Contents lists available at ScienceDirect

### Cellular Immunology

journal homepage: www.elsevier.com/locate/ycimm

Review article

# Exhaustion of T lymphocytes in the tumor microenvironment: Significance and effective mechanisms



Mohammad Davoodzadeh Gholami<sup>a,b</sup>, Gholam Ali kardar<sup>c,\*\*</sup>, Yousef Saeedi<sup>d</sup>, Sahel Heydari<sup>a,b</sup>, Johan Garssen<sup>d</sup>, Reza Falak<sup>a,b,\*</sup>

<sup>a</sup> Immunology Research Center, Iran University of Medical Sciences, Tehran, Iran

<sup>b</sup> Department of Immunology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran

<sup>c</sup> Immunology, Asthma and Allergy Research Institute, Tehran University of Medical Sciences, Tehran, Iran

<sup>d</sup> Department of Pharmaceutical Sciences, Utrecht University, Netherlands

#### ARTICLE INFO

Keywords: Tumor microenvironment T cell exhaustion T cell senescence T cell anergy Inhibitory receptors

#### ABSTRACT

T lymphocytes play crucial roles in adaptive immune responses to tumors. However, due to different tolerance mechanisms and inhibitory effects of the tumor microenvironment (TME) on T cells, responses to tumors are insufficient. In fact, cellular and molecular suppressive mechanisms repress T cell responses in the TME, resulting in senescent, anergic and exhausted lymphocytes. Exhaustion is a poor responsive status of T cells, with up-regulated expression of inhibitory receptors, decreased production of effective cytokines, and reduced cytotoxic activity. Low immunogenicity of tumor antigens and inadequate presentation of tumor-specific antigens results in inappropriate activation of naive T lymphocytes against tumor antigens. Moreover, when effector cytotoxic T cells enter TME, they encounter a complicated network of cells and cytokines that suppress their effectiveness and turn them into exhausted T cells. Thus, the mechanism of T cell exhaustion in cancer is different from that in chronic infections. In this review we will discuss the main components such as inhibitory receptors, inflammatory cells, stromal cells, cytokine milieu as well as environmental and metabolic conditions in TME which play role in development of exhaustion. Furthermore, recent therapeutic methods available to overcome exhaustion will be discussed.

#### 1. Introduction

Tumorigenesis is usually initiated following stepwise accumulation of genetic and epigenetic alterations that control cellular apoptosis and mortality [1]. Mutations may evoke neoplastic phenotypes in normal cells. These accumulated mutations alone generally do not result in cancer formation. Hence, the interaction and crosstalk between cancer cells and the supporting cell types that form the tumor microenvironment (TME) indicate that this environment plays a critical role in cancer development [2]. The TME is a key factor in the escape of tumor cells from the immune system. This region contains mainly angiogenic vascular endothelial cells, infiltrating immune cells, and stromal cells [3].

Pro-angiogenic factors such as vascular endothelial growth factor (VEGF) and platelet-derived growth factor (PDGF) are upregulated in most tumors and can result in the development of immature vessels with abnormal structures in the TME. Vascular hyper-permeability is a hallmark of tumor pathophysiology, and one of its consequence is diminished tumor perfusion. Reduced perfusion leads to hypoxia and acidification of the TME and results in decreased leukocyte infiltration [4].

The altered metabolism of cancer cells influences the nutritional status in the TME and influences the metabolic fitness of leukocytes leading to low glucose and acidification of the TME [5–7]. Hypoxia promotes tumorigenesis by enhancing cancer cell proliferation, and the low pH increases the suppressive activity of tumor-infiltrating myeloid-derived suppressor cells (MDSCs) and tumor-associated macrophages (TAMs), and reduces the cytotoxicity and proliferation of T lymphocytes in the TME [4].

Cytotoxic CD8<sup>+</sup> T cells (CTLs) are considered as one of main effector cell types of the adaptive immune system responsible for combating cancer cells. Despite the presence and activation of several immunologic components, especially CTLs in the TME, tumor cells are not easily eradicated [8]. Mechanisms involved in this impaired

heidari\_sahel69@yahoo.com (S. Heydari), j.garssen@uu.nl (J. Garssen), Falak.r@iums.ac.ir (R. Falak).

http://dx.doi.org/10.1016/j.cellimm.2017.10.002

Received 26 August 2017; Received in revised form 8 October 2017; Accepted 9 October 2017 Available online 10 October 2017 0008-8749/ © 2017 Elsevier Inc. All rights reserved.

ELSEVIER

<sup>\*</sup> Corresponding author at: Department of Immunology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran.

<sup>\*\*</sup> Co-corresponding author at: Department of Medical Biotechnology, School of Advanced Technologies in Medicine, Tehran University of Medical Sciences, Tehran, Iran.

E-mail addresses: dawoodzadeh@gmail.com (M. Davoodzadeh Gholami), gakardar@tums.ac.ir (G.A. kardar), y.saeedi86@gmail.com (Y. Saeedi),



Fig. 1. Dysfunctional subsets of T lymphocytes in the tumor microenvironment and their features.

responsiveness include depletion of naive anti-tumor T cells during thymic lymphocyte development, unresponsiveness of CTLs due to expression of reduced amounts of the co-stimulatory molecules CD80 (B) 7-1) and CD86 (B7-2), and prolonged presence of immunomodulator cells and secretion of soluble factors from those cells, which can directly inhibit the defensive mechanisms of leukocytes or indirectly disrupt their activation process and undermine the CTL response in the TME [9]. The presence of immune suppressive agents in the TME leads to T cell dysfunction in the TME in the context of anergy, senescence, and exhaustion (Fig. 1). Consequently, although immune cells are found in the TME, they are not fully effective.

The concept of T cell exhaustion was first described in CTLs in chronic lymphocytic choriomeningitis virus (LCMV) infection in mice [10,11], and was subsequently reported in human chronic viral infections and cancers as well. T cell exhaustion is a state of T cell dysfunctionality and the severity of T cell exhaustion appears to increase according to antigen concentration and decreased CD4<sup>+</sup> T cell numbers [12]. Exhausted T cells progressively lose their proliferation, cytokine production, and cytotoxic capabilities. Several studies confirm that exhausted T cells express increased levels of inhibitory receptors, including programmed cell death protein 1 (PD-1), cytotoxic T lymphocyte antigen-4 (CTLA-4), lymphocyte activation gene 3 (LAG-3), T cell immunoglobulin and mucin domain containing-3 (TIM-3), B and T lymphocyte attenuator (BTLA), and T cell immunoreceptor with Ig and ITIM domains (TIGIT) [13-19]. Interestingly, it was reported that exhausted CTLs co-express inhibitory receptors and the pattern and number of inhibitory receptors correlate with T cell exhaustion levels [20]. Blocking inhibitory receptors with specific antibodies may reverse the functionality and anti-tumor responses of exhausted T cells. However, application of blocking antibodies against inhibitory receptors in combination therapy has shown promising results in the treatment of advanced stages of cancer. In this paper we discuss mechanisms that may limit CTL effectiveness and promote the development of inefficient T cell subsets, especially those exhausted status in the TME that cannot efficiently combat tumor cells, allowing them to survive and grow.

#### 2. Exhausted T lymphocytes differ from other subsets in the TME

As previously mentioned, evidence indicates that effector T cell phenotypes and capabilities are dramatically impaired in the TME, resulting in T cell exhaustion. Various molecular and cellular mechanisms contribute to this phenomenon. In this section, we discuss lymphocyte dysfunctional phenotypes including anergic, senescent, and exhausted T cells, and also their underlying developmental mechanisms.

#### 2.1. Anergic T lymphocytes

T cell anergy is generally described as an induced state of hyporesponsiveness with impaired proliferation and IL-2 secretion (Table 1). It has been proposed that T cell anergy induces peripheral tolerance and protection from autoimmune disease development [21–25]. Although several studies support T cell anergy in cancer, the exact reasons for anergic T cell development in the TME are unclear.

T cell anergy generally happens following their incomplete activation in response to suboptimal amounts of IL-2 or absence of co-stimulatory signals [21,23]. Obviously, tolerance mechanisms inhibit the immune response against cancer; however, the exact mechanisms of their induction in the TME is not known. Effective mechanisms responsible for T cell anergy in the TME depend mostly on molecular properties of these cells, such as expression of surface molecules. In the TME, B7 family stimulatory and inhibitory receptors do not have the arbitrary distribution as in immunological response to tumor cells. In fact, in this microenvironment, tumor cells and the antigen-presenting cells (APCs) overexpress programmed death-ligand 1 (PD-L1), while expression of the stimulatory receptors CD80 and CD86 are diminished [10,26,27]. Studies in mouse cancer models demonstrated that inducing B7-1 expression on tumor cells or stopping the expression of inhibitory receptor expression on these cells can reduce tumor growth [26-32]. In addition, appropriate stimulation of anergic T cells in the mouse cancer model expands the specific anti-tumor T cell population and returns them from the anergic to the effective mode and helps them to overcome and eradicate the tumor cells [33-35]. Overall T cell anergy is a reversible dysfunctional state of a subset of T cells and tumorinduced T cell anergy in the TME may be one of the immune evasion mechanisms in cancer.

#### 2.2. senescent T lymphocytes

Senescent T cells are characterizes by telomere shortening, arrested cell cycle, and phenotypic changes including downregulated CD28 receptor expression (Table 1) [24,25,36,37]. Telomere shortening is an inherent consequence of cell division, which affects cell function and leads to cellular senescence [38]. Cell cycle controlling proteins including p16, p21, and p53 normally inhibit cell cycle progression and accumulate in senescent cells [39–41]. In addition to phenotypic changes, senescent T cells manifest defective killing abilities and develop negative regulatory functions [42,43].

Aging is a normal physiological process of cells; however, the percentage of senescent T cells in young individuals with autoimmune diseases and chronic infections is higher than in normal individuals [44], indicating that repeated activation and proliferation may promote aging in these cells [45]. Moreover, co-culture of cancer cells with T cells induced senescence in the T cells [46]. Studies of patients with lung or head and neck cancers suggests that following T cell senescence, CD28<sup>-/dim</sup> CTLs will predominate [47,48]. In addition, it has been shown that reduction in CD28 expression and overexpression of TIM-3, CD57, and killer cell lectin-like receptor subfamily G member 1 (KLRG1) is associated with T cell senescence [49–54]. Some researchers believe that DNA damage, which occurs in thymic lymphocyte progenitors, may exit these cells from the normal cell cycle and induce

#### Table 1

Comparison of dysfunctional T cell.

Features	Exhausted T cells	Anergic T cells	Senescent T cells	Refs.
Phenotype	Reduced proliferation Increased experssion of inhibitory receptor Decreased cytokine production Reduced cytotoxicity	Reduced proliferation Reduced IL-2 secretion	Telomere shorting Cell-cycle arrest Phenotypic changes	[12-19,21-25,36,37]
Surface marker profile	CD45RO <sup>+</sup> CD57 <sup>+</sup> CD95 <sup>+</sup> PD-1 <sup>+</sup> CTLA-4 <sup>+</sup> TIM-3 <sup>+</sup> LAG-3 <sup>+</sup> BTLA <sup>+</sup>	LAG-3 <sup>+</sup> PD-1 <sup>+</sup>	KLRG1 <sup>+</sup> CD28 <sup>-</sup> CD57 <sup>+</sup>	[12-19,21-25,36,37]
Transcription profile	NFAT, <i>T</i> -bet <sup>low</sup> /Eomes <sup>high</sup> , Blimp-1, BATF, FoxP3	NFAT, NF-ĸB/RelA, Ikaros, Egr1/Egr2	FoxP3	[12-19,21-25,36,37]
Main causes	Excessive or continuous stimulation	Suboptimal stimulation	Repetitive stimulation	[19,24,25,36]
Detailed reason of development	Persistent overstimulation of signal 1 and signal 3 leads to progressive loss of effector and memory function	Absent or weak signal 2 and/or signal 3 during priming of T cells results dysfunctionality	Intermittent and/or multiple stimulations after the primary response leads to limited cell replication	[19,24,25,37,68]
Time frame of occurrence	Typically within a few weeks	Typically within a few days	Typically within months to years	[19,24,25,36,68]
Proliferative capacity	Low	Low	Low	[19,24,25,36]
Effector function	Low to moderate	None or low	High	[19,24,25,36]
Proper priming	Yes	No	Yes	[19,24,25]

Signal 1: Interaction of MHC-peptide complex with specific T cell receptor.

Signal 2: Involvement of co-stimulatory or co-inhibitory molecules.

Signal 3: Signals from soluble cytokines.

their senescence; however, this mechanism in the TME has not been confirmed [55]. Senescent T cells are dysfunctional and incapable of killing target cells, and may in fact inhibit normal T cell activity in the TME [42,43].

#### 2.3. Exhausted T lymphocytes

Exhaustion is hypo-responsiveness of T cells, along with reduced proliferation, increased expression of the inhibitory receptors PD-1, LAG-3, TIM-3, CTLA-4, BTLA, and TIGIT, and decreased production of IL2, IFNy, TNFa, and granzyme B. The impaired cytotoxicity of these cells is shown in Table 1 [12-19,24,25]. Severity of T cell exhaustion appears to rise with increased antigen load and decreased specific T CD4<sup>+</sup> cells [12]. The typical sign of T cell exhaustion is expression of the inhibitory receptor, PD-1 [13]. Interestingly, results of studies in mice and humans have shown that exhausted CTLs co-express inhibitory receptors and the pattern and number of these receptors correlate with the levels of T cell exhaustion [20]. Approximately one-third to one-half of CD8<sup>+</sup> tumor-infiltrating lymphocytes (TILs) co-express PD-1 and CTLA-4. PD-1<sup>+</sup> CTLA4<sup>+</sup> CD8<sup>+</sup> TILs are more severely exhausted and show lower proliferation and cytokine production capability than normal CTLs [43]. In addition, other inhibitory receptors including TIM-3, LAG-3, BTLA, and TIGIT have been demonstrated to regulate T cell exhaustion in cancer. Most researchers believe that T cell exhaustion occurs due to continuous stimulation of these cells, resulting in their loss of effectiveness and eventual apoptosis. However, some researchers believe that the developmental disorders of memory T cells are mainly due to exhaustion [20]. Although the steps that direct T cell exhaustion in chronic infections are clear, the process in the TME is not yet known. Albeit, since chronic conditions occur in both of those conditions, similar characteristics can be considered for exhausted T cells in either case. Obviously, the TME has a great impact on the phenotype, metabolism, functionality, and maintenance of these cells [56].

#### 3. CD8<sup>+</sup> T lymphocyte exhaustion in the TME

In chronic infections, the characteristics of exhausted T cells are the following: 1) progressive impairment in effectiveness or functionality, 2) upregulated co-expression of inhibitory receptors, 3) diminished

production of effective cytokines such as IL-2, IFN $\gamma$ , and TNF $\alpha$ , 4) impaired in vitro cytotoxic activity, 5) poor responsiveness to survival factors such as IL-7 and IL-15, and 6) alteration in cellular metabolism, DNA transcription level, and functionality of transcription factors including T-bet and Eomes [53,57,58]. While some of the mentioned criteria such as alterations of the expression level of some of receptors, cytokines, and transcription factors are observed in all types of exhausted cells, certain criteria are merely restricted to exhausted T cells in chronic infections. Due to similarities of tumor antigens with selfantigens and also the existence of central and peripheral tolerance mechanisms, effectiveness of CTL responses in cancer is mediocre. In addition, presentation of cancer antigens in the absence of inflammatory cytokines and the presence of inhibitory mechanisms such as self-antigen-specific regulatory T cells ameliorate the condition [59]. Therefore, mechanisms of T cell exhaustion in cancer differ from those of chronic infections.

In chronic infections, the naive T cells naturally encounter presented antigens and differentiate into effective CTLs, but following high resistance of the pathogen and failure in antigen removal, effective CTLs become exhausted. However, in the TME, due to low expression and immunogenicity of tumor antigens, low affinity of specific T cell receptors (TCRs) to tumor antigens [60], insufficient presentation of the tumor antigens to T cells [56], absence of inflammatory cytokines, and reduced expression of co-stimulatory molecules, the process of differentiation of naive CTLs to effective cells is disrupted and naive T cells are not fully activated. On the other hand, following entry of effective CTLs to the TME, they encounter a complicated regulatory network of various cells including cancer, inflammatory, and stromal cells, and secreted cytokines, which could suppress and convert them to exhausted phenotype [9]. Changes in the metabolic state and nutrient availability of cancer cells may alter the functional fate of T cells in the TME. Some researchers believe that exhaustion of CTLs may be triggered by metabolic stresses within the TME [61]. Cancer cells strongly consume glucose through glycolysis as a main metabolic program, thus they capture a high percentage of environmental glucose for energy production. Consequently, T cells will encounter a lack of glucose (hypoglycemia) in the TME due to competition with cancer cells. Hypoglycemia may prevent full activation of CTLs and also decrease the expansion, differentiation, and effector functions of these cells in the TME [62]. In addition, these metabolic conditions increase PD-1



Fig. 2. Differentiation of T CD8<sup>+</sup> T lymphocyte to exhausted cells in the tumor microenvironment. In the secondary lymphoid organs, the encounter of naive T cells with tumor antigens results in proliferation and differentiation of effector T cells. In the tumor microenvironment, effector T cells are polarized into exhausted T cells, with decreased effector cytokine production and increased inhibitory receptor expression. Subsequently, exhausted T cells may differentiate to defective memory T cells or be deleted physically.

expression on activated CTLs and promote differentiation of T cells into regulatory T lymphocytes (Tregs) [63]. On the other hand, excessive consumption of glucose through glycolysis in cancer cells produces large amounts of lactic acid, which can suppress proliferation, production of effective cytokines, and cytotoxic activity of CTLs in the TME [64]. Also excessive metabolism of the amino acids tryptophan, arginine, and glutamine by cancer cells causes an amino acid deficiency that can suppress CTL anti-tumor responses in the TME [65,66]. Accumulation of adipocytes and adipocyte-like fibroblasts, and production of large amounts of fatty acids by cancer cells lead to lipid-enrichment in the TME. These metabolic conditions may promote development of Tregs and suppress effector CTL functions in the TME [67].

## 4. Differentiation of ${\rm CD8}^+$ T lymphocytes to exhausted cells in the TME

After differentiation of CD44<sup>low</sup>, CD62L<sup>high</sup> naive CTLs to CD44<sup>high</sup>, CD62L<sup>low</sup> effective cells in secondary lymph tissues, these cells may enter the TME and become exhausted cells, which eventually will be removed or differentiate to defective memory cells (Fig. 2). So, it seems that in the TME, the exhausted CTLs originate from the effective CTLs that leave the environment and become dysfunctional by the mentioned mechanisms. Some researchers believe that exhaustion of CTLs is due to impaired formation of memory T cells [20]; however, Pauken et al. illustrated that exhausted T cells are a distinct lineage from effector or memory CTLs [68].

## 5. Correlation of the exhaustion of CTLs with deletion of CTLs in the TME

PD-L1, one of the major factors in the TME because of its high expression in cancer tissues and also its capability to induce apoptosis in CTLs, plays a crucial role in the viability of these cells. It was shown that increasing the expression of this molecule on the surface of cancer cells results in impaired CTL survival and elimination of their population in the TME [69]. Immunohistochemical staining showed that

upregulated expression of PD-L1 in hepatocellular carcinoma is directly associated with increased apoptosis of CTLs in the TME. On the other hand, CTLs produce IFN $\gamma$ , which induces the expression of PD-L1 on hepatocytes and causes apoptosis in these cells. These observations suggest that the local reduction of CTLs occurs in advanced stages of exhaustion [70].

#### 6. Effective Internal signals in exhaustion of CTLs in the TME

Involvement of the inhibitory receptors PD-1, LAG-3, and TIM-3, as well as alteration of transcription factors, are effective mechanisms for the induction of exhaustion in CTLs and is discussed in detail below (Fig. 3).

#### 6.1. Inhibitory receptors

T cell activation requires three signals; these include: 1) recognition of MHC-peptide complexes by the TCR, 2) pairing of co-stimulatory or co-inhibitory molecules, and 3) alarms from proper soluble cytokines [71].

Of these signals, the second plays a crucial role in production of the cytokine profile responsible for differentiation of lymphocytes into activator or inhibitor phenotypes. In normal conditions, molecules that generate co-inhibitory messages, such as PD-1 and CTLA-4, inhibit inflammation and prevent the inappropriate escalation of immune system responses and tissue damage. However, excessive increases of co-inhibitory signals weaken immune responses and result in exhaustion of CTLs in the early stages, and finally enhance the expression of the inhibitory receptors PD-1, LAG-3, TIM-3, CTLA-4, BTLA, and TIGIT, as well as reduced production of granzyme B and the stimulatory cytokines IL-2, IFN $\gamma$ , and TNF $\alpha$  [56,72]. During this process, exhausted CTLs gradually lose their ability to produce cytokines. For example, in the early stages of exhaustion, IL-2 production decreases significantly [19]; in the middle stages, TNFa production is decreased, and in the advanced stages, T cells cannot produce IFN $\gamma$  or granzyme B, thus the performance of CTLs as tumor cell eradicators is limited [12].



Fig. 3. Regulatory mechanisms of T cell exhaustion in the tumor microenvironment. Cancer cells, regulatory T lymphocytes (Tregs), tumor-associated macrophages (TAMs), myeloid-derived suppressor cells (MDSCs) and plasmacytoid dendritic cells are major extrinsic cells that promote T cell exhaustion by different mechanisms. Cytokines, including IL-10 and TGF-β, are both extrinsic factors involved in T cell exhaustion in the TME. GARP is shed from the surface of activated Tregs, which in its soluble form (sGARP) induces peripheral Tregs and TAMs with the M2 phenotype. Notably, proliferation of cytotoxic T cells and their effector functions are inhibited by sGARP. Increased expression of inhibitory receptors, including PD-1, CTLA-4, LAG-3, TIM-3, BTLA, and TIGIT, on CD8<sup>+</sup> T cells is the major intrinsic factor for T cell exhaustion. SHP-2, IRF-9, and AP-1 may affect PD-1 expression in the CD8<sup>+</sup> T cells.

#### 6.1.1. PD-1

PD-1 was first established as a negative regulator of T cell activation. Based on possession of a cytoplasmic immunoreceptor tyrosinebased inhibition motif (ITIM), this molecule is regarded as an inhibitory receptor of the immunoglobulin superfamily of receptors [73]. This inhibitory receptor is expressed on T and B cells, and some myeloid cells following their activation [74]. Following TCR engagement, PD-1 expresses and this expression eventually declines after antigen resolving, while during chronic antigen exposure, PD-1 expression is sustained. It has been shown that after co-culture of Jurkat cells with cancer cells, PD-1 expression is upregulated on the Jurkat cell surfaces. Although it is not well understood how PD-1 suppresses T cell functions in vivo, five different mechanisms have been proposed (Fig. 4). PD-1 may: 1) antagonize TCR signaling by recruiting phosphatases [11,75-78] 2) modulate the PI3K / AKT / mTOR pathway, implementing PD-1 in nutrient sensing, metabolism, survival, and cell growth [79-81], 3) modulate the Ras pathway, linking PD-1 to the cell cycle [81], 4) induce expression of basic leucine zipper transcription factor (BATF), which can repress expression of effector genes [82], and, 5) influence T cell motility [83-85]. A significant increase in PD-1 expression on the surface of CTLs in the TME and its correlation with reduction of effective cytokines has been documented in patients with lymphoma, Hodgkin disease, and liver and stomach cancers [73,86,87].

#### 6.1.2. CTLA-4

CTLA-4, an inhibitory receptor belonging to the CD28 protein family, is expressed on the T cell surface. Following connection to its ligands, CD80 and CD86, CTLA-4 exerts inhibitory messages that diminish IL-2 production, thus reducing T cell functionality [88,89]. In effector T cells, CTLA-4 competes strongly with CD28 for effective costimulation by CD80/86, leading to functional inactivation of the specific lymphocytes. CTLA-4 promotes the "stop signals" initiated by T cells upon encounter of the TCR with peptide-loaded MHC molecules [90]. Moreover, CTLA-4 induces *trans*-endocytosis of costimulatory ligands including CD80 and CD86, thus restricting opportunities for further T cell activation [91]. In the TME, 30–50% of CTLs simultaneously express CTLA-4 and PD-1. CTLs that express these two receptors are less capable of proliferation and cytokine production than those that do not, and are prone to exhaustion [92]. Stimulation of these inhibitory receptors can inhibit the AKT signaling pathway. The AKT pathway plays an important role in the regulation of glucose metabolism in T cells and increases glucose transporter 1 expression, resulting in increased glycolysis in these cells. Thus, activation of CTLA-4 and/or PD-1 leads to impaired glucose metabolism and reduced lymphocyte functionality, and eventually, exhaustion [80].

#### 6.1.3. TIM-3

Like PD-1, TIM-3 is transiently upregulated on virus-specific CTLs during acute infections, remains elevated on activated T cells, and may help to drive exhaustion under prolonged antigen exposure, which occurs during chronic infections and cancer [93–95]. In melanoma, co-expression of PD-1 and TIM-3 on exhausted CTL surfaces has been observed in the TME. Interestingly, cells with the TIM-3<sup>+</sup> PD-1<sup>+</sup> phenotype fail more in proliferation and production of typical cytokines such as IL-2, IFN<sub>γ</sub>, and TNF $\alpha$  than TIM-3<sup>-</sup> PD-1<sup>+</sup> and TIM-3<sup>-</sup> PD-1<sup>-</sup> cells; they also show more severe exhaustion [96].

#### 6.1.4. LAG-3

LAG-3, or CD223, is an MHC class-II ligand that is structurally similar to CD4 and expressed on activated and exhausted T cells. LAG-3 suppresses the expansion of CD4<sup>+</sup> T cells in response to antigen recognition [97] and was found to be synergistic with CTLA-4 and PD-1 in



**Fig. 4.** Inhibitory mechanisms of Programmed Cell Death-1 on T lymphocytes. PD-1 affects T cell function by various mechanisms. A) PD-1 directly antagonizes T cell receptor (TCR) signaling by preventing the phosphorylation of zetachain-associated protein kinase 70 (ZAP70); (B) PD-1 hinders the CD28-induced activation of phosphatidyl inositide 3-kinase (PI3K), leading to diminished AKT activation; (C) PD-1 inhibits the Ras pathway; (D) PD-1 upregulates the expression of suppressive transcription factors including basic leucine zipper transcription factor ATF-like (BATF); and (E) PD-1 may affect the contact duration between T cells and antigen-presenting cells (APCs).

mediating T cell suppression during chronic antigenic stimulation [98,99]. Additionally, LAG-3 is important in promoting the functionality of regulatory T cells [100]. LAG-3 has been shown to inhibit calcium fluxes associated with TCR signaling, and dampen cytokine production and lymphocyte proliferation [97]. However, it is believed that upregulated expression of LAG-3 on T cells alone may not be sufficient to drive them toward exhaustion, but it may cooperate with other inhibitory receptors to influence the extent of T cell exhaustion [101]. In the ovarian TME, it was shown that PD-1 and LAG-3 are simultaneously expressed on exhausted CTL surfaces, and lymphocytes with the LAG- $3^{+}$  PD-1<sup>+</sup> phenotype fail more in TNF $\alpha$  and IFN $\gamma$  production than LAG- $3^{-}$  PD-1<sup>+</sup> or LAG-3 ,PD-1<sup>-</sup> cells [58,102].

#### 6.1.5. BTLA

BTLA, an inhibitory receptor on T cell surfaces, through binding to herpes virus molecules, causes induction of inhibitory messages in CTLs. In advanced skin cancers, CTLs with  $BTLA^+$  PD-1 $^+$  TIM-3 $^+$  phenotypes were the most dysfunctional cells. So it seems that in skin cancers, BTLA plays an important role in CTL exhaustion [17,103].

#### 6.1.6. TIGIT

Recently, the role of the co-inhibitory receptor TIGIT was demonstrated in T cell exhaustion. TIGIT and CD226 molecules compete for binding to their CD112 and CD155 ligands, which is very similar to the binding of CTLA-4 to CD28. Notably, binding of CD226 to its ligands transfers positive activation signals while TIGIT binding leads to negative signals in T lymphocytes and their impairment [18,104]. It was shown that melanoma-specific CTLs highly express TIGIT. Furthermore, simultaneous blocking of PD-1 and TIGIT increases T cell proliferation and cytokine production, and enhances CTL performance, leading to the eventual elimination of tumor cells [105,106]. Overall, these findings demonstrate that the inhibitory molecules, including PD-1, have crucial roles in regulation of CTL exhaustion and PD-1 expression, and along with other inhibitory receptors, may determine the severity of CTL exhaustion.

#### 6.2. Transcription factors

Transcription factors including B lymphocyte-induced maturation

protein 1 (Blimp-1), T-bet, nuclear factor of activated T cells, cytoplasmic 1 (NFATc1), and BATF play substantial roles in CTL exhaustion in chronic infections [82,107,108]. However, the signaling pathways and transcription factors that affect CTL exhaustion in cancer are not yet clear. Downstream signaling pathways should also be considered. It has been demonstrated that Src homology 2 domain-containing phosphatase-2 (SHP2) plays an important role in the downstream signaling pathway of PD-1 [109]. On the other hand, PD-1 expression is upregulated on the surface of CTLs in Hodgkin's lymphoma. Blocking the PD-1 pathway affects the phosphorylation of tyrosine phosphatases such as SHP2, which consequently results in reduced IFN<sub>Y</sub> production, and hence, impaired lymphocyte function. Alpha interferon via IFNresponsive factor 9 (IRF9) signaling causes stable expression of PD-1 on T cell surfaces, indicating that IRF9 may induce T cell exhaustion [110]. Studies have also demonstrated that aberrant expression of the C-FOS subunit of the AP1 transcription factor in CTLs increases PD-1 expression through binding to the PD-1 promoter, which indicates the crucial role of C-FOS in the regulation of CTL exhaustion in the TME [111].

#### 7. Effective external signals in exhaustion of CTLs in the TME

The TME consists of stromal and inflammatory cells that express inhibitory ligands such as PD-L1 and PD-L2 enclosed with tumor cells and a network of regulatory cells and cytokines. This alters the metabolic status, lowers nutrient accessibility, and provides a complex arrangement of immunosuppressors, which limits cytotoxic activity CTLs and causes their dysfunction. In addition, the abundance of tumor antigens in this microenvironment leads to chronic T cell activation and probably CTL exhaustion [20]. Effective external signals in exhaustion of CTLs in the TME include the inhibitory ligands of PD-L1 and PD-L2, immunosuppressive cells, soluble factors, and environmental and metabolic conditions (Fig. 3).

#### 7.1. The inhibitory ligands PD-L1 and PD-L2

The main inhibitory ligands of PD-1 are PD-L1 and PD-L2. PD-L1 is highly expressed on stromal and cancer cell surfaces, and the PD-1/PD-L1 signaling pathway is considered as one of substantial regulatory routes of CTL exhaustion in the TME. Therefore, blocking this signaling pathway may augment the antitumor activity of CTLs [112]. PD-L2 is intermediately expressed on macrophages in response to cytokine activation, as well as dendritic cells (DCs) and mast cells. It was shown that coupling of these ligands to PD-1 drives T cells toward regulatory types [113].

#### 7.2. Immunosuppressive cells

A complex of immunosuppressor cells in the TME includes regulatory  $CD4^+$  T cells, plasmacytoid DCs, macrophages, and myeloidderived suppressor cells that directly or indirectly suppress CTL responses.

#### 7.2.1. Regulatory T CD4<sup>+</sup> lymphocytes

Regulatory CD4<sup>+</sup> T cells (Tregs) are defined as subsets of inhibitory T cells that in normal conditions maintain peripheral tolerance and prevent autoimmune diseases. These lymphocytes are abundant in peripheral blood and tumor tissues and may aid tumor cells to escape from immune responses [114,115]. These cells, through their functional ectoenzymes including CD39 and CD73, mediate production of extracellular adenosine, which may couple with adenosine A2A receptors on the cell membrane of effective CTLs and exert inhibitory effects [116,117]. By production of inhibitory cytokines such as IL-10 and TGF $\beta$ , and overexpression of CD25 (IL-2R), Tregs suppress CTL functions [118–120]. Furthermore, the increased IL-10 secretion from regulatory lymphocytes results in upregulated expression of PD-L1 on DC surfaces.

Glycoprotein A repetitions predominant (GARP), also known as LLRC32, which induces peripheral tolerance via TGF- $\beta$ , is an activation marker for Tregs. GARP is shed from activated Treg surfaces, and in its soluble form (sGARP), induces peripheral Tregs and M2 phenotype TAMs. Notably, cytotoxic T cell proliferation and their effector function is inhibited by sGARP [121].

#### 7.2.2. Plasmacytoid dendritic cells

Dendritic cells are distinct subsets of professional APCs. Plasmacytoid DCs, by production of indolamine 2, 3-dioxygenase (IDO), induce Tregs and eventually suppress immune responses in the TME [122]. It was shown that a population of plasmacytoid DCs in the TME of mice with prostate cancer expressed lower levels of the co-stimulatory ligands CD80, CD86, and CD40 and greater levels of the inhibitory ligands PD-L1 and IDO than non-cancerous mice. These findings reflect the crucial role of plasmacytoid DCs in the induction of CTL exhaustion [123].

#### 7.2.3. Macrophages

Macrophages play a vital role in innate immunity against foreign pathogens. M1 phenotypes are capable of producing a significant amount of pro-inflammatory cytokines, while M2 phenotypes, by producing several growth factors, are involved in tissue remodeling and control of innate immune responses. Macrophages that accumulate in the TME, known as TAMs, mainly demonstrate M2 phenotype and favor tumor establishment [124]. An increased number of these cells in the TME may worsen the prognosis. Tumor cells, by production of VEGF, CCL2, M-CSF, and angiopoietin 2, promote migration of bloodstream monocytes to the TME, which then differentiate to TAMs [125,126]. CCL2 overexpression in mice fibrocarcinoma cells results in recruitment of TAMs and assists tumorigenesis [62]. Tumor-associated macrophages, by production of the inhibitory cytokines IL-10 and TGF $\beta$ , suppress lymphocyte proliferation in the TME and also lead the immune responses toward conversion to Tregs [127].

#### 7.2.4. Myeloid-derived suppressor cells

Accumulation of MDSCs is a substantial mechanism of tumor promotion. These suppressor cells are progenitors of DCs, monocytes/ macrophages, and granulocytes [128]. In the case of human cancers they exhibit CD11b<sup>+</sup> CD33<sup>+</sup> CD34<sup>+</sup> CD14<sup>-</sup> HLA-DR<sup>-</sup> phenotype, while in mice tumors they exhibit CD11b<sup>+</sup> Gr-1<sup>+</sup> characteristics. MDSCs are commonly divided into the monocytic subgroup, with typical characteristics of CD11b<sup>+</sup> Ly6G<sup>low</sup> Ly6C<sup>high</sup>, and the granulocytic subgroup, with cellular characteristics of CD11b<sup>+</sup> Ly6G<sup>high</sup> Ly6C<sup>low</sup> [129]. By several mechanisms, MDSCs may inhibit the functionality of activated lymphocytes and induce CTL exhaustion. For example, in a mouse model of ovarian cancer, MDSCs highly express PD-L1 and CD80, which significantly inhibits the immune response against tumor antigens [130,131]. IL-10-producing cells, such as some DCs, may stimulate MDSCs to increase PD-L1 expression on their surfaces, which ultimately leads to CTL dysfunction through the PD-1/PD-L1 pathway, emphasizing the role of MDSCs in CTL exhaustion [132].

#### 7.3. Soluble factors (IL-10 and TGF- $\beta$ as inhibitory cytokines)

Several inhibitory cytokines contribute to CTL exhaustion. Among them IL-10 and TGFB could be released from tumor cells, TAMs, and Tregs in the TME and contribute to reduced effectiveness of local lymphocytes [127,133]. These cytokines can impair the cytotoxic effects of natural killer cells, which play a major role in innate immunity against tumors. They also may suppress macrophage and DC functions in the TME [134]. These cytokines may also upregulate PD-L1 expression on DCs [132], which represents a crucial step in CTL exhaustion [135]. Cancer cells, local fibroblasts, and several leukocytes are the main sources of TGFB [136]. TGFB shows various roles in tumor immunobiology, depending on cancer cell type and disease stage. In the early stages of cancer, TGF<sup>β</sup> hinders tumor cell growth and induces their apoptosis, however in the later stages  $TGF\beta$  may inhibit immune responses, upregulate cancer cell invasiveness, and increase the possibility of metastasis [137]. It was recently shown that TGF<sub>β</sub>-mediated transcription inhibition of lymphocyte effector molecules including perforin, granzyme, and cytokines, directly inhibits cytotoxic properties of CTLs [120,138]. Moreover, tumor cell-derived TGFB may increase miR-23a expression and reduce Blimp-1 expression and consequently inhibit CTL toxicity [139]. In addition, TGF<sub>β</sub>-treated naive T cells differentiate to Tregs, which have a major role in CTL exhaustion [140].

#### 7.4. Environmental and metabolic conditions

Researchers believe that differentiation status and metabolic programing determine the functional fate of T cells in the TME. As previously mentioned, changes in metabolic state of and nutrient availability to cancer cells may alter T cell function in the TME. Under physiological conditions, most nonmalignant cells rely on mitochondrial respiration, in which oxidative phosphorylation (OXPHOS) occurs as a primary metabolic pathway to generate energy in the form of adenosine triphosphate (ATP). But cancer cells, due to oncogenic mutations and dysfunction of the tumor suppressor genes, including p53, switch their metabolism to glycolysis, a biochemical process that occurs in the cytoplasm without the requirement for oxygen. This phenomenon, called the "Warburg effect," is a hallmark for cancer cell metabolism [141].

In cancer cells, glycolysis occurs in parallel with the tricarboxylic acid (TCA) cycle, which is linked to OXPHOS and oxidizes acetyl-CoA, and also other metabolic pathways to enhance the biosynthetic processes and thus support cancer cell proliferation and growth. Similar metabolic features were discovered in T lymphocytes during activation, even though this metabolic transition in T cells is part of a physiological process. Naive CTLs primarily use OXPHOS to produce ATP, but activated CTLs switch their metabolic program to glycolysis [142]. Signal transduction via the TCR and CD28 results in activation of phosphatidylinositol 3-kinase (PI3K), protein kinase B (Akt), and mammalian target of rapamycin (mTOR) pathways, which consequently may increase the activity of hypoxia-inducible factor (HIF)-1 $\alpha$  and Myc [143]. Upregulation of HIF-1 $\alpha$  results in increased expression of glucose

transporter protein (Glut1), which augments glucose uptake. PI3K activation decreases the expression of enzymes involved in the TCA cycle as well as OXPHOS, whereas Akt and Myc increase the activity of several glycolytic enzymes [144–146]. All these events induce CTLs to increase glycolysis after activation steps [147]. Glycolysis is less efficient than OXPHOS due to lower ATP production per glucose molecule, but generates ATP molecules faster than OXPHOS and supports differentiation and functionality of effector T cells [61]. Hypoxia, excessive consumption of essential metabolites such as glucose and essential amino acids, production of large amounts of lactic acid, fatty acids, reactive oxygen species (ROS) by cancer cells may lead to decreased functionality of effector TCD8<sup>+</sup> lymphocytes and suppression of antitumor immune responses in TME [142].

#### 7.4.1. Hypoxia

Hypoxia is low oxygen concentration status that commonly occurs following reduction of blood perfusion within tumoral tissue due to structural and functional alterations in the vessels, and also increased oxygen consumption, which refers to the uncontrolled proliferation of cancer cells. This results in insufficient delivery of oxygen and nutrients into the TME [148,149]. Different opinions were proposed about the effects of hypoxia on metabolism and CTL functions in the TME. Some researchers believe that HIF-1a, the main transcription factor that senses and responds to hypoxia, regulates CTL metabolism and suppresses their responses in the TME. Upon entry of CTLs to the TME and their confrontation with hypoxic conditions, HIF-1 $\alpha$  becomes activated. In hypoxia, HIF-1a, by increasing pyruvate dehydrogenase kinase 1 (PDK1) expression, inhibits mitochondrial respiration, prevents the oxidation of pyruvate to acetyl-CoA, and by promoting the activity of lactate dehydrogenase A (LDHa), enhances glycolysis for energy generation [150,151]. Furthermore, increased HIF-1 $\alpha$  activity inhibits the sustained Ca<sup>2+</sup>-NFAT pathway, which controls production of effector molecules in activated T cells [152]. Several in vitro and in vivo studies have shown that hypoxia and increased HIF-1 $\alpha$  activity suppress T cell activation, reducing their proliferation and decreasing their ability to produce necessary cytokines and lytic enzymes [153-157]. Additionally, hypoxia, due to increasing ROS accumulation, may induce apoptosis in activated CTLs [158,159]. Interestingly, some studies reported that cytokine production was increased in activated CTLs with a partial deficiency in HIF-1a [160,161]. Doedens et al. demonstrated that hypoxia or increased HIF-1a activity in CTLs may increase expression of co-inhibitors such as CTLA-4, LAG-3, and TIM-3, and decrease expression of T-bet, a key transcription factor that controls of T cell function. Moreover, these studies showed that hypoxia and increased HIF-1a activity may promote T cell effector functions, especially production of the proteolytic enzymes granzyme B and perforin. Of note, in these studies after 48 h of activation, CTLs were rested for several days in the IL2-supplemented medium before being subjected to hypoxia. Thus, maintenance of T cells in the IL-2-supplemented medium, due to metabolic reprograming and their decreased energy demand, may allow them to improve some functions in hypoxia [162]. Some other researchers believe that the effect of hypoxia or HIF-1 $\alpha$  is limited to CTLs. They confirmed that deletion of VHL, a protein involved in the ubiquitination and degradation of HIF, may lead to HIF upregulation, resulting in differentiation of cytotoxic CD8<sup>+</sup> cells (C-TLs), while depletion of HIF-1 $\alpha$  reduces CTL function [61]. Hypoxia, in addition to its direct effect on CTL responses in the TME through increased surface expression of PD-L1 on cancer cells and enhanced the suppressive activity of tumor-infiltrating myeloid suppressor cells and tumor-associated macrophages, impairs PD-1<sup>+</sup> CTL survival and function [163–165].

#### 7.4.2. Hypoglycemia and lactic acid

Glucose is critical to the survival, growth, and differentiation of T lymphocytes. Several studies in vitro and in vivo have demonstrated that hypoglycemia suppresses the effector functions of CTLs. High competition for glucose intake by cancer cells and activated T lymphocytes leads to hypoglycemia in the TME. Activated CTLs express increased levels of Glut1 to increase glucose uptake, but this effect may be neutralized by tumor cells. Consequently, impaired naive CTL activation may dampen their effectiveness and decrease generation of effector cytokines, even though some levels of proliferation may be preserved through OXPHOS [62,166–171].

The mTOR pathway and AMP-activated protein kinase (AMPK) are central energy monitoring systems of cells. Disorders of mTOR and AMPK activity are key signals that merge metabolic activity with cell activation and differentiation in T cells [146,151].

Hypoglycemia exerts its immunosuppressive effects via increasing PD-1 expression and decreasing mTOR signaling in activated CTLs, which lead to reduced glycolysis, enhanced fatty acid metabolism, and diminished IFN- $\gamma$  and IL-2 production [63], as well as blockade of PD-1, resulting in augmentation of the glycolytic capacity through increased mTOR signaling and improvement of CTL function in the TME [62].

Glucose deprivation, with enhanced AMP and alteration of the AMP/ATP cellular ratio, leads to activation of AMP-activated protein kinase (AMPK) as an energy sensor in activated CTLs. AMPK, through inhibition of the mTOR pathway, decreases glycolysis and anabolic processes, while enhancement of OXPHOS by fatty acids and glutamine, as well as AMPK via blocking cytokine production, decreases T cell energy expenditure [172,173]. Furthermore, AMPK activation promotes Treg formation in both in vitro and in vivo conditions [174,175].

The glycolytic metabolite phosphoenolpyruvate (PEP) promotes T cell activation by sustaining TCR-mediated Ca<sup>2+</sup>-NFAT signaling and suppressing sarco/ER Ca<sup>2+</sup>-ATPase (SERCA) activity. SERCA is a Ca<sup>2+</sup> ATPase that transfers Ca<sup>2+</sup> from the cell cytosol to the sarcoplasmic reticulum (SR) lumen, increasing T cell effector functions [170].

Hypoglycemia due to high glucose flux in cancer cells results in limited glucose availability for T cell utilization. As a consequence, T cells not only are unable to develop tumoricidal effects, but also alter their differentiation program resulting in the generation of cell types that develop due to limited glucose supplies, such as Tregs and exhausted T lymphocytes [174,176].

Although increased glycolysis in activated CTLs causes lactic acid production and release, PD-1 signaling in these cells reduces glycolysis and lactic acid production; therefore, tumor cells are most likely the main source of lactic acid in the TME [176]. Increasing glycolysis in cancer cells produces excess lactic acid resulting in microenvironmental acidification. Microenvironmental acidification can: (1) suppress proliferation, cytokine production, and cytotoxic activity of CTLs [65,177], (2) alter macrophage polarization and shift them to the M2 suppressive phenotype [178], and (3) induce arginase 1 (Arg1), which promotes the depletion of extracellular arginine levels, resulting in inhibition of efficient T cell proliferation and activation [178,179]. Increasing lactic acid concentration in CTLs causes local acidification, which inhibits the activity of phosphofructokinase, a key glycolytic enzyme [177].

#### 7.4.3. Fatty acids and cholesterol metabolism

Accumulation of adipocytes and adipocyte-like fibroblasts and production of large amounts of fatty acids by tumor cells indicate that the TME could be lipid-enriched [67,180,181]. Excessive amounts of fatty acids lead to leukocyte dysfunction in the TME. Many studies have shown that tumor-infiltrating MDSCs and DCs with abnormally high lipid content are respectively associated with strong immunosuppressive activity and weakened antigen presentation [182–185]. The fatty acid-enriched TME may affect effector T cell function, promote Treg development, and shift macrophages to the M2 suppressive phenotype [5,186].

Cholesterol is an important component of the plasma membrane in mammalian cells that often clusters in lipid rafts. Lipid rafts cluster at TCR-rich regions of T cells, thus cholesterol is regarded as a critical factor for TCR signaling [187]. Free cholesterol can be esterified by the cholesteryl esters ACAT1 and ACAT2, which can help their intracellular storage [188]. Yang et al. proposed a mechanism by which the antitumor response of mouse CTLs can be potentiated via modulating cholesterol metabolism. Inhibiting cholesterol esterification in T cells by genetic ablation or pharmacological inhibition of ACAT1 leads to enhanced functionality and proliferation of CD8+, but not CD4+, T cells. This is probably due to selective increment of cholesterol in CTL plasma membranes, which causes enhanced TCR clustering and signaling, as well as increased immunological synapse formation. CTLs controlled melanoma growth and metastasis in ACAT1-deficient mice better than wild-type cells. Yang et al. indicated that avasimibe, an ACAT inhibitor that was previously applied in clinical trials to treat atherosclerosis, and also to treat melanoma in mice, has proved safe in humans, thus it may be an effective human antitumoric. Recently, a combined therapy of avasimibe plus an anti-PD-1 antibody showed greater efficacy than monotherapies in control of tumor progression [189].

#### 7.4.4. Amino acid depletion

Tryptophan, an essential amino acid, has a crucial role in cancer cell survival. Overexpression of IDO, a metabolic enzyme that degrades tryptophan to kynurenine via reduced infiltration of tryptophan to Tlymphocytes, may suppress lymphocyte proliferation. Furthermore, kynurenine production promotes Tregs in the TME [65,190]. Cancer cells, MDSCs, and M2 type macrophages, through increased expression of Arg1, which degrades arginine, may have reduced arginine levels in the TME [178,191]. Glutamine is also required for T cell differentiation and function. This amino acid is also utilized by lymphocytes as an essential nutrient and helps in effector CTL development [172,192]. In several cancer types, enhanced glutamine metabolism, due to mutations and altered signaling pathways, may result in its decreased availability. Therefore, T cells encounter glutamine-deprived conditions in the TME [66]. Overall, excessive amino acid metabolism by cancer cells causes amino acid paucity in the TME, which can suppress CTL anti-tumor responses in this region.

#### 8. The rapeutic interventions to restore exhausted CTL function in the $\ensuremath{\mathsf{TME}}$

Recently, cancer immunotherapy via inhibitory receptor targeting by specific antibodies has been a major breakthrough. Blocking inhibitory receptors with specific antibodies may reverse exhausted T cell functions and anti-tumor responses. Currently-available antibodies that block T cell inhibitory receptors target CTLA-4, PD-1, and PD-L1 (Table 2). Among CTLA-4 blocking monoclonal antibodies, Tremelimumab and Ipilimumab are the most studied. Early-phase clinical trials demonstrated that Tremelimumab treatment resulted in sustained antitumor responses [193]. Ipilimumab was approved in 2011 by the American Food and Drug Administration (FDA) for the treatment of patients with advanced skin cancer [194,195], and is currently under study for treatment of patients with metastatic prostate cancer. As previously mentioned, blocking the PD-1-PD-L1 pathway could restore exhausted CTL functions and increase their anti-tumor responses. For this purpose, various antibodies have been produced and applied. Nivolumab was the first monoclonal anti-PD-1 antibody used to treat patients with advanced skin, kidney, and lung cancers; unfortunately, this antibody was only effective for up to one year and could not be applied for prolonged treatments [112, 196]. Pembrolizumab, another anti-PD-1 monoclonal antibody, in the first phase of clinical trials, caused satisfactory anti-tumor responses and no toxic side effects on T lymphocytes were seen [197]. Similarly, Pidilizumab, another anti-PD-1 monoclonal antibody, was initially used to treat hematologic malignancies and generated some steady-state responses [198]. In other studies, humanized monoclonal antibodies, including BMS-936559 and MPDL3280A, were used to block PD-L1. In clinical trials BMS-936559 led to long-term tumor regression in patients with kidney, lung, ovarian and skin cancers, caused limited side effects, and was well tolerated by

Table 2

Therapeutic interventions for blocking immune checkpo
---

Antibody	Target	Status of clinical trial	Cancer type	Refs.
Tremelimumab	CTLA-4	Phase II	Mesothelioma	[188]
Ipilimumab	CTLA-4	FDA approved Phase II and III	Melanoma	[189,190]
			Solid tumors	
Nivolumab	PD-1	Phases I and II	Solid tumors, melanoma, NSCLC, RCC, ovarian cancer	[65,191]
Pembrolizumab	PD-1	Phase I	Melanoma, NSCLC, head and neck cancer	[192]
Pidilizumab	PD-1	Phases I and II	Hematologic malignancies	[66]
BMS-936559	PD-L1	Phase I	Solid tumors	[193]
MPDL3280A	PD-L1	Phase I	Solid tumors, melanoma, NSCLC, bladder cancer	[194]
MEDI4736	PD-L1	Phase I	Solid tumors, melanoma, head and neck cancer, gastric cancer	[195]

*Abbreviations*: CTLA-4, cytotoxic T lymphocyte antigen-4; NSCLC, non-small cell lung cancer; PD-1, programmed cell death protein 1; PD-L1, programmed cell death1 ligand 1; RCC, renal cell carcinoma.

patients [199]. Similarly, MPDL3280A was antitumoric in the treatment of prostate cancer and other advanced and metastatic solid tumors [200]. MEDI4736, another PD-L1 blocking antibody showed acceptable results in early stage clinical trials for treatment of advanced solid tumors; moreover, molecular engineering of the Fc region of this antibody improved results [201]. Currently, combinations of blocking antibodies against inhibitory receptors have shown promising results in advance stage cancer treatments. For example, a combination of Pidilizumab and Rituximab is, chimeric monoclonal antibody against CD20, recommended for the treatment of follicular lymphoma [113], a combination of Nivolumab and Ipilimumab has been used to treat advanced skin cancers [202], and a combination of Pembrolizumab and Ipilmumab has been approved for the treatment of malignant skin cancers [203]. Overall, combination therapies have been shown to be more effective against tumors than monoclonal monotherapies.

Another cancer immunotherapy approach is application of CAR T cells. In this method T cells from peripheral blood are modified to express chimeric antigen receptors (CARs), which recognize cell surfaceexpressed tumor-associated antigens independent from the major histocompatibility complex and also co-stimulatory molecules, enhancing T cell anti-tumor responses [204]. The therapeutic outcomes of CAR T cells on acute lymphatic leukemia and B cell lymphoma were acceptable; however, in patients with solid tumors the results were disappointing. It is believed the TME can reduce CAR T cell-induced antitumor immunity [205-209]. Reducing the number of Tregs, MDSCs, or TAMs, or suppressing their functions as well as blocking the immune checkpoints, could improve the efficacy of CAR T cells in cancer immunotherapy [210,211]. Interestingly, CAR T cells, combined with PD-1 blockade, strongly augmented anti-tumor responses [212], Laboratory results confirmed that inhibitory checkpoint pathways such as PD-1 and CTLA-4 signaling can modify the metabolic programs of T cells [77,176]. PD-1-dependent signaling may inhibit glucose and glutamine transport and hinder hexokinase 2, which catalyzes the first step of glycolysis, leading to reduced glycolysis and amino acid metabolism in T cells. However, this signals through inhibiting the lipid oxidation PI3K pathway induce lipolysis and decrease lipid biosynthesis that increase the rate of fatty acid oxidation (FAO) in T lymphocytes

[176,213]. Increasing FAO after receiving PD-1 signals in T cells causes longevity of these cells in patients with chronic infections and cancer, thus PD-1 ligation alters the metabolic programming of T lymphocytes by inhibiting glycolysis and promoting FAO. In contrast, CTLA-4 signals without FAO augmentation may decrease glycolysis in T cells only through inhibiting expression of the glucose and glutamine transporters. It seems that CTLA-4 ligation maintains immune tolerance by preserving the metabolic profile of T lymphocytes [214]. As previously mentioned, the metabolic state of the tumor cells has a significant influence on longevity and functional fate of CTLs by altering nutrient availability and modifying their metabolic profile before their infiltration to the TME. Some studies revealed that pharmaceutical targeting of the glycolytic pathway in tumor cells by inhibitors of GLUTs. HK. PKM2, or LDHA may lead to tumor regression and increased glucose sources within the TME [215,216]. Chang et al. indicated that blocking PD-L1 reduces glucose consumption through suppressed glycolysis in tumor cells and increases glycolysis and effective function of CTLs in the TME [62]. Ho et al. showed that a glucose-deficient TME leads to reduction of phosphoenolpyruvate (PEP), a glycolytic metabolite essential for  $Ca^{\bar{2}+}$ -NFAT signaling in CTLs, and increases the expression of phosphoenolpyruvate carboxykinase 1 (PCK1), which converts the TCA cycle intermediate oxaloacetate to PEP, augments NFAT signaling, and improves CTLs function within the TME [170]. On the other hand, some studies indicated that inhibition of glycolysis in CTLs increases their efficacy. During their activation naive CTLs switch their metabolic program to glycolysis, which may lead to terminal differentiation and shorten their survival. Thus, reducing glycolysis in these cells may improve therapeutic effects [217]. For example, inhibition of glycolysis in CTLs by 2-deoxyglucose enhances their anti-tumor function in a mouse melanoma model [218]. Furthermore, a series of studies demonstrated that CTLs, when treated with IL-7 or IL-15 in vitro, differentiate toward memory and enhancement of anti-tumor function in vivo [219-221]. Also in adoptive transfer condition, central memory CTLs have stronger anti-tumor responses than effector CD8<sup>+</sup> T lymphocytes [222,223]. We think that use of FAO and OXPHOS as the primary metabolic pathway in memory CTLs causes their superior performance within the TME.

#### 9. Conclusion

Tumor cells utilize various mechanisms to overcome patients' immune systems. The presence and functions of specific T cells in the TME are directly related to the fate of tumor cells and subsequent patient prognosis. Numerous mechanisms limit T cell activity and tumor cell survival. Lack of nutrients, increased metabolic waste, and expression of inhibitory ligands by tumors lead to reduced metabolic fitness and decrease the ability of T cells to import nutrients. Moreover, immune suppressive agents in the TME lead to T cell dysfunction characterized by exhaustion and Treg phenotypes. In the TME, exhaustion is an important mechanism that reduces the cytotoxic function of CTLs and induces secretion of regulatory cytokines. Currently, restoring the function of these cells and increasing the antitumor response is regarded as a novel strategy in cancer treatment. Blocking inhibitory receptors, especially PD-1 and PD-L1, with monoclonal antibodies, has opened a new perspective in cancer treatment. Application of combination therapy has improved results in this area. In the case of using a singular blocking antibody, some types of exhausted CTLs change their phenotypes to effective ones, while simultaneous blocking of multiple inhibitory receptors resulted in improved outcomes. However, limitations exist in this therapeutic approach: first, the various functions of inhibitory receptors are not yet well known, for example, PD-1 and TIM-3 may regulate as yet undefined pathways in CTLs, second, restoring CTL function may intensify their cytotoxic activity, and third, blocking each inhibitory receptor alone had little effect, while application of combination therapies was more effective.

On the other hand, drugs that directly target key metabolic enzymes or their upstream regulators will likely effect metabolism of both cancer and T cells. Thus, understanding the similar and different metabolic processes of the two cell types may help to develop therapies that simultaneously improve anti-tumor responses while eradicating tumor cells. We have only begun to understand how the TME effects cancer and its treatment options, but we are confident that restoring the function of exhausted CTLs can be considered as a promising therapeutic approach for cancer treatment.

#### References

- F. Chen, X. Zhuang, L. Lin, P. Yu, Y. Wang, Y. Shi, et al., New horizons in tumor microenvironment biology: challenges and opportunities, BMC Med. 13 (1) (2015) 45.
- [2] D. Hanahan, R.A. Weinberg, The hallmarks of cancer, Cell 100 (1) (2000) 57–70.
  [3] D. Hanahan, L.M. Coussens, Accessories to the crime: functions of cells recruited to
- the tumor microenvironment, Cancer Cell 21 (3) (2012) 309-322.
- [4] V. Gkretsi, A. Stylianou, P. Papageorgis, C. Polydorou, T. Stylianopoulos, Remodeling components of the tumor microenvironment to enhance cancer therapy, Front. Oncol. 5 (2015).
- [5] N.J. MacIver, R.D. Michalek, J.C. Rathmell, Metabolic regulation of T lymphocytes, Annu. Rev. Immunol. 31 (2013) 259–283.
- [6] A.J. Freemerman, A.R. Johnson, G.N. Sacks, J.J. Milner, E.L. Kirk, M.A. Troester, et al., Metabolic reprogramming of macrophages glucose transporter 1 (GLUT1)mediated glucose metabolism drives a proinflammatory phenotype, J. Biol. Chem. 289 (11) (2014) 7884–7896.
- [7] E.J. Pearce, B. Everts, Dendritic cell metabolism, Nat. Rev. Immunol. 15 (1) (2015) 18–29.
- [8] G. Verdeil, S.A.F. Marraco, T. Murray, D.E. Speiser, From T cell "exhaustion" to anti-cancer immunity, Biochim. Biophys. Acta (BBA) – Rev. Cancer 1865 (1) (2016) 49–57.
- [9] L. Baitsch, S.A. Fuertes-Marraco, A. Legat, C. Meyer, D.E. Speiser, The three main stumbling blocks for anticancer T cells, Trends Immunol. 33 (7) (2012) 364–372.
- [10] W. Zou, Immunosuppressive networks in the tumour environment and their therapeutic relevance, Nat. Rev. Cancer 5 (4) (2005) 263.
- [11] M. Vanneman, G. Dranoff, Combining immunotherapy and targeted therapies in cancer treatment, Nat. Rev. Cancer 12 (4) (2012) 237–251.
- [12] E.J. Wherry, J.N. Blattman, K. Murali-Krishna, R. Van Der Most, R. Ahmed, Viral persistence alters CD8 T-cell immunodominance and tissue distribution and results in distinct stages of functional impairment, J. Virol. 77 (8) (2003) 4911–4927.
- [13] D.L. Barber, E.J. Wherry, D. Masopust, B. Zhu, J.P. Allison, A.H. Sharpe, et al., Restoring function in exhausted CD8 T cells during chronic viral infection, Nature 439 (7077) (2006) 682.
- [14] H.-T. Jin, A.C. Anderson, W.G. Tan, E.E. West, S.-J. Ha, K. Araki, et al., Cooperation of Tim-3 and PD-1 in CD8 T-cell exhaustion during chronic viral infection, Proc. Natl. Acad. Sci. 107 (33) (2010) 14733–14738.
- [15] A. Crawford, E.J. Wherry, The diversity of costimulatory and inhibitory receptor pathways and the regulation of antiviral T cell responses, Curr. Opin Immunol. 21 (2) (2009) 179–186.
- [16] S.D. Blackburn, H. Shin, W.N. Haining, T. Zou, C.J. Workman, A. Polley, et al., Coregulation of CD8 + T cell exhaustion by multiple inhibitory receptors during chronic viral infection, Nat. Immunol. 10 (1) (2009) 29–37.
- [17] J. Fourcade, Z. Sun, O. Pagliano, P. Guillaume, I.F. Luescher, C. Sander, et al., CD8+ T cells specific for tumor antigens can be rendered dysfunctional by the tumor microenvironment through upregulation of the inhibitory receptors BTLA and PD-1, Cancer Res. 72 (4) (2012) 887–896.
- [18] N. Joller, J.P. Hafler, B. Brynedal, N. Kassam, S. Spoerl, S.D. Levin, et al., Cutting edge: TIGIT has T cell-intrinsic inhibitory functions, J. Immunol. 186 (3) (2011) 1338–1342.
- [19] E.J. Wherry, T cell exhaustion, Nat. Immunol. 12 (6) (2011) 492–499.
- [20] Y. Jiang, Y. Li, B. Zhu, T-cell exhaustion in the tumor microenvironment, Cell Death Dis. 6 (6) (2015) e1792.
- [21] R.H. Schwartz, A cell culture model for T lymphocyte clonal anergy, Science 248 (4961) (1990) 1349–1356.
- [22] R.H. Schwartz, T cell anergy, Annu. Rev. Immunol. 21 (2003) 305–334.
- [23] A.D. Wells, New insights into the molecular basis of T cell anergy: anergy factors, avoidance sensors, and epigenetic imprinting, J. Immunol. 182 (12) (2009) 7331–7341.
- [24] J. Reiser, A. Banerjee, Effector, Memory, and Dysfunctional CD8(+) T Cell Fates in the Antitumor Immune Response, J. Immunol. Res. 2016 (2016) 8941260.
- [25] E.J. Wherry, M. Kurachi, Molecular and cellular insights into T cell exhaustion, Nat. Rev. Immunol. 15 (8) (2015) 486–499.
- [26] W. Zou, L. Chen, Inhibitory B7-family molecules in the tumour microenvironment, Nat. Rev. Immunol. 8 (6) (2008) 467.
- [27] C. Blank, I. Brown, A.C. Peterson, M. Spiotto, Y. Iwai, T. Honjo, et al., PD-L1/B7H-1 inhibits the effector phase of tumor rejection by T cell receptor (TCR) transgenic CD8 + T cells, Cancer Res. 64 (3) (2004) 1140–1145.
- [28] T.J. Curiel, S. Wei, H. Dong, X. Alvarez, P. Cheng, P. Mottram, et al., Blockade of B7–H1 improves myeloid dendritic cell-mediated antitumor immunity, Nat. Med. 9 (5) (2003) 562.
- [29] I. Kryczek, L. Zou, P. Rodriguez, G. Zhu, S. Wei, P. Mottram, et al., B7-H4

expression identifies a novel suppressive macrophage population in human ovarian carcinoma, J. Exp. Med. 203 (4) (2006) 871–881.

- [30] L. Chen, S. Ashe, W.A. Brady, I. Hellström, K.E. Hellström, J.A. Ledbetter, et al., Costimulation of antitumor immunity by the B7 counterreceptor for the T lymphocyte molecules CD28 and CTLA-4, Cell 71 (7) (1992) 1093–1102.
- [31] T.F. Gajewski, B7–1 but not B7–2 efficiently costimulates CD8+ T lymphocytes in the P815 tumor system in vitro, J. Immunol. 156 (2) (1996) 465–472.
- [32] C. Blank, J. Kuball, S. Voelkl, H. Wiendl, B. Becker, B. Walter, et al., Blockade of PD-L1 (B7–H1) augments human tumor-specific T cell responses in vitro, Int. J. Cancer 119 (2) (2006) 317–327.
- [33] I.E. Brown, C. Blank, J. Kline, A.K. Kacha, T.F. Gajewski, Homeostatic proliferation as an isolated variable reverses CD8 + T cell anergy and promotes tumor rejection, J. Immunol. 177 (7) (2006) 4521–4529.
- [34] M.R. Nazareth, L. Broderick, M.R. Simpson-Abelson, R.J. Kelleher, S.J. Yokota, R.B. Bankert, Characterization of human lung tumor-associated fibroblasts and their ability to modulate the activation of tumor-associated T cells, J. Immunol. 178 (9) (2007) 5552–5562.
- [35] L. Broderick, S.P. Brooks, H. Takita, A.N. Baer, J.M. Bernstein, R.B. Bankert, IL-12 reverses anergy to T cell receptor triggering in human lung tumor-associated memory T cells, Clin. Immunol. 118 (2) (2006) 159–169.
- [36] M. Adibzadeh, H. Pohla, A. Rehbein, G. Pawelec, Long-term culture of monoclonal human T lymphocytes: models for immunosenescence? Mech. Ageing Dev. 83 (3) (1995) 171–183.
- [37] R.B. Effros, Replicative senescence in the immune system: impact of the Hayflick Limit on T-cell function in the elderly, Am. J. Hum. Genet. 62 (5) (1998) 1003–1007.
- [38] L. Hayflick, P.S. Moorhead, The serial cultivation of human diploid cell strains, Exp. Cell Res. 25 (3) (1961) 585–621.
- [39] J.F. Passos, G. Nelson, C. Wang, T. Richter, C. Simillion, C.J. Proctor, et al., Feedback between p21 and reactive oxygen production is necessary for cell senescence, Mol. Syst. Biol. 6 (1) (2010) 347.
- [40] F. Rodier, J.-P. Coppé, C.K. Patil, W.A. Hoeijmakers, D.P. Muñoz, S.R. Raza, et al., Persistent DNA damage signaling triggers senescence-associated inflammatory cytokine secretion, Nat. Cell Biol. 11 (8) (2009) 973.
- [41] C.M. Beauséjour, A. Krtolica, F. Galimi, M. Narita, S.W. Lowe, P. Yaswen, et al., Reversal of human cellular senescence: roles of the p53 and p16 pathways, The EMBO J. 22 (16) (2003) 4212–4222.
- [42] V. Appay, D.F. Nixon, S.M. Donahoe, G.M. Gillespie, T. Dong, A. King, et al., HIV-specific CD8 + T cells produce antiviral cytokines but are impaired in cytolytic function, J. Exp. Med. 192 (1) (2000) 63–76.
- [43] A.N. Akbar, S.M. Henson, Are senescence and exhaustion intertwined or unrelated processes that compromise immunity? Nat. Rev. Immunol. 11 (4) (2011) 289.
- [44] G. Pawelec, R. Solana, Immunoageing-the cause or effect of morbidity? Trends Immunol. 22 (7) (2001) 348–349.
- [45] A.N. Vallejo, C.M. Weyand, J.J. Goronzy, T-cell senescence: a culprit of immune abnormalities in chronic inflammation and persistent infection, Trends Mol. Med. 10 (3) (2004) 119–124.
- [46] C.L. Montes, A.I. Chapoval, J. Nelson, V. Orhue, X. Zhang, D.H. Schulze, et al., Tumor-induced senescent T cells with suppressor function: a potential form of tumor immune evasion, Cancer Res. 68 (3) (2008) 870–879.
- [47] F. Meloni, M. Morosini, N. Solari, I. Passadore, C. Nascimbene, M. Novo, et al., Foxp3 expressing CD4+ CD25+ and CD8+ CD28- T regulatory cells in the peripheral blood of patients with lung cancer and pleural mesothelioma, Hum. Immunol. 67 (1) (2006) 1–12.
- [48] T. Tsukishiro, A.D. Donnenberg, T.L. Whiteside, Rapid turnover of the CD8 + CD28-T-cell subset of effector cells in the circulation of patients with head and neck cancer, Cancer Immunol. Immunother. 52 (10) (2003) 599–607.
- [49] H. Li, K. Wu, K. Tao, L. Chen, Q. Zheng, X. Lu, et al., Tim-3/galectin-9 signaling pathway mediates T-cell dysfunction and predicts poor prognosis in patients with hepatitis B virus-associated hepatocellular carcinoma, Hepatology 56 (4) (2012) 1342–1351.
- [50] J.M. Brenchley, N.J. Karandikar, M.R. Betts, D.R. Ambrozak, B.J. Hill, L.E. Crotty, et al., Expression of CD57 defines replicative senescence and antigen-induced apoptotic death of CD8+ T cells, Blood 101 (7) (2003) 2711–2720.
- [51] M. Heffner, D.T. Fearon, Loss of T cell receptor-induced Bmi-1 in the KLRG1 + senescent CD8 + T lymphocyte, Proc. Natl. Acad. Sci. 104 (33) (2007) 13414–13419.
- [52] D. Voehringer, C. Blaser, P. Brawand, D.H. Raulet, T. Hanke, H. Pircher, Viral infections induce abundant numbers of senescent CD8 T cells, J. Immunol. 167 (9) (2001) 4838–4843.
- [53] J. Fourcade, Z. Sun, M. Benallaoua, P. Guillaume, I.F. Luescher, C. Sander, et al., Upregulation of Tim-3 and PD-1 expression is associated with tumor antigen–specific CD8 + T cell dysfunction in melanoma patients, J. Exp. Med. 207 (10) (2010) 2175–2186.
- [54] X. Huang, X. Bai, Y. Cao, J. Wu, M. Huang, D. Tang, et al., Lymphoma endothelium preferentially expresses Tim-3 and facilitates the progression of lymphoma by mediating immune evasion, J. Exp. Med. (2010) jem. 20090397.
- [55] T. Van Nguyen, N. Puebla-Osorio, H. Pang, M.E. Dujka, C. Zhu, DNA damageinduced cellular senescence is sufficient to suppress tumorigenesis: a mouse model, J. Exp. Med. 204 (6) (2007) 1453–1461.
- [56] K.E. Pauken, E.J. Wherry, Overcoming T cell exhaustion in infection and cancer, Trends Immunol. 36 (4) (2015) 265–276.
- [57] T. Zenz, Exhausting T cells in CLL, Blood 121 (9) (2013) 1485-1486.
- [58] J. Matsuzaki, S. Gnjatic, P. Mhawech-Fauceglia, A. Beck, A. Miller, T. Tsuji, et al., Tumor-infiltrating NY-ESO-1–specific CD8+ T cells are negatively regulated by LAG-3 and PD-1 in human ovarian cancer, Proc. Natl. Acad. Sci. 107 (17) (2010)

7875-7880.

- [59] P.A. Savage, D.S. Leventhal, S. Malchow, Shaping the repertoire of tumor-infiltrating effector and regulatory T cells, Immunol. Rev. 259 (1) (2014) 245–258.
- [60] P.S. Kim, R. Ahmed, Features of responding T cells in cancer and chronic infection, Curr. Opin. Immunol. 22 (2) (2010) 223–230.
- [61] Y. Zhang, H.C. Ertl, Starved and asphyxiated: How can CD8 + T cells within a tumor microenvironment prevent tumor progression, Front. immunol. 7 (2016).
- [62] C.-H. Chang, J. Qiu, D. O'Sullivan, M.D. Buck, T. Noguchi, J.D. Curtis, et al., Metabolic competition in the tumor microenvironment is a driver of cancer progression, Cell 162 (6) (2015) 1229–1241.
- [63] C.-H. Chang, J.D. Curtis, L.B. Maggi, B. Faubert, A.V. Villarino, D. O'Sullivan, et al., Posttranscriptional control of T cell effector function by aerobic glycolysis, Cell 153 (6) (2013) 1239–1251.
- [64] A.N. Mendler, B. Hu, P.U. Prinz, M. Kreutz, E. Gottfried, E. Noessner, Tumor lactic acidosis suppresses CTL function by inhibition of p38 and JNK/c-Jun activation, Int. J. Cancer 131 (3) (2012) 633–640.
- [65] J.D. Mezrich, J.H. Fechner, X. Zhang, B.P. Johnson, W.J. Burlingham, C.A. Bradfield, An interaction between kynurenine and the aryl hydrocarbon receptor can generate regulatory T cells, J. Immunol. 185 (6) (2010) 3190–3198.
- [66] C.T. Hensley, A.T. Wasti, R.J. DeBerardinis, Glutamine and cancer: cell biology, physiology, and clinical opportunities, J. Clin. Invest. 123 (9) (2013) 3678.
- [67] E. Currie, A. Schulze, R. Zechner, T.C. Walther, R.V. Farese, Cellular fatty acid metabolism and cancer, Cell Metab. 18 (2) (2013) 153–161.
- [68] K.E. Pauken, M.A. Sammons, P.M. Odorizzi, S. Manne, J. Godec, O. Khan, et al., Epigenetic stability of exhausted T cells limits durability of reinvigoration by PD-1 blockade, Science 354 (6316) (2016) 1160–1165.
- [69] B. Lu, O.J. Finn, T-cell death and cancer immune tolerance, Cell Death Differ. 15 (1) (2008) 70–79.
- [70] F. Shi, M. Shi, Z. Zeng, R.Z. Qi, Z.W. Liu, J.Y. Zhang, et al., PD-1 and PD-L1 upregulation promotes CD8(+) T-cell apoptosis and postoperative recurrence in hepatocellular carcinoma patients, Int. J. Cancer 128 (4) (2011) 887–896.
- [71] N. Murakami, L.V. Riella, Co-inhibitory pathways and their importance in immune regulation, Transplantation 98 (1) (2014) 3–14.
- [72] T. Maj, S. Wei, T. Welling, W. Zou, T cells and costimulation in cancer, Cancer J. 19 (6) (2013) 473–482.
- [73] M. Ahmadzadeh, L.A. Johnson, B. Heemskerk, J.R. Wunderlich, M.E. Dudley, D.E. White, et al., Tumor antigen-specific CD8 T cells infiltrating the tumor express high levels of PD-1 and are functionally impaired, Blood 114 (8) (2009) 1537–1544.
- [74] M.E. Keir, M.J. Butte, G.J. Freeman, A.H. Sharpe, PD-1 and its ligands in tolerance and immunity, Annu. Rev. Immunol. 26 (2008) 677–704.
- [75] T. Yokosuka, M. Takamatsu, W. Kobayashi-Imanishi, A. Hashimoto-Tane, M. Azuma, T. Saito, Programmed cell death 1 forms negative costimulatory microclusters that directly inhibit T cell receptor signaling by recruiting phosphatase SHP2, J. Exp. Med. 209 (6) (2012) 1201–1217.
- [76] J.M. Chemnitz, R.V. Parry, K.E. Nichols, C.H. June, J.L. Riley, SHP-1 and SHP-2 associate with immunoreceptor tyrosine-based switch motif of programmed death 1 upon primary human T cell stimulation, but only receptor ligation prevents T cell activation, J. immunol. 173 (2) (2004) 945–954.
- [77] J.L. Riley, PD-1 signaling in primary T cells, Immunol. Rev. 229 (1) (2009) 114–125.
- [78] K.A. Sheppard, L.J. Fitz, J.M. Lee, C. Benander, J.A. George, J. Wooters, et al., PD-1 inhibits T-cell receptor induced phosphorylation of the ZAP70/CD3zeta signalosome and downstream signaling to PKCtheta, FEBS Lett. 574 (1–3) (2004) 37–41.
- [79] M.M. Staron, S.M. Gray, H.D. Marshall, I.A. Parish, J.H. Chen, C.J. Perry, et al., The transcription factor FoxO1 sustains expression of the inhibitory receptor PD-1 and survival of antiviral CD8(+) T cells during chronic infection, Immunity 41 (5) (2014) 802–814.
- [80] R.V. Parry, J.M. Chemnitz, K.A. Frauwirth, A.R. Lanfranco, I. Braunstein, S.V. Kobayashi, et al., CTLA-4 and PD-1 receptors inhibit T-cell activation by distinct mechanisms, Mol. Cell Biol. 25 (21) (2005) 9543–9553.
- [81] N. Patsoukis, J. Brown, V. Petkova, F. Liu, L. Li, V.A. Boussiotis, Selective effects of PD-1 on Akt and Ras pathways regulate molecular components of the cell cycle and inhibit T cell proliferation, Sci Signal. 5 (230) (2012) ra46.
- [82] M. Quigley, F. Pereyra, B. Nilsson, F. Porichis, C. Fonseca, Q. Eichbaum, et al., Transcriptional analysis of HIV-specific CD8 + T cells shows that PD-1 inhibits T cell function by upregulating BATF, Nat. Med. 16 (10) (2010) 1147–1151.
- [83] B.T. Fife, K.E. Pauken, T.N. Eagar, T. Obu, J. Wu, Q. Tang, et al., Interactions between PD-1 and PD-L1 promote tolerance by blocking the TCR-induced stop signal, Nat. Immunol. 10 (11) (2009) 1185–1192.
- [84] T. Honda, J.G. Egen, T. Lammermann, W. Kastenmuller, P. Torabi-Parizi, R.N. Germain, Tuning of antigen sensitivity by T cell receptor-dependent negative feedback controls T cell effector function in inflamed tissues, Immunity 40 (2) (2014) 235–247.
- [85] B.H. Zinselmeyer, S. Heydari, C. Sacristan, D. Nayak, M. Cammer, J. Herz, et al., PD-1 promotes immune exhaustion by inducing antiviral T cell motility paralysis, J. Exp. Med. 210 (4) (2013) 757–774.
- [86] J. Fourcade, P. Kudela, Z. Sun, H. Shen, S.R. Land, D. Lenzner, et al., PD-1 is a regulator of NY-ESO-1-specific CD8+ T cell expansion in melanoma patients, J. Immunol. 182 (9) (2009) 5240–5249.
- [87] H. Saito, H. Kuroda, T. Matsunaga, T. Osaki, M. Ikeguchi, Increased PD-1 expression on CD4+ and CD8+ T cells is involved in immune evasion in gastric cancer, J. Surg. Oncol. 107 (5) (2013) 517–522.
- [88] C.A. Chambers, M.S. Kuhns, J.G. Egen, J.P. Allison, CTLA-4-mediated inhibition in regulation of T cell responses: mechanisms and manipulation in tumor

immunotherapy, Annu. Rev. Immunol. 19 (2001) 565-594.

- [89] W.A. Teft, M.G. Kirchhof, J. Madrenas, A molecular perspective of CTLA-4 function, Annu. Rev. Immunol. 24 (2006) 65–97.
- [90] H. Schneider, J. Downey, A. Smith, B.H. Zinselmeyer, C. Rush, J.M. Brewer, et al., Reversal of the TCR stop signal by CTLA-4, Science 313 (5795) (2006) 1972–1975.
- [91] O.S. Qureshi, Y. Zheng, K. Nakamura, K. Attridge, C. Manzotti, E.M. Schmidt, et al., Trans-endocytosis of CD80 and CD86: a molecular basis for the cell-extrinsic function of CTLA-4, Science 332 (6029) (2011) 600–603.
- [92] J. Duraiswamy, K.M. Kaluza, G.J. Freeman, G. Coukos, Dual blockade of PD-1 and CTLA-4 combined with tumor vaccine effectively restores T-cell rejection function in tumors, Cancer Res. 73 (12) (2013) 3591–3603.
- [93] T. Fujita, B.J. Burwitz, G.M. Chew, J.S. Reed, R. Pathak, E. Seger, et al., Expansion of dysfunctional Tim-3-expressing effector memory CD8 + T cells during simian immunodeficiency virus infection in rhesus macaques, J. Immunol. 193 (11) (2014) 5576–5583.
- [94] R.B. Jones, L.C. Ndhlovu, J.D. Barbour, P.M. Sheth, A.R. Jha, B.R. Long, et al., Tim-3 expression defines a novel population of dysfunctional T cells with highly elevated frequencies in progressive HIV-1 infection, J. Exp. Med. 205 (12) (2008) 2763–2779.
- [95] R.H. McMahan, L. Golden-Mason, M.I. Nishimura, B.J. McMahon, M. Kemper, T.M. Allen, et al., Tim-3 expression on PD-1 + HCV-specific human CTLs is associated with viral persistence, and its blockade restores hepatocyte-directed in vitro cytotoxicity, J. Clin. Invest. 120 (12) (2010) 4546–4557.
- [96] K. Sakuishi, L. Apetoh, J.M. Sullivan, B.R. Blazar, V.K. Kuchroo, A.C. Anderson, Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore antitumor immunity, J. Exp. Med. 207 (10) (2010) 2187–2194.
- [97] S. Hannier, M. Tournier, G. Bismuth, F. Triebel, CD3/TCR complex-associated lymphocyte activation gene-3 molecules inhibit CD3/TCR signaling, J. Immunol. 161 (8) (1998) 4058–4065.
- [98] T. Okazaki, Okazaki I-m, J. Wang, D. Sugiura, F. Nakaki, T. Yoshida, et al., PD-1 and LAG-3 inhibitory co-receptors act synergistically to prevent autoimmunity in mice, J. Exp. Med. 208 (2) (2011) 395–407.
- [99] C.J. Workman, L.S. Cauley, I.J. Kim, M.A. Blackman, D.L. Woodland, D.A. Vignali, Lymphocyte activation gene-3 (CD223) regulates the size of the expanding T cell population following antigen activation in vivo, J. Immunol. 172 (9) (2004) 5450–5455.
- [100] C.T. Huang, C.J. Workman, D. Flies, X. Pan, A.L. Marson, G. Zhou, et al., Role of LAG-3 in regulatory T cells, Immunity 21 (4) (2004) 503–513.
- [101] K. Richter, P. Agnellini, A. Oxenius, On the role of the inhibitory receptor LAG-3 in acute and chronic LCMV infection, Int. Immunol. 22 (1) (2010) 13–23.
- [102] S.R. Woo, M.E. Turnis, M.V. Goldberg, J. Bankoti, M. Selby, C.J. Nirschl, et al., Immune inhibitory molecules LAG-3 and PD-1 synergistically regulate T-cell function to promote tumoral immune escape, Cancer Res. 72 (4) (2012) 917–927.
- [103] L. Derré, J.-P. Rivals, C. Jandus, S. Pastor, D. Rimoldi, P. Romero, et al., BTLA mediates inhibition of human tumor-specific CD8(+) T cells that can be partially reversed by vaccination, J. Clin. Invest. 120 (1) (2010) 157–167.
- [104] X. Yu, K. Harden, L.C. Gonzalez, M. Francesco, E. Chiang, B. Irving, et al., The surface protein TIGIT suppresses T cell activation by promoting the generation of mature immunoregulatory dendritic cells. Nat. Immunol. 10 (1) (2009) 48–57.
- [105] J.M. Chauvin, O. Pagliano, J. Fourcade, Z. Sun, H. Wang, C. Sander, et al., TIGIT and PD-1 impair tumor antigen-specific CD8(+) T cells in melanoma patients, J. Clin. Invest. 125 (5) (2015) 2046–2058.
- [106] R.J. Johnston, L. Comps-Agrar, J. Hackney, X. Yu, M. Huseni, Y. Yang, et al., The immunoreceptor TIGIT regulates antitumor and antiviral CD8(+) T cell effector function, Cancer Cell 26 (6) (2014) 923–937.
- [107] H. Shin, S.D. Blackburn, A.M. Intlekofer, C. Kao, J.M. Angelosanto, S.L. Reiner, et al., A role for the transcriptional repressor Blimp-1 in CD8(+) T cell exhaustion during chronic viral infection, Immunity 31 (2) (2009) 309–320.
- [108] P. Agnellini, P. Wolint, M. Rehr, J. Cahenzli, U. Karrer, A. Oxenius, Impaired NFAT nuclear translocation results in split exhaustion of virus-specific CD8(+) T cell functions during chronic viral infection, Proc. Natl. Acad. Sci. U.S.A. 104 (11) (2007) 4565–4570.
- [109] R. Yamamoto, M. Nishikori, T. Kitawaki, T. Sakai, M. Hishizawa, M. Tashima, et al., PD-1-PD-1 ligand interaction contributes to immunosuppressive microenvironment of Hodgkin lymphoma, Blood 111 (6) (2008) 3220–3224.
- [110] S. Terawaki, S. Chikuma, S. Shibayama, T. Hayashi, T. Yoshida, T. Okazaki, et al., IFN-alpha directly promotes programmed cell death-1 transcription and limits the duration of T cell-mediated immunity, J. immunol. 186 (5) (2011) 2772–2779.
- [111] G. Xiao, A. Deng, H. Liu, G. Ge, X. Liu, Activator protein 1 suppresses antitumor Tcell function via the induction of programmed death 1, Proc. Natl. Acad. Sci. U.S.A. 109 (38) (2012) 15419–15424.
- [112] S.L. Topalian, F.S. Hodi, J.R. Brahmer, S.N. Gettinger, D.C. Smith, D.F. McDermott, et al., Safety, activity, and immune correlates of anti-PD-1 antibody in cancer, N. Engl. J. Med. 366 (26) (2012) 2443–2454.
- [113] K.C. Ohaegbulam, A. Assal, E. Lazar-Molnar, Y. Yao, X. Zang, Human cancer immunotherapy with antibodies to the PD-1 and PD-L1 pathway, Trends Mol. Med. 21 (1) (2015) 24–33.
- [114] H. Nishikawa, S. Sakaguchi, Regulatory T cells in tumor immunity, Int. J. Cancer 127 (4) (2010) 759–767.
- [115] D. Mougiakakos, A. Choudhury, A. Lladser, R. Kiessling, C.C. Johansson, Regulatory T cells in cancer, Adv. Cancer Res. 107 (2010) 57–117.
- [116] G. Borsellino, M. Kleinewietfeld, D. Di Mitri, A. Sternjak, A. Diamantini, R. Giometto, et al., Expression of ectonucleotidase CD39 by Foxp3 + Treg cells: hydrolysis of extracellular ATP and immune suppression, Blood 110 (4) (2007) 1225–1232.
- [117] J.J. Kobie, P.R. Shah, L. Yang, J.A. Rebhahn, D.J. Fowell, T.R. Mosmann, T

regulatory and primed uncommitted CD4 T cells express CD73, which suppresses effector CD4 T cells by converting 5'-adenosine monophosphate to adenosine, J. Immunol. 177 (10) (2006) 6780–6786.

- [118] J.D. Fontenot, J.P. Rasmussen, M.A. Gavin, A.Y. Rudensky, A function for interleukin 2 in Foxp3-expressing regulatory T cells, Nat. Immunol. 6 (11) (2005) 1142–1151.
- [119] O. Annacker, C. Asseman, S. Read, F. Powrie, Interleukin-10 in the regulation of T cell-induced colitis, J. Autoimmun. 20 (4) (2003) 277–279.
- [120] D.A. Thomas, J. Massague, TGF-beta directly targets cytotoxic T cell functions during tumor evasion of immune surveillance, Cancer Cell 8 (5) (2005) 369–380.
- [121] S.A. Hahn, A. Neuhoff, J. Landsberg, J. Schupp, D. Eberts, P. Leukel, et al., A key role of GARP in the immune suppressive tumor microenvironment, Oncotarget 7 (28) (2016) 42996–43009.
- [122] M.D. Sharma, B. Baban, P. Chandler, D.Y. Hou, N. Singh, H. Yagita, et al., Plasmacytoid dendritic cells from mouse tumor-draining lymph nodes directly activate mature Tregs via indoleamine 2,3-dioxygenase, J. Clin. Invest. 117 (9) (2007) 2570–2582.
- [123] A.A. Hurwitz, S.K. Watkins, Immune suppression in the tumor microenvironment: a role for dendritic cell-mediated tolerization of T cells, Cancer Immunol. Immunother. CII. 61 (2) (2012) 289–293.
- [124] P. Allavena, A. Mantovani, Immunology in the clinic review series; focus on cancer: tumour-associated macrophages: undisputed stars of the inflammatory tumour microenvironment, Clin. Exp. Immunol. 167 (2) (2012) 195–205.
- [125] C. Murdoch, S. Tazzyman, S. Webster, C.E. Lewis, Expression of Tie-2 by human monocytes and their responses to angiopoietin-2, J. immunol. 178 (11) (2007) 7405–7411.
- [126] A. Mantovani, W.J. Ming, C. Balotta, B. Abdeljalil, B. Bottazzi, Origin and regulation of tumor-associated macrophages: the role of tumor-derived chemotactic factor, Biochim. Biophys. Acta 865 (1) (1986) 59–67.
- [127] A. Mantovani, S. Sozzani, M. Locati, P. Allavena, A. Sica, Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes, Trends Immunol. 23 (11) (2002) 549–555.
- [128] V. Bronte, Myeloid-derived suppressor cells in inflammation: uncovering cell subsets with enhanced immunosuppressive functions, Eur. J. Immunol. 39 (10) (2009) 2670–2672.
- [129] S. Ostrand-Rosenberg, P. Sinha, Myeloid-derived suppressor cells: linking inflammation and cancer, J. Immunol. 182 (8) (2009) 4499–4506.
- [130] Y. Liu B. Zeng Z. Zhang Y. Zhang R. Yang B7-H1 on myeloid-derived suppressor cells in immune suppression by a mouse model of ovarian cancer. Clin. immunol. 2008 129 3 471 481.
- [131] R. Yang, Z. Cai, Y. Zhang, Yutzy WHt, Roby KF, Roden RB. CD80 in immune suppression by mouse ovarian carcinoma-associated Gr-1+CD11b+ myeloid cells, Cancer Res. 66 (13) (2006) 6807–6815.
- [132] Y.J. Kim, S.J. Park, H.E. Broxmeyer, Phagocytosis, a potential mechanism for myeloid-derived suppressor cell regulation of CD8 + T cell function mediated through programmed cell death-1 and programmed cell death-1 ligand interaction, J. immunol. 187 (5) (2011) 2291–2301.
- [133] L. Strauss, C. Bergmann, W. Gooding, J.T. Johnson, T.L. Whiteside, The frequency and suppressor function of CD4+CD25highFoxp3+ T cells in the circulation of patients with squamous cell carcinoma of the head and neck, Clin. Cancer Res. 13 (21) (2007) 6301–6311.
- [134] G. Landskron, M. De la Fuente, P. Thuwajit, C. Thuwajit, M.A. Hermoso, Chronic inflammation and cytokines in the tumor microenvironment, J. Immunol. Res. 2014 (2014) 149185.
- [135] M.G. Roncarolo, R. Bacchetta, C. Bordignon, S. Narula, M.K. Levings, Type 1 T regulatory cells, Immunol. Rev. 182 (2001) 68–79.
- [136] J. Massague, TGFbeta in Cancer, Cell 134 (2) (2008) 215-230.
- [137] L.H. Katz, Y. Li, J.S. Chen, N.M. Munoz, A. Majumdar, J. Chen, et al., Targeting TGF-beta signaling in cancer, Expert Opin. Ther. Targets. 17 (7) (2013) 743–760.
- [138] J.A. Trapani, The dual adverse effects of TGF-beta secretion on tumor progression, Cancer Cell 8 (5) (2005) 349–350.
- [139] R. Lin, L. Chen, G. Chen, C. Hu, S. Jiang, J. Sevilla, et al., Targeting miR-23a in CD8+ cytotoxic T lymphocytes prevents tumor-dependent immunosuppression, J. Clin. Invest. 124 (12) (2014) 5352–5367.
- [140] W. Chen, W. Jin, N. Hardegen, K.J. Lei, L. Li, N. Marinos, et al., Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGFbeta induction of transcription factor Foxp3, J. Exp. Med. 198 (12) (2003) 1875–1886.
- [141] N.N. Pavlova, C.B. Thompson, The emerging Hallmarks of cancer metabolism, Cell Metab. 23 (1) (2016) 27–47.
- [142] C. Herbel, N. Patsoukis, K. Bardhan, P. Seth, J.D. Weaver, V.A. Boussiotis, Clinical significance of T cell metabolic reprogramming in cancer, Clin. Trans. Med. 5 (1) (2016) 29.
- [143] K.A. Frauwirth, J.L. Riley, M.H. Harris, R.V. Parry, J.C. Rathmell, D.R. Plas, et al., The CD28 signaling pathway regulates glucose metabolism, Immunity 16 (6) (2002) 769–777.
- [144] R.L. Elstrom, D.E. Bauer, M. Buzzai, R. Karnauskas, M.H. Harris, D.R. Plas, et al., Akt stimulates aerobic glycolysis in cancer cells, Cancer Res. 64 (11) (2004) 3892–3899.
- [145] C.V. Dang, MYC, metabolism, cell growth, and tumorigenesis, Cold Spring Harb. Perspect. Med. 3 (8) (2013).
- [146] V.G. Antico Arciuch, M.A. Russo, K.S. Kang, A. Di Cristofano, Inhibition of AMPK and Krebs cycle gene expression drives metabolic remodeling of Pten-deficient preneoplastic thyroid cells, Cancer Res. 73 (17) (2013) 5459–5472.
- [147] R. Wang, D.R. Green, Metabolic checkpoints in activated T cells, Nat. Immunol. 13 (10) (2012) 907–915.

- [148] P. Vaupel, A. Mayer, Hypoxia in cancer: significance and impact on clinical outcome, Cancer Metastasis Rev. 26 (2) (2007) 225–239.
- [149] K.L. Eales, K.E. Hollinshead, D.A. Tennant, Hypoxia and metabolic adaptation of cancer cells, Oncogenesis 5 (2016) e190.
- [150] J.W. Kim, I. Tchernyshyov, G.L. Semenza, C.V. Dang, HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia, Cell Metab. 3 (3) (2006) 177–185.
- [151] J.D. Firth, B.L. Ebert, C.W. Pugh, P.J. Ratcliffe, Oxygen-regulated control elements in the phosphoglycerate kinase 1 and lactate dehydrogenase A genes: similarities with the erythropoietin 3' enhancer, Proc. Natl. Acad. Sci. U.S.A. 91 (14) (1994) 6496–6500.
- [152] A.K. Neumann, J. Yang, M.P. Biju, S.K. Joseph, R.S. Johnson, V.H. Haase, et al., Hypoxia inducible factor 1 alpha regulates T cell receptor signal transduction, Proc. Natl. Acad. Sci. U.S.A. 102 (47) (2005) 17071–17076.
- [153] M.V. Sitkovsky, D. Lukashev, S. Apasov, H. Kojima, M. Koshiba, C. Caldwell, et al., Physiological control of immune response and inflammatory tissue damage by hypoxia-inducible factors and adenosine A2A receptors, Annu. Rev. Immunol. 22 (2004) 657–682.
- [154] K.R. Atkuri, L.A. Herzenberg, L.A. Herzenberg, Culturing at atmospheric oxygen levels impacts lymphocyte function, Proc. Natl. Acad. Sci. U.S.A. 102 (10) (2005) 3756–3759.
- [155] A. Larbi, H. Zelba, D. Goldeck, G. Pawelec, Induction of HIF-1alpha and the glycolytic pathway alters apoptotic and differentiation profiles of activated human T cells, J. Leukoc. Biol. 87 (2) (2010) 265–273.
- [156] A.L. Zuckerberg, L.I. Goldberg, H.M. Lederman, Effects of hypoxia on interleukin-2 mRNA expression by T lymphocytes, Crit. Care Med. 22 (2) (1994) 197–203.
- [157] H. Kim, G. Peng, J.M. Hicks, H.L. Weiss, E.G. Van Meir, M.K. Brenner, et al., Engineering human tumor-specific cytotoxic T cells to function in a hypoxic environment, Molecular Ther. 16 (3) (2008) 599–606.
- [158] E.L. Bell, T.A. Klimova, J. Eisenbart, C.T. Moraes, M.P. Murphy, G.R. Budinger, et al., The Qo site of the mitochondrial complex III is required for the transduction of hypoxic signaling via reactive oxygen species production, J. Cell Biol. 177 (6) (2007) 1029–1036.
- [159] P. Kesarwani, A.K. Murali, A.A. Al-Khami, S. Mehrotra, Redox regulation of T-cell function: from molecular mechanisms to significance in human health and disease, Antioxid. Redox Signal. 18 (12) (2013) 1497–1534.
- [160] J. Guo, W. Lu, L.A. Shimoda, G.L. Semenza, S.N. Georas, Enhanced interferongamma gene expression in T Cells and reduced ovalbumin-dependent lung eosinophilia in hypoxia-inducible factor-1-alpha-deficient mice, Int. Arch. Allergy Immunol. 149 (2) (2009) 98–102.
- [161] D. Lukashev, B. Klebanov, H. Kojima, A. Grinberg, A. Ohta, L. Berenfeld, et al., Cutting edge: hypoxia-inducible factor 1alpha and its activation-inducible short isoform 1.1 negatively regulate functions of CD4+ and CD8+ T lymphocytes, J. Immunol. 177 (8) (2006) 4962–4965.
- [162] A.L. Doedens, A.T. Phan, M.H. Stradner, J.K. Fujimoto, J.V. Nguyen, E. Yang, et al., Hypoxia-inducible factors enhance the effector responses of CD8(+) T cells to persistent antigen, Nat. Immunol. 14 (11) (2013) 1173–1182.
- [163] I.B. Barsoum, C.A. Smallwood, D.R. Siemens, C.H. Graham, A mechanism of hypoxia-mediated escape from adaptive immunity in cancer cells, Cancer Res. 74 (3) (2014) 665–674.
- [164] H.Y. Fang, R. Hughes, C. Murdoch, S.B. Coffelt, S.K. Biswas, A.L. Harris, et al., Hypoxia-inducible factors 1 and 2 are important transcriptional effectors in primary macrophages experiencing hypoxia, Blood 114 (4) (2009) 844–859.
- [165] V. Nizet, R.S. Johnson, Interdependence of hypoxic and innate immune responses, Nat. Rev. Immunol. 9 (9) (2009) 609–617.
- [166] C.M. Cham, T.F. Gajewski, Glucose availability regulates IFN-gamma production and p70S6 kinase activation in CD8 + effector T cells, J. Immunol. 174 (8) (2005) 4670–4677.
- [167] C.M. Cham, G. Driessens, J.P. O'Keefe, T.F. Gajewski, Glucose deprivation inhibits multiple key gene expression events and effector functions in CD8 + T cells, Eur. J. Immunol. 38 (9) (2008) 2438–2450.
- [168] S.R. Jacobs, C.E. Herman, N.J. Maciver, J.A. Wofford, H.L. Wieman, J.J. Hammen, et al., Glucose uptake is limiting in T cell activation and requires CD28-mediated Akt-dependent and independent pathways, J. Immunol. 180 (7) (2008) 4476–4486.
- [169] A.N. Macintyre, V.A. Gerriets, A.G. Nichols, R.D. Michalek, M.C. Rudolph, D. Deoliveira, et al., The glucose transporter Glut1 is selectively essential for CD4 T cell activation and effector function, Cell Metab. 20 (1) (2014) 61–72.
- [170] P.C. Ho, J.D. Bihuniak, A.N. Macintyre, M. Staron, X. Liu, R. Amezquita, et al., Phosphoenolpyruvate is a metabolic checkpoint of anti-tumor T cell Responses, Cell 162 (6) (2015) 1217–1228.
- [171] O. Warburg, On the origin of cancer cells, Science 123 (3191) (1956) 309-314.
- [172] J. Blagih, F. Coulombe, E.E. Vincent, F. Dupuy, G. Galicia-Vazquez, E. Yurchenko, et al., The energy sensor AMPK regulates T cell metabolic adaptation and effector responses in vivo, Immunity 42 (1) (2015) 41–54.
- [173] L.A. O'Neill, D.G. Hardie, Metabolism of inflammation limited by AMPK and pseudo-starvation, Nature 493 (7432) (2013) 346–355.
- [174] R.D. Michalek, V.A. Gerriets, S.R. Jacobs, A.N. Macintyre, N.J. MacIver, E.F. Mason, et al., Cutting edge: distinct glycolytic and lipid oxidative metabolic programs are essential for effector and regulatory CD4+ T cell subsets, J. Immunol. 186 (6) (2011) 3299–3303.
- [175] H.J. Son, J. Lee, S.Y. Lee, E.K. Kim, M.J. Park, K.W. Kim, et al., Metformin attenuates experimental autoimmune arthritis through reciprocal regulation of Th17/Treg balance and osteoclastogenesis, Mediators inflamm. 2014 (2014) 973986.
- [176] N. Patsoukis, K. Bardhan, P. Chatterjee, D. Sari, B. Liu, L.N. Bell, et al., PD-1 alters

T-cell metabolic reprogramming by inhibiting glycolysis and promoting lipolysis and fatty acid oxidation, Nat. Commun. 6 (2015) 6692.

- [177] K. Fischer, P. Hoffmann, S. Voelkl, N. Meidenbauer, J. Ammer, M. Edinger, et al., Inhibitory effect of tumor cell-derived lactic acid on human T cells, Blood 109 (9) (2007) 3812–3819.
- [178] O.R. Colegio, N.Q. Chu, A.L. Szabo, T. Chu, A.M. Rhebergen, V. Jairam, et al., Functional polarization of tumour-associated macrophages by tumour-derived lactic acid, Nature 513 (7519) (2014) 559–563.
- [179] T. Ohashi, T. Akazawa, M. Aoki, B. Kuze, K. Mizuta, Y. Ito, et al., Dichloroacetate improves immune dysfunction caused by tumor-secreted lactic acid and increases antitumor immunoreactivity, Int. J. Cancer 133 (5) (2013) 1107–1118.
- [180] D.K. Nomura, J.Z. Long, S. Niessen, H.S. Hoover, S.W. Ng, B.F. Cravatt, Monoacylglycerol lipase regulates a fatty acid network that promotes cancer pathogenesis, Cell 140 (1) (2010) 49–61.
- [181] K.M. Nieman, H.A. Kenny, C.V. Penicka, A. Ladanyi, R. Buell-Gutbrod, M.R. Zillhardt, et al., Adipocytes promote ovarian cancer metastasis and provide energy for rapid tumor growth, Nat. Med. 17 (11) (2011) 1498–1503.
- [182] F. Hossain, A.A. Al-Khami, D. Wyczechowska, C. Hernandez, L. Zheng, K. Reiss, et al., Inhibition of fatty acid oxidation modulates immunosuppressive functions of myeloid-derived suppressor cells and enhances cancer therapies, Cancer Immunol. Res. 3 (11) (2015) 1236–1247.
- [183] D.L. Herber, W. Cao, Y. Nefedova, S.V. Novitskiy, S. Nagaraj, V.A. Tyurin, et al., Lipid accumulation and dendritic cell dysfunction in cancer, Nat. Med. 16 (8) (2010) 880–886.
- [184] F. Gao, C. Liu, J. Guo, W. Sun, L. Xian, D. Bai, et al., Radiation-driven lipid accumulation and dendritic cell dysfunction in cancer, Sci. Rep. 5 (2015) 9613.
- [185] J.R. Cubillos-Ruiz, P.C. Silberman, M.R. Rutkowski, S. Chopra, A. Perales-Puchalt, M. Song, et al., ER Stress sensor XBP1 controls anti-tumor immunity by disrupting dendritic cell homeostasis, Cell 161 (7) (2015) 1527–1538.
- [186] D. Vats, L. Mukundan, J.I. Odegaard, L. Zhang, K.L. Smith, C.R. Morel, et al., Oxidative metabolism and PGC-1beta attenuate macrophage-mediated inflammation, Cell Metab. 4 (1) (2006) 13–24.
- [187] M.B. Fessler, Regulation of adaptive immunity in health and disease by cholesterol metabolism, Curr. Allergy Asthma Rep. 15 (8) (2015) 48.
- [188] T.Y. Chang, C.C. Chang, N. Ohgami, Y. Yamauchi, Cholesterol sensing, trafficking, and esterification, Annu. Rev. Cell Dev. Biol. 22 (2006) 129–157.
- [189] W. Yang, Y. Bai, Y. Xiong, J. Zhang, S. Chen, X. Zheng, et al., Potentiating the antitumour response of CD8(+) T cells by modulating cholesterol metabolism, Nature 531 (7596) (2016) 651–655.
- [190] C.A. Opitz, U.M. Litzenburger, F. Sahm, M. Ott, I. Tritschler, S. Trump, et al., An endogenous tumour-promoting ligand of the human aryl hydrocarbon receptor, Nature 478 (7368) (2011) 197–203.
- [191] P.C. Rodriguez, M.S. Ernstoff, C. Hernandez, M. Atkins, J. Zabaleta, R. Sierra, et al., Arginase I-producing myeloid-derived suppressor cells in renal cell carcinoma are a subpopulation of activated granulocytes, Cancer Res. 69 (4) (2009) 1553–1560.
- [192] M. Nakaya, Y. Xiao, X. Zhou, J.H. Chang, M. Chang, X. Cheng, et al., Inflammatory T cell responses rely on amino acid transporter ASCT2 facilitation of glutamine uptake and mTORC1 kinase activation, Immunity 40 (5) (2014) 692–705.
- [193] L.H. Camacho, S. Antonia, J. Sosman, J.M. Kirkwood, T.F. Gajewski, B. Redman, et al., Phase I/II trial of tremelimumab in patients with metastatic melanoma, J. Clin. Oncol. 27 (7) (2009) 1075–1081.
- [194] P. Momtaz, M.A. Postow, Immunologic checkpoints in cancer therapy: focus on the programmed death-1 (PD-1) receptor pathway, Pharmgenomics Pers Med. 7 (2014) 357–365.
- [195] G. Graziani, L. Tentori, P. Navarra, Ipilimumab: a novel immunostimulatory monoclonal antibody for the treatment of cancer, Pharmacol. Res. 65 (1) (2012) 9–22.
- [196] S.S. Ramalingam, J. Mazières, D. Planchard, T.E. Stinchcombe, G.K. Dy,
- S.J. Antonia, et al., Phase II study of nivolumab (anti-PD-1, BMS-936558, ONO-4538) in patients with advanced, refractory squamous non-small cell lung cancer, Int. J. Rad. Oncol. Biol. Phys. 90 (5) (2014) 1266–1267.
- [197] O. Hamid, C. Robert, A. Daud, F.S. Hodi, W.-J. Hwu, R. Kefford, et al., Safety and tumor responses with lambrolizumab (Anti–PD-1) in melanoma, N Eng. J. Med. 369 (2) (2013) 134–144.
- [198] R. Berger, R. Rotem-Yehudar, G. Slama, S. Landes, A. Kneller, M. Leiba, et al., Phase I safety and pharmacokinetic study of CT-011, a humanized antibody interacting with PD-1, in patients with advanced hematologic malignancies, Clin. Cancer Res. 14 (10) (2008) 3044–3051.
- [199] S.S. Tykodi, PD-1 as an emerging therapeutic target in renal cell carcinoma: current evidence, OncoTargets Ther. 7 (2014) 1349–1359.
- [200] T. Powles, J.P. Eder, G.D. Fine, F.S. Braiteh, Y. Loriot, C. Cruz, et al., MPDL3280A (anti-PD-L1) treatment leads to clinical activity in metastatic bladder cancer, Nature 515 (7528) (2014) 558–562.
- [201] J. Lu, L. Lee-Gabel, M.C. Nadeau, T.M. Ferencz, S.A. Soefje, Clinical evaluation of compounds targeting PD-1/PD-L1 pathway for cancer immunotherapy, J. Oncol. Pharm. Pract. 21 (6) (2015) 451–467.
- [202] M.A. Postow, J. Chesney, A.C. Pavlick, C. Robert, K. Grossmann, D. McDermott, et al., Nivolumab and ipilimumab versus ipilimumab in untreated melanoma, N Engl. J. Med. 372 (21) (2015) 2006–2017.
- [203] J. Larkin, V. Chiarion-Sileni, R. Gonzalez, J.J. Grob, C.L. Cowey, C.D. Lao, et al., Combined nivolumab and ipilimumab or monotherapy in untreated melanoma, N Engl. J. Med. 373 (1) (2015) 23–34.
- [204] M.V. Maus D.J. Powell Jr. Chimeric antigen receptor t-cells: new approaches to improve their efficacy and reduce toxicity Cancer J. 2015 21 6 475 479.
- [205] R.J. Brentjens, M.L. Davila, I. Riviere, J. Park, X. Wang, L.G. Cowell, et al., CD19-

targeted T cells rapidly induce molecular remissions in adults with chemotherapyrefractory acute lymphoblastic leukemia, Sci. Transl. Med. 5 (177) (2013) 177ra38.

- [206] S.A. Grupp, M. Kalos, D. Barrett, R. Aplenc, D.L. Porter, S.R. Rheingold, et al., Chimeric antigen receptor-modified T cells for acute lymphoid leukemia, N Eng. J. Med. 368 (16) (2013) 1509–1518.
- [207] S. Kakarla, S. Gottschalk, CAR T cells for solid tumors: armed and ready to go? Cancer J. 20 (2) (2014) 151–155.
- [208] J.R. Park, D.L. Digiusto, M. Slovak, C. Wright, A. Naranjo, J. Wagner, et al., Adoptive transfer of chimeric antigen receptor re-directed cytolytic T lymphocyte clones in patients with neuroblastoma, Mol. Ther. 15 (4) (2007) 825–833.
- [209] C.H. Lamers, S. Sleijfer, A.G. Vulto, W.H. Kruit, M. Kliffen, R. Debets, et al., Treatment of metastatic renal cell carcinoma with autologous T-lymphocytes genetically retargeted against carbonic anhydrase IX: first clinical experience, J. Clin. Oncol. 24 (13) (2006) e20–e22.
- [210] P.A. Beavis, C.Y. Slaney, M.H. Kershaw, D. Gyorki, P.J. Neeson, P.K. Darcy, Reprogramming the tumor microenvironment to enhance adoptive cellular therapy, Semin. Immunol. 28 (1) (2016) 64–72.
- [211] L.B. John, M.H. Kershaw, P.K. Darcy, Blockade of PD-1 immunosuppression boosts CAR T-cell therapy, Oncoimmunology 2 (10) (2013) e26286.
- [212] L.B. John, C. Devaud, C.P. Duong, C.S. Yong, P.A. Beavis, N.M. Haynes, et al., Anti-PD-1 antibody therapy potently enhances the eradication of established tumors by gene-modified T cells, Clin. Cancer Res. 19 (20) (2013) 5636–5646.
- [213] R.J. Deberardinis, J.J. Lum, C.B. Thompson, Phosphatidylinositol 3-kinase-dependent modulation of carnitine palmitoyltransferase 1A expression regulates lipid metabolism during hematopoietic cell growth, J. Biol. Chem. 281 (49) (2006) 37372–37380.
- [214] A. Saha, K. Aoyama, P.A. Taylor, B.H. Koehn, R.G. Veenstra, A. Panoskaltsis-Mortari, et al., Host programmed death ligand 1 is dominant over programmed death ligand 2 expression in regulating graft-versus-host disease lethality, Blood

122 (17) (2013) 3062–3073.

- [215] Y. Liu, Y. Cao, W. Zhang, S. Bergmeier, Y. Qian, H. Akbar, et al., A small-molecule inhibitor of glucose transporter 1 downregulates glycolysis, induces cell-cycle arrest, and inhibits cancer cell growth in vitro and in vivo, Mol. Cancer Ther. 11 (8) (2012) 1672–1682.
- [216] K. Birsoy, T. Wang, R. Possemato, O.H. Yilmaz, C.E. Koch, W.W. Chen, et al., MCT1-mediated transport of a toxic molecule is an effective strategy for targeting glycolytic tumors, Nat. Genet. 45 (1) (2013) 104–108.
- [217] L. Gattinoni, C.A. Klebanoff, D.C. Palmer, C. Wrzesinski, K. Kerstann, Z. Yu, et al., Acquisition of full effector function in vitro paradoxically impairs the in vivo antitumor efficacy of adoptively transferred CD8 + T cells, J. Clin. Invest. 115 (6) (2005) 1616–1626.
- [218] M. Sukumar, J. Liu, Y. Ji, M. Subramanian, J.G. Crompton, Z. Yu, et al., Inhibiting glycolytic metabolism enhances CD8 + T cell memory and antitumor function, J. Clin. Invest. 123 (10) (2013) 4479–4488.
- [219] R. Zeng, R. Spolski, S.E. Finkelstein, S. Oh, P.E. Kovanen, C.S. Hinrichs, et al., Synergy of IL-21 and IL-15 in regulating CD8+ T cell expansion and function, J. Exp. Med. 201 (1) (2005) 139–148.
- [220] S. Liu, G. Lizee, Y. Lou, C. Liu, W.W. Overwijk, G. Wang, et al., IL-21 synergizes with IL-7 to augment expansion and anti-tumor function of cytotoxic T cells, Int. Immunol. 19 (10) (2007) 1213–1221.
- [221] N.M. Fewkes, C.L. Mackall, Novel gamma-chain cytokines as candidate immune modulators in immune therapies for cancer, Cancer J. 16 (4) (2010) 392–398.
- [222] C.A. Klebanoff, L. Gattinoni, P. Torabi-Parizi, K. Kerstann, A.R. Cardones, S.E. Finkelstein, et al., Central memory self/tumor-reactive CD8+ T cells confer superior antitumor immunity compared with effector memory T cells, Proc. Natl. Acad. Sci. U.S.A. 102 (27) (2005) 9571–9576.
- [223] C.A. Klebanoff, L. Gattinoni, D.C. Palmer, P. Muranski, Y. Ji, C.S. Hinrichs, et al., Determinants of successful CD8+ T-cell adoptive immunotherapy for large established tumors in mice, Clin. Cancer Res. 17 (16) (2011) 5343–5352.