2020). Limoges et al. (2010) and Vásquez-Bedoya et al. (2008) recorded a widespread distribution of *Brigantedinium* spp. along the Mexican coast and the lowest abundances in nearshore environments. Based on the global distribution of this taxon, *Brigantedinium* spp. can be considered cosmopolitan. Its presence has been reported from the tropics to regions permanently covered by sea ice and in water depths that range from coastal areas to the deep ocean (de Vernal et al., 1997; Marret and Zonneveld, 2003; Pospelova et al., 2008, 2010; Zonneveld et al., 2013).



Plate I. 1–2: Bitectatodinium spongium; 3–4: Brigantedinium spp.; 5: Archaeperidinium bailongense; 6: cysts of Archaeperidinium constrictum; 7: cysts of Polykrikos schwartzii; 8: cysts of Protoceratium reticulatum; 9: Echinidinium aculeatum; 10: Echinidinium granulatum. 11: Echinidinium spp.; 12: Impagidinium aculeatum. Scale bar = 10 μm.





Plate II. 1: Lejeunecysta sp.; 2: Lingulodinium machaerophorum; 3: Nematosphaeropsis lemniscata; 4: Operculodinium israelianum; 5: Selenopemphix nephroides; 6: Selenopemphix quanta; 7–8: Spiniferites spp.; 9: Spiniferites spp.; 10: Stelladinium bifurcatum, 11: Tuberculodinium vancampoae; 12: Votadinium spinosum. Scale bar = 10 µm.



Fig. 7. Shade plot of square root transformed dinocyst relative abundances for the southern Caribbean (SC) and eastern tropical Pacific (ETP). Highest abundances are shown in dark orange and lowest transformed relative abundances are shown in light orange.



Fig. 8. a- The Shannon-Wiener diversity index (*H'*) of dinocysts. The insert figure shows box plots for the southern Caribbean (red box) and the ETP (black box). b-Biplot of the Shannon-Wiener index (*H'*) and *Brigantedinium* spp. relative abundances.

5.3. Cysts of Protoceratium reticulatum (Claparède and Lachmann, 1859) Bűttschli, 1885, Figs. 12, 16c; Plate I, 8. Morphological remark: Following Paez-Reyes and Head (2013), cysts with similar morphology to Operculodinium centrocarpum sensu Wall and Dale, 1966 are referred to in this paper as cysts of Protoceratium reticulatum

Distribution: Cysts of *Protoceratium reticulatum* were more abundant in the SC than in the ETP, with the highest abundances in the SC being related to areas with high SSS values. This species represented 2% of the entire cyst assemblages of the SC and the ETP and the cysts of *Protoceratium reticulatum* were observed in sediments deposited over a wide range of water depths (mean of =1708 m, sd = 1698, min = 1 m max = 5040 m). The highest abundances of cysts of *Protoceratium reticulatum* were found where annual temperatures range from 27 $^{\circ}$ C to 29 $^{\circ}$ C and with SSS at 35 psu and above 36.8 psu.

The cysts of *Protoceratium reticulatum* recorded in the SC and the ETP show a morphological variation expressed as changes in the length of the processes, which ranged from 2.7 μ m to 9.4 μ m with an average length of 5.53 μ m and a standard deviation of 1.56 μ m for twenty-one specimens. Mertens et al. (2011) and Verleye et al. (2012) demonstrated that the length of the processes in this species is controlled mainly by salinity and temperature. However, its genetic variation and the presence of pseudo-cryptic species, means that its use as an environmental tracer is not straightforward (Mertens et al., 2011).

Scores for axes 1 and 2, according to the non-metric multidimensional scaling (NMDS) of environmental parameters for the southern Caribbean (SC) and eastern tropical Pacific (ETP). Bold numbers correspond to NMDS-scores of most explanatory environmental variables.

Environmental parameter	NMDS1	NMDS2
SST_A	-0.06089072	-0.03252157
SST_JJA	-0.05773889	-0.03205266
SST-JFM	-0.05242172	-0.03229783
SSS_A	-0.07261274	-0.02336757
SSS_JJA	-0.06867755	-0.02783331
SSS_JFM	-0.07378798	-0.02929745
PO4_A	0.28471409	-0.02362933
PO4_JJA	0.37472981	-0.03265386
PO4_JFM	0.04031442	-0.18018941
NO3_A	0.26765733	-0.11289742
NO3_JJA	-0.67271222	0.52257962
NO3_JFM	0.53042314	-0.14296472
WD	-0.00488358	0.32575543
DC	-0.05720408	0.29670788



Fig. 9. a- Non-metric multidimensional scaling (NMDS) for environmental parameters based on Bray-Curtis similarities for the southern Caribbean (SC) (red dots) and eastern tropical Pacific (ETP) (black dots); b- Redundancy analysis (RDA) for more abundant species and environmental parameters according to axes 1 and 2 of the variance. Species abbreviations: *Bitectatodinium spongium* (BSPO), *Brigantedinium spp.* (BSPP), cysts of *Protoceratium reticulatum* (PRET), *Echinidinium aculeatum* (EACU), *Selenopemphix nephroides* (SNEP), *Spiniferites* spp. (SSPP).

Table 8

Scores for axes 1 and 2, according to the redundancy analysis (RDA) of the correlation coefficient between the environmental parameters and the scores for the more representative dinocysts. Parameter abbreviations are: sea surface temperature (SST), sea surface salinity (SSS), phosphates concentration ([PO₄]), nitrates concentration ([NO₃]), annual (A), June–July–August (JJA), January–February–, March (JFM). Water depth (WD) and distance to the coast (DC) for the southern Caribbean (SC) and eastern tropical Pacific (ETP). Bold numbers correspond to RDA-scores of most explanatory variables and species.

Environmental parameter	RDA1	RDA2
SST_A	-0.36404	0.070137
SST_JJA	-0.45797	0.095381
SST_JFM	0.14178	0.225984
SSS_A	-0.77344	0.107213
SSS_JJA	-0.85130	-0.007678
SSS_JFM	-0.88660	0.007302
PO4_A	0.82990	0.010562
PO4_JJA	0.72494	-0.113969
PO4_JFM	0.04251	-0.009680
NO3_A	0.55406	0.096139
NO3_JJA	-0.60477	-0.061774
NO3_JFM	0.24242	-0.064725
WD	-0.23448	0.490224
DC	-0.21446	0.291055
Dinocyst taxa		
Brigantedinium spp.	-0.03801832	-0.41565202
Bitectatodinium spongium	0.67531513	0.04883838
Echinidinium aculeatum	0.32477966	-0.09893145
Cysts of Protoceratium reticulatum	-0.24688309	-0.06643120
Selenopemphix nephroides	-0.17706919	0.22263322
Spiniferites spp.	-0.11956780	-0.05325828

5.4. Echinidinium aculeatumZonneveld, 1997* Figs.13, 16d; Plate I, 9. Morphological remark: * the name was not validly published in Zonneveld (1997) because the original description of the species based on a modern cyst did not include a latin diagnosis (see discussion in Head, 2002)

Distribution: *E. aculeatum* was a minor component of the assemblages from the SC but present in almost every sample from the ETP (Fig. 10a). *E. aculeatum* represented 4% of the entire cyst assemblages and was observed in sediments deposited over a wide range of water depths (mean of $=1171 \pm 1168$ m, min = 1 m, max = 4649 m). Although this taxon is only found in low-relative abundances in sediments from the SC compared to the ETP, the cyst has often been reported to be found in sediment traps and sediment cores from the Cariaco Basin with a strong association to upwelling (González et al., 2008; Bringué et al., 2019). Consistent with these observations, the highest abundances of *E. aculeatum* in the study area occurred in close proximity to upwelling zones (Fig. 13a). The peak cyst abundance of *E. aculeatum* was found in the ETP at 32 psu.

On a global scale, the highest abundances of *E. aculeatum* have been found in coastal regions of active upwelling and zones with increased nutrient availability and low-salinity variations (Zonneveld, 1997; Ribeiro and Amorim, 2008; Limoges et al., 2010; Pospelova and Kim, 2010; Zonneveld et al., 2013). Bringué et al. (2019) found that the highest production of *E. aculeatum* in the SC was associated with active but seasonal upwelling in the Cariaco Basin. In the southeast Pacific, *E. aculeatum* appeared to be the best indicator for the presence of yearround active upwelling zones (Verleye and Louwye, 2010). In the Gulf of Mexico, this taxon was observed in nutrient-rich areas and nearby urban centers with extensive industrial activity (Vásquez-Bedoya et al., 2008; Limoges et al., 2010, 2013).

5.5. Selenopemphix nephroides (Benedek 1972) Benedek et Sarjeant, 1981, Figs. 14, 16e; Plate II, 5

Distribution: *S. nephroides* was observed in almost all localities analyzed in the SC and the ETP but was generally found in low relative



Fig. 10. a- Geographic distribution of *Bitectatodinium spongium*, b- Distance to the coast, c- Water depth, surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), h- June–July–August (SST_JJA), l- January–February–March (SST_JFM). Sea-surface salinity (SSS) in psu: e- Annual (SSS_A), i- June–July–August (SSS_JJA), m- January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: f- Annual ([PO4]_A), j- June–July–August ([PO4]_JJA), n- January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: g- Annual ([NO3]_A), k- June–July–August ([NO3]_JJA), o- January–February–March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).



Fig. 11. a- Geographic distribution of *Brigantedinium* spp., b- Distance to the coast, c- Water depth, surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), h- June–July –August (SST_JJA), l- January–February –March (SST_JFM). Sea-surface salinity (SSS) in psu: e- Annual (SSS_A), i- June–July–August (SSS_JJA), m- January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: f- Annual ([PO4]_A), j- June–July–August ([PO4]_JJA), n- January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: g- Annual ([NO3]_A), k- June–July –August ([NO3]_JJA), o- January–February –March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).



Fig. 12. a- Geographic distribution of cysts of *Protoceratium reticulatum*, b- Distance to the coast, c- Water depth, surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), h- June–July–August (SST_JJA), l- January–February–March (SST_JFM). Sea-surface salinity (SSS) in psu: e- Annual (SSS_A), i- June–July–August (SSS_JJA), m- January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: f- Annual ([PO4]_A), j- June–July–August ([PO4]_JJA), n- January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: g- Annual ([NO3]_A), k- June–July –August ([NO3]_JJA), o- January–February–March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).



Fig. 13. a- Geographic distribution of *Echinidinium aculeatum*, b- Distance to the coast, c- Water depth, surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), h- June–July–August (SST_JJA), l- January–February–March (SST_JFM). Sea-surface salinity (SSS) in psu: e- Annual (SSS_A), i- June–July–August (SSS_JJA), m- January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: f- Annual ([PO4]_A), j- June–July–August ([PO4]_JJA), n- January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: g- Annual ([NO3]_A), k- June–July–August ([NO3]_JJA), o- January–February–March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).



Fig. 14. a- Geographic distribution of *Selenopemphix nephroides*, b- Distance to the coast, c- Water depth, surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), h- June–July–August (SST_JJA), l- January–February –March (SST_JFM). Sea-surface salinity (SSS) in psu: e- Annual (SSS_A), i- June–July–August (SSS_JJA), m- January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: f- Annual ([PO4]_A), j- June–July–August ([PO4]_JJA), n- January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: g- Annual ([NO3]_A), k- June–July–August ([NO3]_JJA), o- January–February–March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).



Fig. 15. a- Geographic distribution of *Spiniferites* spp., b- Distance to the coast, c- Water depth, surface-water conditions in the southern Caribbean (SC) and the eastern tropical Pacific (ETP). Sea-surface temperature (SST) in °C: a- Annual (SST_A), h- June_July_August (SST_JJA), l- January–February–March (SST_JFM). Sea-surface salinity (SSS) in psu: e- Annual (SSS_A), i- June_July_August (SSS_JJA), m- January–February–March (SSS_JFM). The sea-surface concentration of phosphates ([PO4]) in µmol/L: f- Annual ([PO4]_A), j- June_July_August ([PO4]_JJA), n- January–February–March ([PO4]_JFM). The sea-surface concentration of nitrates ([NO3]) in µmol/L: g- Annual ([NO3]_A), k- June_July_August ([NO3]_JJA), o- January–February–March ([NO3]_JFM). Values were downloaded from the World Ocean Atlas 2013 (Boyer et al., 2013).



Fig. 16. Bubble plot of relative abundances (Ra %) overlaying the NMDS plot of the environmental parameters (Fig. 8a) for the Southern Caribbean (SC) (red dots) and the eastern tropical Pacific (ETP) (black dots) of: a- *Bitectatodinium spongium* b- *Brigantedinium spp.*, c- cysts of *Protoceratium reticulatum*, d- *Echinidinium aculeatum*, e- *Selenopemphix nephroides* and f- *Spiniferites* spp.

abundances (less than 20% per sample) except for Zapsurro and VM12–110 from the SC where *S. nephroides* was found in relative abundances of up to (24% and 86% respectively).

The highest relative abundances of *S. nephroides* in the region occurred in shallow waters. In this environment, the cyst represents 4% of the dinocyst assemblage. In the RDA, the relative abundance of this dinocyst seems to be related to WD and DC. *S. nephroides* was observed in sediments deposited over a wide range of water depths (mean of =1324

 \pm 1512 m, min = 1 m, max = 5040 m) and occurs in waters influenced by coastal upwelling in the California Basin, Mexico, and Peru (Lewis et al., 1990; Vásquez-Bedoya et al., 2008; Limoges et al., 2010, 2013; Bringué et al., 2013). In the study area, the highest abundances of *S. nephroides* are reached in areas where the annual SST is higher than 27 °C and the annual SSS is between 34 and 36 psu.

On a global scale, the highest abundances of *S. nephroides* have been found in warm and high-productivity environments. The species is

classified as neritic with a strong affinity to low SSS values (Marret, 1994; Pospelova et al., 2008; Elshanawany et al., 2010; Candel et al., 2012; Zonneveld et al., 2013; Mudie et al., 2017). In Holocene sediments from the Cariaco Basin, the highest abundances of *S. nephroides* (ca. 5%) were associated with stronger trade winds and active upwelling zones during Heinrich-Stadials (González et al., 2008; Mertens et al., 2009).

5.6. Spiniferites spp. Figs. 15, 16f; Plate II, 7-9

Distribution: *Spiniferites* spp. was a common cyst in both the SC and ETP and represented 2% of the assemblages from both areas. The greatest cyst abundances were found to be associated with the highest SSS. *Spiniferites* spp. were observed in sediments deposited over a wide range of water depths (mean of $=1475 \pm 1591$ m, min = 4 m, max = 4674 m), and the highest abundances were found at annual temperatures of >26 °C and an SSS of between 34 and 36 psu.

On a global scale, the highest abundances of *Spiniferites* spp. have been found in waters with high SST values, weak upwelling zones, and fully marine conditions (Alves de Souza et al., 2008; Holzwarth et al., 2010; Candel et al., 2012; Bringué et al., 2013; Zonneveld et al., 2013). *Spiniferites* spp. is one of the most abundant species reported in the Holocene of the Cariaco Basin and Gulf of Mexico (González et al., 2008; Mertens et al., 2009; Limoges et al., 2014).

6. Conclusions

The study of fifty-two sediment samples from the SC and the ETP reveals that of the twenty taxa recorded, six taxa (Brigantedinium species, Bitectatodinium spongium, Echinidinium aculeatum, Selenopemphix nephroides, cysts of Protoceratium reticulatum, and some Spiniferites spp.) dominate the cyst assemblages from these basins. Bitectatodinium spongium, E. aculeatum, and E. granulatum are more abundant in the ETP, while the cysts of P. reticulatum and Spiniferites spp. -both autotrophic cysts- are predominant in the SC. A lower absolute abundance of cysts was found in the SC than the ETP. This pattern is explained either by the coarser grain size of the substrate in the SC compared to the ETP, the higher primary productivity in the ETP than in the SC, and/or the lower accumulation rate of sediments that characterizes the SC compared to the ETP. NMDS and RDA analyses reveal that the relative abundance, composition, and distribution of dinocysts in the SC and the ETP are better explained by the variation of phosphates ([PO₄] Annual), seasonal nitrates ([NO₃] JJA), and seasonal SSS (JJA, and JFM). A higher Shannon Index for the ETP is consistent with a larger environmental heterogeneity, and a lower dominance of Brigantedinium spp. Our results provide a modern regional scale baseline for ecological, biodiversity, and paleoceanographic reconstructions in the region.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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