

ORIGINAL ARTICLE

Quantifying Hospital-Acquired Carriage of Extended-Spectrum Beta-Lactamase-Producing Enterobacteriaceae Among Patients in Dutch Hospitals

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BACKGROUND. Extended-spectrum β -lactamase-producing Enterobacteriaceae (ESBL-E) are emerging worldwide. Contact precautions are recommended for known ESBL-E carriers to control the spread of ESBL-E within hospitals.

OBJECTIVE. This study quantified the acquisition of ESBL-E rectal carriage among patients in Dutch hospitals, given the application of contact precautions.

METHODS. Data were used from 2 cluster-randomized studies on isolation strategies for ESBL-E: (1) the SoM study, performed in 14 Dutch hospitals from 2011 through 2014 and (2) the R-GNOSIS study, for which data were limited to those collected in a Dutch hospital in 2014. Perianal cultures were obtained, either during ward-based prevalence surveys (SoM), or at admission and twice weekly thereafter (R-GNOSIS). In both studies, contact precautions were applied to all known ESBL-E carriers. Estimates for acquisition of ESBL-E were based on the results of admission and discharge cultures from patients hospitalized for more than 2 days (both studies) and a Markov chain Monte Carlo (MCMC) model, applied to all patients hospitalized (R-GNOSIS).

RESULTS. The absolute risk of acquisition of ESBL-E rectal carriage ranged from 2.4% to 2.9% with an ESBL-E acquisition rate of 2.8 to 3.8 acquisitions per 1,000 patient days. In addition, 28% of acquisitions were attributable to patient-dependent transmission, and the per-admission reproduction number was 0.06.

CONCLUSIONS. The low ESBL-E acquisition rate in this study demonstrates that it is possible to control the nosocomial transmission of ESBL in a low-endemic, non-ICU setting where *Escherichia coli* is the most prevalent ESBL-E and standard and contact precautions are applied for known ESBL-E carriers.

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The emergence and global spread of extended-spectrum β -lactamase-producing Enterobacteriaceae (ESBL-E) is a major threat to human health.^{1–5} Infections with ESBL-E are difficult to treat and are associated with increased morbidity, mortality, and healthcare costs.^{6–8} Estimates for the prevalence of ESBL-E rectal carriage at hospital admission range from 4% to 12% in Europe.^{9–17} Nosocomial transmission of ESBL-E is known to occur, and infection control guidelines, therefore, recommend several measures to control the spread of ESBL-E in healthcare settings.^{18,19} In the Netherlands, contact precautions are recommended for all patients known to be colonized or infected with ESBL-E.¹⁸ The risk of acquisition of ESBL-E during hospitalization while applying contact precautions in addition to standard precautions for known ESBL-E carriers is unknown. The present study is based on data from 2 cluster-randomized studies comparing isolation strategies for ESBL-E. We aimed to provide estimates for the acquisition of ESBL-E rectal carriage amongst patients in Dutch hospitals, given the application of contact precautions for known ESBL-E carriers.

METHODS

Contributing Studies

The analyses were performed on data collected in 2 multi-center cluster-randomized studies comparing different isolation strategies for known ESBL-E carriers: (1) the SoM study (Single- or Multiple-occupancy room isolation of patients colonized with ESBL-E)²⁰ and (2) the R-GNOSIS study (Resistance in Gram-Negative Organisms: Studying Intervention Strategies), Work Package 5.²¹ The methods differed slightly between these studies and are presented in Table 1.

Contact Precautions

In both studies, contact precautions were applied in addition to standard precautions for all patients known to be colonized or infected with ESBL-E. In accordance with the national guidelines, standard precautions included the performance of hand hygiene and the use of personal protective equipment (gloves and gown) when anticipating contact with blood or body fluids.^{25–27} Contact precautions additionally included the wearing of gloves at all direct contacts with the patient or the patient's immediate environment or belongings.²⁸

Acquisition of ESBL-E Rectal Carriage During Hospitalization

We used 2 approaches to produce estimates for hospital-acquisition of ESBL-E rectal carriage: (1) a pragmatic

approach, using the results of admission and discharge cultures from patients hospitalized for more than 2 days (both studies) and (2) a Markov chain Monte Carlo (MCMC) model, applied to all patients hospitalized on the participating wards, including those without cultures taken (R-GNOSIS study). The assumptions for each of the approaches are listed in Table 2.

Pragmatic approach. Hospital-acquired ESBL-E rectal carriage was assumed not to be detectable within 2 days of hospital admission.²⁹ Consequently, ESBL-E rectal carriage that was detected within 2 days of hospital admission was considered community-acquired. In addition, patients who were discharged within 2 days of hospital admission were considered not to be at risk for (detectable) hospital-acquired ESBL-E rectal carriage and were excluded from the pragmatic analysis. Admission cultures comprised all cultures taken within 2 days of hospital admission; discharge cultures were all cultures taken on the day of discharge. The prevalence of ESBL-E rectal carriage at hospital admission and hospital discharge were calculated and were used to estimate (1) the prevalence of hospital-acquired ESBL-E rectal carriage at discharge, (2) the cumulative incidence of ESBL-E rectal carriage during hospitalization, and (3) the ESBL-E acquisition rate. For the SoM study, a Markov chain Monte Carlo (MCMC) random-effects analysis was performed to estimate the mean prevalence of ESBL-E rectal carriage at hospital admission and hospital discharge across hospitals, considering within-hospital dependency of the data collected in the 14 participating hospitals. Leave-one-out sensitivity analyses were conducted to evaluate the robustness of the overall estimates.³⁰ By iteratively removing 1 hospital at a time and recalculating parameter estimates, the impact of each hospital on the overall estimates was assessed. Details on the calculations performed can be found in Appendix B online.

Markov chain Monte Carlo model. A previously developed MCMC model was used to quantify hospital-acquisition of ESBL-E rectal carriage in the R-GNOSIS study.^{31,32} This model distinguishes between patient-dependent acquisition and background acquisition. Patient-dependent acquisitions comprise all ESBL-E acquisitions that are dependent on the colonization pressure on the ward³³ and include the transmission of ESBL-E from colonized to noncolonized patients, either directly or indirectly (through the contaminated hands of healthcare workers or the contaminated environment). Background acquisitions cover all other ESBL-E acquisitions, including acquisition from visitors or healthcare workers moving between wards, acquisition from the environment independent of the colonization pressure on the ward, and acquisition through the endogenous route. The latter represents the situation where

TABLE 1. Study Methods

| | SoM Study | R-GNOSIS Study |
|--|---|---|
| Design | Multicenter cluster-randomized study | Multicenter cluster-randomized study ^a |
| Study period | April 2011–February 2014 | January 2014–January 2015 |
| Country | The Netherlands | The Netherlands |
| Hospitals | 6 university and 8 nonuniversity hospitals | 1 university hospital |
| Wards | 124 non-ICU, nonhematology wards | 1 medical and 3 surgical wards |
| Population | Adult patients | Adult patients |
| Detection of ESBL-E rectal carriage | Ward-based prevalence surveys ^b | At admission, followed by twice weekly |
| Microbiological procedures | | |
| Specimen | Perianal and (if applicable) gastrointestinal stoma swabs | Perianal and (if applicable) gastrointestinal stoma swabs |
| Pre-enrichment | Tryptic soy broth with vancomycin (8 mg/L) and cefotaxime (0.125 mg/L) (TSB-VC, Cepheid Benelux, Apeldoorn, the Netherlands) ^c | No |
| ESBL screening agar plate | EbSA (Cepheid Benelux, Apeldoorn, the Netherlands) | ChromID ESBL (bioMérieux, Marcy l'Etoile, France) |
| Species identification | Vitek MS (bioMérieux, Marcy l'Etoile, France) | MALDI Biotyper (Bruker Daltonics, Bremen, Germany) |
| Phenotypic ESBL confirmation | Combination disk diffusion: cefotaxime (30 ug), ceftazidime (30 ug), cefepime (30 ug), alone and combined with clavulanic acid (10 ug) (Neo-Sensitabs, Rosco, Taastrup, Denmark) ^d | Etest ESBL: cefotaxime/cefotaxime + clavulanic acid, ceftazidime/ceftazidime + clavulanic acid, cefepime/cefepime + clavulanic acid (bioMérieux, Marcy l'Etoile, France) ^d |
| Reporting of study culture result | No | Yes |
| Contact precautions following ESBL-E-positive study culture result | No | Yes |

NOTE. ESBL-E, extended-spectrum β -lactamase-producing Enterobacteriaceae; ICU, intensive care unit.

^aThe R-GNOSIS study was performed on medical and surgical wards of 4 university hospitals in the Netherlands, Germany, Switzerland and Spain. The present analysis was limited to the data that were collected in 1 Dutch university hospital during the study period in which contact precautions were applied.

^b5–9 days after institution of contact precautions for a patient known to be colonized or infected with ESBL-E.

^cAs described previously.²²

^dAccording to national and international guidelines.^{23,24}

TABLE 2. Assumptions

| |
|---|
| Pragmatic approach and MCMC model |
| <ul style="list-style-type: none"> • The ESBL-E acquisition rate is constant over time (on average). • ESBL-E carriers remain colonized during their entire hospital stay. • ESBL-E carriers cannot acquire a second ESBL-E. • All patients on a ward on a given day are exposed to the same colonization pressure and have the same risk of patient-dependent acquisition. • The specificity of the method used to detect ESBL-E rectal carriage is 100%. |
| Pragmatic approach only |
| <ul style="list-style-type: none"> • ESBL-E rectal carriage that is detected within 2 days of admission is community-acquired. • Patients are at risk for acquisition of ESBL-E until 2 days before the detection of ESBL-E rectal carriage. • The sensitivity of the method used to detect ESBL-E rectal carriage is 100%. |
| MCMC model only |
| <ul style="list-style-type: none"> • Wards are separate units and acquisition of ESBL-E rectal carriage occurs independently of the colonization pressure on other wards. |

NOTE. ESBL-E, extended-spectrum β -lactamase-producing Enterobacteriaceae; MCMC, Markov chain Monte Carlo.

bacteria are already present in the host at undetectable levels and reach detectable levels under antibiotic pressure. The model accounts for false-negative and missing cultures and, thus, allows estimation of the sensitivity of the method used to detect ESBL-E rectal carriage and the most likely time of ESBL-E acquisition for each patient. A detailed description of the model is provided in Appendix B online. Model parameter estimates were used to obtain estimates for (1) the prevalence of ESBL-E rectal carriage at admission and discharge, (2) the prevalence of hospital-acquired ESBL-E at discharge, (3) the cumulative incidence of ESBL-E rectal carriage during hospitalization, (4) the ESBL-E

acquisition rate, (5) the relative contribution of patient-dependent acquisition to the total ESBL-E acquisition rate, and (6) the average number of ESBL-E acquisitions caused by 1 ESBL-E carrier during a single admission, that is, the per-admission reproduction number (R_A).^{32,34}

Ethical Considerations

The SoM study and the R-GNOSIS study were reviewed by the medical research and ethics committees of the Elisabeth-TweeSteden Hospital (Tilburg, the Netherlands) and the

University Medical Center Utrecht (Utrecht, the Netherlands), respectively. Both studies were judged to be beyond the scope of the Medical Research Involving Human Subjects Act (WMO), and a waiver of written informed consent was granted (SoM: METC/jv/2010.034; R-GNOSIS: WAG/om/13/069083). Patients provided verbal consent for the use of demographic, clinical, and culture data.

RESULTS

In total, 660 prevalence surveys were performed in the SoM study. During these surveys, 10,263 cultures were obtained from 9,136 patients, including 1,718 admission cultures and 1,111 discharge cultures from patients hospitalized for more than 2 days (Table 3). In the R-GNOSIS study, 8,133 cultures were available for 2,787 patients and included 1,483 admission cultures and 680 discharge cultures from patients hospitalized for >2 days.

Table 4 lists the ESBL-E rectal carriage estimates per study and per analytic approach. The prevalence of ESBL-E rectal carriage at admission and discharge was comparable between studies and approaches and varied from 6.4% to 7.4% at admission and from 8.7% to 10.1% at discharge. In both studies, *Escherichia coli* was the most prevalent ESBL-E identified at admission (SoM study 79.9%; R-GNOSIS study 88.8%) (Table 5). The absolute risk of acquisition (cumulative incidence) of ESBL-E rectal carriage during hospitalization varied from 2.4% to 2.9%, and estimates for the ESBL-E acquisition rate ranged from 2.8 to 3.8 acquisitions per 1,000 patient days, with largely overlapping confidence or credible intervals. With the MCMC model, the median background acquisition rate

was estimated to be 0.0028 (95% credible interval [CrI], 0.00088–0.0045) acquisitions per patient day, and the median patient-dependent acquisition rate was 0.010 (95% CrI, 0.00055–0.030) acquisitions per colonized patient day. Based on these estimates and an estimated mean daily prevalence of ESBL-E rectal carriage of 10.6% (95% CrI, 9.0%–12.2%), it was calculated that 28.0% (95% CrI, 1.5%–74.5%) of acquisitions in the R-GNOSIS study were attributable to patient-dependent transmission and the remaining 72.0% (95% CrI, 25.5%–98.5%) resulted from background transmission. Multiplying the patient-dependent acquisition rate by the mean length of hospital stay (6 days) yielded a per-admission reproduction number (R_A) of 0.06. Finally, the MCMC model provided an estimate of 77% (95% CrI, 73%–81%) for the median sensitivity of the method used to detect ESBL-E rectal carriage in the R-GNOSIS study.

In the leave-one-out sensitivity analyses of the SoM study data, all parameter estimates were within the 95% credible intervals of the overall estimates for acquisition of ESBL-E rectal carriage, indicating that the results were not driven by any single hospital (Appendix C online, Figures S1–S5).

DISCUSSION

In this study, performed in the low-endemic setting of Dutch hospitals, where contact precautions are applied for known ESBL-E carriers, the absolute risk of acquisition of ESBL-E rectal carriage was 2.4% to 2.9% with an ESBL-E acquisition rate of 2.8 to 3.8 acquisitions per 1,000 patient days. Estimates for the acquisition of ESBL-E rectal carriage were similar across studies and analytic approaches.

TABLE 3. Study Population Characteristics

| Characteristic | SoM Study | | R-GNOSIS Study | |
|--|--------------------------------|--|--------------------------------|--|
| | All Patients, No. ^a | Length of Hospital Stay >2 d, No. ^a | All Patients, No. ^a | Length of Hospital Stay >2 d, No. ^a |
| Surveys | 660 | 660 | NA | NA |
| Patients | Unknown ^b | Unknown ^b | 4,161 | 2,531 |
| Patients with at least 1 culture | 9,136 | 8,261 | 2,787 | 2,183 |
| Admissions | Unknown ^b | Unknown ^b | 5,188 | 3,003 |
| Admissions with at least 1 culture | 9,604 | 8,682 | 3,315 | 2,558 |
| Cultures | 10,263 | 9,341 | 8,133 | 7,376 |
| Admission cultures | 2,640 | 1,718 | 2,240 | 1,483 |
| Discharge cultures | 1,740 | 1,111 | 1,057 | 680 |
| Culture response, % | 73.4 ^c | Unknown ^b | 63.9 ^d | 85.2 ^d |
| Length of hospital stay | | | | |
| Admissions with at least 1 culture, median d (IQR) | 11 (6–22) | 13 (7–24) | 6 (3–11) | 8 (5–14) |
| Admissions with an admission culture, median d (IQR) | 4 (2–8) | 6 (4–10) | 4 (2–8) | 6 (4–10) |
| Admissions with a discharge culture, median d (IQR) | 4 (1–8) | 7 (5–11) | 4 (1–8) | 7 (4–12) |

NOTE. IQR, interquartile range; NA, not applicable.

^aUnless otherwise specified.

^bNo data were available for nonresponding patients, except for the number of patients per survey.

^cCulture response is calculated as the number of cultures obtained divided by the number of potential cultures.

^dCulture response is calculated as the number of admissions with at least 1 culture divided by the number of admissions.

TABLE 4. Rectal Carriage of Extended-Spectrum β -Lactamase-Producing Enterobacteriaceae (ESBL-E)

| | Pragmatic Approach | | MCMC Model |
|---|---|--|--|
| | SoM Study, % (95% CI/CrI) ^a | R-GNOSIS Study, % (95% CI/CrI) ^a | R-GNOSIS Study, % (95% CI/CrI) ^a |
| ESBL-E rectal carriage | | | |
| Prevalence at admission | 7.4 (5.5–9.9) | 6.4 (5.3–7.8) | 7.0 (6.2–7.8) |
| Prevalence at discharge | 10.1 (7.1–15.0) | 8.7 (6.8–11.0) | 9.3 (8.6–10.0) |
| Hospital-acquired prevalence at discharge | 2.7 (0.6–5.0) | 2.3 (–0.2 to 4.9) | 2.3 (1.7–2.9) |
| Cumulative incidence during hospitalization | 2.9 (0.6–5.4) | 2.4 (–0.2 to 5.2) | 2.5 (2.1–3.0) |
| Acquisition rate, n per 1,000 patient days | 3.7 (0.8–6.9) | 2.8 (–0.3 to 6.1) | 3.8 (2.9–4.9) |

NOTE. CI, confidence interval; CrI, credible interval; ESBL-E, extended-spectrum β -lactamase-producing Enterobacteriaceae; MCMC, Markov chain Monte Carlo.

^aIntervals are either 95% confidence intervals (R-GNOSIS study, pragmatic approach) or 95% credible intervals (SoM study, pragmatic approach and R-GNOSIS study, MCMC model).

TABLE 5. Distribution of Extended-Spectrum β -Lactamase-Producing Enterobacteriaceae in Admission Cultures

| Microorganism | SoM Study | | R-GNOSIS Study | |
|-------------------------------------|------------------|------|-----------------|------|
| | No. | % | No. | % |
| <i>Citrobacter</i> spp | 3 | 2.2 | 1 | 1.0 |
| <i>Enterobacter cloacae</i> complex | 5 | 3.7 | 4 | 4.1 |
| <i>Escherichia coli</i> | 107 | 79.9 | 87 | 88.8 |
| <i>Klebsiella oxytoca</i> | 4 | 3.0 | 0 | 0.0 |
| <i>Klebsiella pneumoniae</i> | 14 | 10.4 | 5 | 5.1 |
| <i>Morganella morganii</i> | 0 | 0.0 | 1 | 1.0 |
| <i>Raoultella</i> spp | 1 | 0.7 | 0 | 0.0 |
| Total | 134 ^a | | 98 ^b | |

^aIn 118 admission cultures, 1 ESBL-E isolate was identified; in 8 admission cultures, 2 ESBL-E isolates were identified.

^bIn 92 admission cultures, 1 ESBL-E isolate was identified; in 3 admission cultures, 2 ESBL-E isolates were identified.

The estimates for the prevalence of ESBL-E rectal carriage at admission and discharge in the present analyses were consistent with those reported for other European hospital-based studies, despite differences in setting, study population, and microbiological methods (Online Appendix A, Table S1).^{9–17}

Although ESBL-E are known to spread within hospitals, quantitative data on the acquisition of ESBL-E rectal carriage during hospitalization in low-endemic settings are limited.¹⁹ In 2 European studies that performed active surveillance cultures at admission and during hospitalization, the cumulative incidence of ESBL-E rectal carriage on non-ICU wards without contact precautions for known ESBL-E carriers was ~4.5%,^{12,13} with an ESBL-E acquisition rate of 1.8 acquisitions per 1,000 patient days.¹² The limited availability of paired samples in both studies may have biased the results; the acquisition of ESBL-E as well as the availability of a second culture are dependent on the length of hospital stay.

Three Swiss studies assessed the acquisition of ESBL-E rectal carriage in roommates of patients with ESBL-E-positive clinical cultures.^{35–37} In 1 study, the acquisition of clonally related ESBL-E

was identified in 5.4% of roommates during hospitalization at a rate of 7.0 acquisitions per 1,000 patient days.³⁵ In the other 2 studies, a single culture was obtained from roommates shortly after the detection of ESBL-E in the index patient. Acquisition of clonally related ESBL-E was identified in 1.5% and 2.6% of roommates, respectively.^{36,37} The ESBL-E acquisition rate, assessed in 1 of these studies, was 3.5 acquisitions per 1,000 patient days.³⁶ The estimates for acquisition of ESBL-E in the Swiss studies were comparable to those in the present analyses, even though the assessment of acquisition of ESBL-E in the Swiss studies was limited to roommates of known ESBL-E carriers, contact precautions were either not applied or only for patients at high risk for ESBL-E carriage, and the possibility of horizontal transfer of resistance genes was not taken into account.

To the best of our knowledge, this is the first study to include an MCMC model approach to provide quantitative data on the acquisition of ESBL-E rectal carriage in hospitals. Other MCMC model-based studies on the acquisition of antimicrobial-resistant Enterobacteriaceae were performed in nonhospital settings or were aimed at other resistance mechanisms.^{32,38} The per-admission reproduction number estimated in the current study was far below 1, indicating that patient-to-patient transmission of ESBL-E during a single admission of an ESBL-E carrier is not sufficient to maintain endemicity of ESBL-E in Dutch hospitals that use contact precautions for known ESBL-E carriers. The estimate for the sensitivity of the method used to detect rectal carriage of antimicrobial-resistant Enterobacteriaceae was comparable to those reported in the other studies, which supports the robustness of the MCMC model.^{32,38}

One of the benefits of the MCMC algorithm used in this study is that it allows estimation of the most likely time of ESBL-E acquisition for each patient, including patients with missing or false-negative cultures. In addition, separate estimates are provided for patient-dependent acquisition and background acquisition. In the present study, the relative contribution of patient-dependent acquisition to the total number of hospital acquisitions was estimated to be 28.0% (95% confidence interval [CI], 1.5%–74.5%). The rather high

level of uncertainty around this estimate may be due to a relatively high percentage (36%) of admissions with no cultures taken. The relative contribution of patient-dependent acquisition might be interpreted as the maximum achievable reduction in hospital acquisition of ESBL-E rectal carriage when infection control measures would be optimized. It is obvious that the relative importance of patient-dependent acquisition is dependent on the number of colonized patients present on the ward and the effectiveness of infection control measures. The low estimate for patient-dependent acquisition in the present study can, therefore, not be generalized to settings with a high-endemic level of ESBL-E rectal carriage or settings with less effective infection control policies.

The analysis in the pragmatic approach was restricted to cultures taken at hospital admission and hospital discharge. Estimates for hospital-acquisition of ESBL-E rectal carriage that are based on cultures taken during hospitalization may be biased, as ESBL-E rectal carriage is associated with a prolonged length of hospital stay, leading to overrepresentation of ESBL-E carriers in prevalence surveys, and thus overestimation of hospital-acquired ESBL-E carriage. This finding is clearly illustrated by the MCMC model estimates for the R-GNOSIS study, where the mean daily prevalence was estimated to be 10.6% with a prevalence of 7.0% at admission and 9.3% at discharge.

Both studies and analytic approaches were based on phenotypic ESBL confirmation methods. The use of phenotypic data, without species identification and molecular typing, allows for the detection of transmission of the ESBL phenotype due to horizontal gene transfer and, herewith, increases the sensitivity to detect transmission of ESBL-encoding genes between patients.

Several assumptions were made for quantifying hospital-acquired ESBL-E rectal carriage, which may all have resulted in underestimating the risk and rate of acquisition.

In both studies, contact precautions were applied in addition to standard precautions for all known ESBL-E carriers, according to the national guideline²⁸. Nevertheless, some acquisition of ESBL-E rectal carriage was observed, partly due to patient-to-patient transmission. For the SoM study, the results of study cultures were blinded, and identification of ESBL-E carriers was based on clinical cultures only. Asymptomatic ESBL-E carriers who were not detected by clinical cultures might, thus, have contributed to the observed acquisition of ESBL-E. In the R-GNOSIS study, the results of all study cultures were reported to the treating physicians and contact precautions were applied for all ESBL-E carriers, including those detected in study cultures. However, as not all patients were sampled, and some culture results might have been falsely negative, the acquisition of ESBL-E from undetected ESBL-E carriers cannot be excluded in the R-GNOSIS study either. Finally, noncompliance with recommended infection control measures may have contributed to the observed spread of ESBL-E in both studies.

Recent studies suggest that *E. coli* has a lower intrinsic transmission capacity than *K. pneumoniae*.^{35,36} Hence, estimates for hospital-acquisition of ESBL-E rectal carriage will be dependent on the distribution of ESBL-producing bacterial

species in ESBL-E carriers. The high relative prevalence of ESBL-producing *E. coli* in the present study may well have contributed to the low estimates for acquisition.

This study provides quantitative data on the prevalence and acquisition of ESBL-E rectal carriage amongst patients in Dutch hospitals. The hospital acquisition rate of ESBL-E rectal carriage was low and the per-admission reproduction rate far below 1. This demonstrates that it is possible to control the nosocomial transmission of ESBL in a low-endemic, non-ICU setting where *E. coli* is the most prevalent ESBL-E and standard and contact precautions are applied for known ESBL-E carriers, which is promising considering the global emergence of Enterobacteriaceae with plasmid-mediated resistance.

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SUPPLEMENTARY MATERIAL

To view supplementary material for this article, please visit <https://doi.org/10.1017/ice.2017.241>

REFERENCES

1. Paterson DL, Bonomo RA. Extended-spectrum beta-lactamases: a clinical update. *Clin Microbiol Rev* 2005;18:657–686.
2. Hawkey PM, Jones AM. The changing epidemiology of resistance. *J Antimicrob Chemother* 2009;64:i3–i10.
3. Woerther PL, Burdet C, Chachaty E, Andremont A. Trends in human fecal carriage of extended-spectrum beta-lactamases in the community: toward the globalization of CTX-M. *Clin Microbiol Rev* 2013;26:744–758.
4. Antimicrobial resistance surveillance in Europe. 2014. Annual report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). European Centre for Disease Prevention and Control website. http://ecdc.europa.eu/en/publications/_layouts/forms/Publication_DispForm.aspx?List=4f55ad51-4aed-4d32-b960-af70113dbb90&ID=1400. Published 2015. Accessed August 15, 2017.
5. NethMap. 2015. Consumption of antimicrobial agents and antimicrobial resistance among medically important bacteria in the Netherlands in 2014. Dutch Foundation of the Working Party on Antibiotic Policy website. [http://www.swab.nl/swab/cms3.nsf/uploads/4F5A0D8E6F0DD139C1257E6E0051833A/\\$FILE/NethmapMaran2015%20_webversie.pdf](http://www.swab.nl/swab/cms3.nsf/uploads/4F5A0D8E6F0DD139C1257E6E0051833A/$FILE/NethmapMaran2015%20_webversie.pdf). Published 2015. Accessed August 15, 2017.

6. Ammerlaan HSM, Troelstra A, Kruitwagen CLJJ, Kluytmans JAJW, Bonten MJM. Quantifying changes in incidences of nosocomial bacteraemia caused by antibiotic-susceptible and antibiotic-resistant pathogens. *J Antimicrob Chemother* 2009;63:1064–1070.
7. Rottier WC, Ammerlaan HSM, Bonten MJM. Effects of confounders and intermediates on the association of bacteraemia caused by extended-spectrum beta-lactamase-producing Enterobacteriaceae and patient outcome: a meta-analysis. *J Antimicrob Chemother* 2012;67:1311–1320.
8. Stewardson A, Fankhauser C, De Angelis G, et al. Burden of bloodstream infection caused by extended-spectrum beta-lactamase-producing Enterobacteriaceae determined using multistate modeling at a Swiss university hospital and a nationwide predictive model. *Infect Control Hosp Epidemiol* 2013;34:133–143.
9. Esposito S, Capuano A, Noviello S, et al. Modification of patients' endogenous bacterial flora during hospitalization in a large teaching hospital in Naples. *J Chemother* 2003;6:568–573.
10. Ben-Ami R, Schwaber MJ, Navon-Venezia S, et al. Influx of extended-spectrum beta-lactamase-producing Enterobacteriaceae into the hospital. *Clin Infect Dis* 2006;42:925–934.
11. Ruppé E, Pitsch A, Tubach F, et al. Clinical predictive values of extended-spectrum beta-lactamase carriage in patients admitted to medical wards. *Eur J Clin Microbiol Infect Dis* 2012;31:319–325.
12. Schoevaerdt D, Verroken A, Huang TD, et al. Multidrug-resistant bacteria colonization amongst patients newly admitted to a geriatric unit: a prospective cohort study. *J Infect* 2012;65:109–118.
13. Pasricha J, Koessler T, Harbarth S, et al. Carriage of extended-spectrum beta-lactamase-producing Enterobacteriaceae among internal medicine patients in Switzerland. *Antimicrob Resist Infect Control* 2013;2:20.
14. Shitrit P, Reisfeld S, Paitan Y, et al. Extended-spectrum beta-lactamase-producing Enterobacteriaceae carriage upon hospital admission: prevalence and risk factors. *J Hosp Infect* 2013;85:230–232.
15. Platteel TN, Leverstein-van Hall MA, Cohen Stuart JW, et al. Predicting carriage with extended-spectrum beta-lactamase-producing bacteria at hospital admission: a cross-sectional study. *Clin Microbiol Infect* 2015;21:141–146.
16. Willemsen I, Oome S, Verhulst C, Pettersson A, Verduin K, Kluytmans J. Trends in extended spectrum beta-lactamase (ESBL) producing Enterobacteriaceae and ESBL genes in a Dutch teaching hospital, measured in 5 yearly point prevalence surveys (2010–2014). *PLoS ONE* 2015;10:e0141765.
17. Huizinga P, Kluytmans-van den Bergh M, Rijen M, Willemsen I, van 't Veer N, Kluytmans J. Proton pump inhibitor use is associated with extended-spectrum beta-lactamase-producing Enterobacteriaceae rectal carriage at hospital admission: a cross-sectional study. *Clin Infect Dis* 2017;64:361–363.
18. Kluytmans-VandenBergh MFQ, Kluytmans JAJW, Voss A. Dutch guideline for preventing nosocomial transmission of highly resistant microorganisms (HRMO). *Infection* 2005;33:309–313.
19. Tacconelli E, Cataldo MA, Dancer SJ, et al. ESCMID guidelines for the management of the infection control measures to reduce transmission of multidrug-resistant gram-negative bacteria in hospitalized patients. *Clin Microbiol Infect* 2014;20:S1–S55.
20. Kluytmans JAJW. Single- or multiple-occupancy room isolation of patients colonised with ESBL-producing Enterobacteriaceae. Trial ID NTR2799. Netherlands Trialregister website. <http://www.trialregister.nl/trialreg/admin/rctview.asp?TC=2799>. Published 2011. Accessed August 15, 2017.
21. Gastmeier P. Patient isolation strategies for extended spectrum beta lactamase (ESBL) carriers in medical and surgical hospital wards. ISRCTN57648070. ISRCTN Registry website. <http://www.isrctn.com/ISRCTN57648070>. Published 2014. Accessed August 15, 2017.
22. Kluytmans-van den Bergh MFQ, Verhulst C, Willemsen LE, Verkade E, MJM Bonten, Kluytmans JAJW. Rectal carriage of extended-spectrum beta-lactamase-producing Enterobacteriaceae in hospitalized patients: selective preenrichment increases yield of screening. *J Clin Microbiol* 2015;53:2709–2712.
23. NVMM guideline laboratory detection of highly resistant microorganisms, version 2.0. Netherlands Society for Medical Microbiology website. <http://www.nvmm.nl/richtlijnen/hrmo-laboratory-detection-highly-resistant-microorganisms>. Published 2012. Accessed August 15, 2017.
24. EUCAST guidelines for detection of resistance mechanisms and specific resistances of clinical and/or epidemiological importance, version 1.0. European Committee on Antimicrobial Susceptibility Testing website. http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Resistance_mechanisms/EUCAST_detection_of_resistance_mechanisms_v1.0_20131211.pdf. Published 2012. Accessed August 15, 2017.
25. Dutch Working Party on Infection Prevention. Richtlijn Handhygiëne. Rijksinstituut voor Volksgezondheid en Milieu website. <http://www.rivm.nl/dsresource?objectid=3f4dd7ba-f3c4-48ea-9fed-8876b2ca5845&type=org&disposition=inline>. Published 2014. Accessed August 15, 2017.
26. Dutch Working Party on Infection Prevention. WIP-richtlijn Persoonlijke beschermingsmiddelen. Rijksinstituut voor Volksgezondheid en Milieu website. <http://www.rivm.nl/dsresource?objectid=5426e1c5-5355-4d03-98ef-e51e9e27e9dd&type=org&disposition=inline>. Published 2015. Accessed August 15, 2017.
27. Dutch Working Party on Infection Prevention. WIP-richtlijn Persoonlijke hygiëne medewerker. Rijksinstituut voor Volksgezondheid en Milieu website. <http://www.rivm.nl/dsresource?objectid=dfa15c98-834b-4d5c-a392-1278ad81345c&type=org&disposition=inline>. Published 2014. Accessed August 15, 2017.
28. Dutch Working Party on Infection Prevention. WIP-richtlijn Contactisolatie. Rijksinstituut voor Volksgezondheid en Milieu website. <http://www.rivm.nl/dsresource?objectid=3ce6b915-2169-4bbf-be05-7b7fc0dc527d&type=org&disposition=inline>. Published 2014. Accessed August 15, 2017.
29. National Healthcare Safety Network (NHSN) patient safety component manual. National Health Safety Network website. https://www.cdc.gov/nhsn/pdfs/pscmanual/pscmanual_current.pdf. Published 2017. Accessed August 15, 2017.
30. Higgins JPT. Commentary: Heterogeneity in meta-analysis should be expected and appropriately quantified. *Int J Epidemiol* 2008;37:1158–1160.
31. Worby CJ, Jeyaratnam D, Robotham JV, et al. Estimating the effectiveness of isolation and decolonization measures in reducing transmission of methicillin-resistant *Staphylococcus aureus* in hospital general wards. *Am J Epidemiol* 2013;177:1306–1313.
32. Haverkate MR, Bootsma MCJ, Weiner S, et al. Modeling spread of KPC-producing bacteria in long-term acute care hospitals in the Chicago region, USA. *Infect Control Hosp Epidemiol* 2015;36:1148–1154.
33. Bonten MJM, Slaughter S, Ambergen AW, et al. The role of "colonization pressure" in the spread of vancomycin-resistant enterococci. *Arch Intern Med* 1998;158:1127–1132.

34. Cooper BS, Kypraios T, Batra R, Wyncoll D, Tosas O, Edgeworth JD. Quantifying type-specific reproduction numbers for nosocomial pathogens: evidence for heightened transmission of an Asian sequence type 239 MRSA clone. *PLoS Comput Biol* 2004;8:e1002454.
35. Hilty M, Betsch BY, Bögli-Stuber K, et al. Transmission dynamics of extended-spectrum beta-lactamase-producing Enterobacteriaceae in the tertiary care hospital and the household setting. *Clin Infect Dis* 2012;55:967–975.
36. Tschudin-Sutter S, Frei R, Dangel M, Stranden A, Widmer AF. Rate of transmission of extended-spectrum beta-lactamase-producing Enterobacteriaceae without contact isolation. *Clin Infect Dis* 2012;55:1505–1511.
37. Tschudin-Sutter S, Frei R, Schwahn F, et al. Prospective validation of cessation of contact precautions for extended-spectrum beta-lactamase-producing *Escherichia coli*. *Emerg Infect Dis* 2016;22:1094–1097.
38. Haverkate MR, Platteel TN, Fluit AC, et al. Quantifying within-household transmission of extended-spectrum beta-lactamase-producing bacteria. *Clin Microbiol Infect* 2017;23:46.e1–46.e7.