



Mini-review

Immunotherapy holds the key to cancer treatment and prevention in constitutional mismatch repair deficiency (CMMRD) syndrome



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ABSTRACT

Monoallelic germline mutations in one of the DNA mismatch repair (MMR) genes cause Lynch syndrome, with a high lifetime risks of colorectal and endometrial cancer at adult age. Less well known, is the constitutional mismatch repair deficiency (CMMRD) syndrome caused by biallelic germline mutations in MMR genes. This syndrome is characterized by the development of childhood cancer. Patients with CMMRD are at extremely high risk of developing multiple cancers including hematological, brain and intestinal tumors. Mutations in MMR genes impair DNA repair and therefore most tumors of patients with CMMRD are hypermutated. These mutations lead to changes in the translational reading frame, which consequently result in neoantigen formation. Neoantigens are recognized as foreign by the immune system and can induce specific immune responses. The growing evidence on the clinical efficacy of immunotherapies, such as immune checkpoint inhibitors, offers the prospect for treatment of patients with CMMRD. Combining neoantigen-based vaccination strategies and immune checkpoint inhibitors could be an effective way to conquer CMMRD-related tumors. Neoantigen-based vaccines might also be a preventive treatment option in healthy biallelic MMR mutation carriers. Future studies need to reveal the safety and efficacy of immunotherapies for patients with CMMRD.

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Introduction

Lynch syndrome (LS) is an autosomal dominant tumor syndrome predisposing to predominantly colorectal cancer (CRC) and endometrial carcinoma at an early age of onset, with a mean age of 45 years [1]. Monoallelic germline mutations in one of the DNA mismatch repair (MMR) genes cause malignancies in patients with LS when a second hit inactivates the wildtype allele (Fig. 1) [2–5]. The MMR genes involved in LS are *MLH1*, *MSH2*, *MSH6*, and *PMS2*.

Abbreviations: CMMRD, constitutional mismatch repair deficiency; COX, cyclooxygenase; CRC, colorectal cancer; CTLs, cytotoxic T lymphocytes; GBM, glioblastoma multiforme; LS, Lynch syndrome; MMR, mismatch repair; MSI, microsatellite instability; Tregs, regulatory T cells.

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Biallelic mutations in one of these MMR genes cause constitutional mismatch repair-deficiency (CMMRD). CMMRD is a rare recessively inherited syndrome mainly characterized by café-au-lait spots and a broad spectrum of childhood malignancies, primarily hematological, brain and intestinal tract tumors [6–9]. The prognosis for patients with CMMRD is much worse than for patients with LS due to the types of cancer and the high risk of multiple primary malignancies [10]. In children with CMMRD, no somatic mutations have to arise in the MMR genes to start tumorigenesis, since affected individuals inherit a germline MMR mutation from each parent. These biallelic MMR mutations, either homozygous or compound heterozygous, cause loss of genomic integrity by the inability of cells to repair DNA damage. This results in high numbers of mutations, mainly consisting of insertion and deletion mutations at repetitive DNA sequences known as microsatellites. Repeated DNA structures are prone to DNA polymerase slippage during DNA replication [11]. Due to these insertions and deletions, the length of the repeating sequences increases or decreases leading to

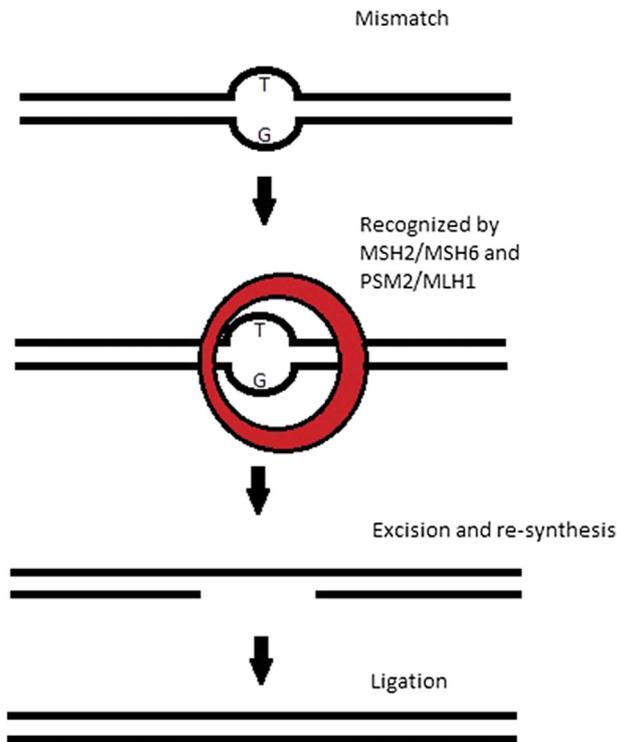


Fig. 1. Schematic figure of the mismatch repair pathway. A base mismatch is recognized by mismatch repair proteins (MSH2/MSH6 and PMS2/MLH1). The mismatch on the newly synthesized strand is excised and the correct nucleotides are synthesized. Finally, the DNA strands are ligated.

microsatellite instability (MSI). When microsatellites in gene-encoding regions are affected it can cause inactivation of the gene products through shifting of the translational reading frame leading to truncated or nonfunctional proteins (Fig. 2) [12–14]. Truncated proteins can be processed into peptides and presented on the surface of mutated cells. Eventually, so-called ultra-hypermutated tumor cells arise. All ultra-hypermutated tumors harbor mutations in polymerase genes *POLE* or *POLD1* and have an upper limit of

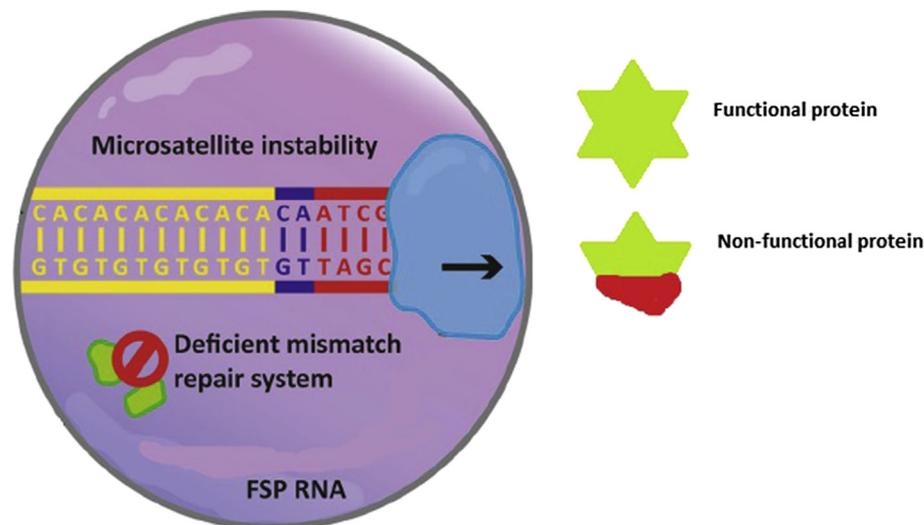


Fig. 2. Normally an insertion or deletion in a microsatellite sequence is repaired by the mismatch repair pathway to prevent that mutations become permanent and affect the protein. Mismatch repair deficient cells are unable to repair insertions or deletion, here indicated with an inserted CA sequence (purple), leading to a frameshift mutation. The coding sequence is altered due to shifting of the translational reading frame, which can result in a truncated or non-functional protein. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

exonic mutations which is tolerable [15,16]. Fortunately, endogenous ultra-hypermutated tumor cells can be recognized by the immune system as foreign as they can be identified by their private mutanome-derived epitopes called neoantigens. Most of the tumors that present in patients with CMMRD are ultra-hypermutated and thus express neoantigens, which can result in an immune response against these tumor-specific neoantigens [17]. Generally speaking, tumors that are recognized by the patients' own immune system have an improved prognosis [18,19]. Studies have shown that microsatellite instable CRCs of LS patients with a high density of tumor-infiltrating lymphocytes are associated with a better prognosis than microsatellite stable (MSS) CRCs [20,21]. Until now, very little is known about immune responses against neoantigens in CMMRD patients. In this review we discuss the potential of neoantigens to elicit immune responses in patients with CMMRD and probable immunotherapeutic treatment options.

Biallelic mutations in MMR genes and their risk of cancer

Individuals with CMMRD are at increased risk for developing malignant gliomas, hematologic malignancies, and gastrointestinal tract cancers at young age. It is a highly penetrating cancer predisposition syndrome, with most biallelic mutation carriers developing cancer in the first two decades of life [8,9]. Hematological malignancies usually arise in infancy or early childhood and are more frequent in patients with *MLH1* or *MSH2* mutations than in patients with mutations in *MSH6* or *PSM2* [6]. The latter group appears to have a higher prevalence of brain tumors which develop later during childhood. CRC in patients with CMMRD is most frequently found as a second or third primary malignancy and arises in adolescence or young adulthood. The prevalence of CRC is higher in patients with biallelic *PMS2* and *MSH6* [6,22–24].

Prophylactic cancer surveillance and preventive treatments

When a patient is diagnosed with CMMRD syndrome, family members can be screened for heterozygous and homozygous mutations by performing a mutation analysis of peripheral blood DNA. Especially, siblings need to be screened since the chance for a homozygous mutation is 25% resulting in CMMRD, and an additional

chance of 50% for heterozygous mutation resulting in LS. Patients with CMMRD may benefit from prophylactic cancer surveillance or preventive treatments through use of aspirin or ibuprofen. Surveillance may lead to earlier recognition of asymptomatic tumors which are better resectable and might be curable [25]. To improve the prognosis of CMMRD patients, surveillance should be offered for CRC and brain tumors [9]. The effectiveness of hematological screening is questionable since non-Hodgkin lymphomas and acute lymphoid leukemia are rapidly growing tumors and surveillance may not improve the outcome for patients with these malignancies [10]. In heterozygous MMR mutation carriers, daily intake of aspirin for approximately two years reduced the CRC incidence after a period of almost five years [26]. Another study reported that both the use of aspirin and ibuprofen might be effective in reducing the CRC risk for MMR gene mutation carriers [27]. Aspirin and ibuprofen, which belongs to the group of drugs called non-steroidal anti-inflammatory drugs, both inhibit cyclooxygenase (COX) enzymes resulting in decreased prostaglandin synthesis [28,29]. One isoform of COX, named COX-2 promotes inflammation and cell proliferation. In colorectal carcinogenesis the COX-2 enzyme is often overexpressed [28]. Prostaglandins are thought to regulate apoptosis, angiogenesis, and tumor-cell invasiveness [30]. Therefore, aspirin and non-steroidal anti-inflammatory drugs could reduce the risk of COX-2 expressing tumors. Whether there is overexpression of COX-2 in LS-associated CRC or CMMRD-related CRC is unknown. Regular aspirin use was also shown to lower the risk of CRC with low numbers of tumor-infiltrating lymphocytes. This was independent of MSI status [31]. It is postulated that aspirin, either related or unrelated to the inhibition of prostaglandins, may inhibit cancer development by overcoming suppression of T cell mediated antitumor immunity [32]. These results improve the understanding of the mechanisms through which aspirin exerts antineoplastic effects and provides support for the potential of exploiting immune mechanisms for cancer prevention [31]. Preventive treatments with aspirin and non-steroidal anti-inflammatory drugs might also reduce the incidence of cancer in patients with CMMRD.

Current treatment strategies

The treatment of childhood cancer is dependent of the specific location and the type of cancer. Since CMMRD is very rare, there is limited information on optimal therapeutic strategies. All described responses to chemotherapy are based on case report studies only. Chemotherapy treatment comes at the cost of toxicity. Therefore, careful selection of therapies is required for children with CMMRD. This should be based on the toxicity profile of specific agents as well as known tumor resistance to specific therapies [33].

MMR deficient cells are profoundly resistant to alkylating anti-neoplastic agents, such as temozolomide, cisplatin or busulfan. Temozolomide is frequently used in the standard treatment of glioblastomas multiforme (GBM) [10,34]. This is of clinical importance in patients with CMMRD, since it has become clear that these agents are less effective in MMR-deficient tumors and even may provide a growth advantage for the tumor cells [35]. Indeed, several studies have reported that treatment of GBM with temozolomide can promote further MMR deficiency due to loss of MSH2 or MSH6 expression, leading to temozolomide resistance [36,37]. *In vitro* studies also showed reduced sensitivity to the DNA damaging agents cisplatin and busulfan. Only one out of six patients with CMMRD-related GBM treated with temozolomide and radiotherapy showed a clinical response. The other patients' tumors were resistant to this therapy [10]. Another study showed that temozolomide treatment leads to accumulation of somatic mutations [38]. Moreover, temozolomide increases the risk of second primary

tumors in patients with CMMRD because of their inability to repair the accumulated somatic mutations [9,10,39,40]. On the contrary, recent data supports that temozolomide treatment in glioma cells can boost the adaptive immune response [41].

Whether temozolomide, cisplatin and busulfan are safe in patients with CMMRD is still controversial and requires further clinical studies [10]. Until now it remains unclear which chemotherapeutic regimen is most effective and the least dangerous.

The role of the immune system in cancer

The immune system is able to protect the body against cancer development. Cancer cells are immunogenic through the generation of tumor-specific antigens that are the consequence of somatic mutations and epigenetic alterations in the DNA. Immune cells, such as killer T lymphocytes and natural killer cells, are capable of tumor cell killing by the release of cytolytic granules, such as perforin and granzymes. Tumor cells with a less immunogenic phenotype are able to escape the immune attack and progress further, leading to resistance for immune detection. Tumors can acquire several immune escape mechanisms which include defective antigen presentation, lack of tumor antigen recognition, loss of sensitivity by cancer cells through secretion of immunosuppressive cytokines, induction of inhibitory checkpoint receptors, and infiltrating immunosuppressive immune cells [42–44]. Immunosuppressive cells, such as Foxp3-positive regulatory T cells (Tregs) and myeloid-derived suppressor cells can suppress the immune response against tumor-specific antigens by suppression of cytotoxic T lymphocytes (CTLs) [22]. Although, several studies have shown that high Treg infiltration in MSI CRC is correlated with a poor outcome [45,46]. Others did not confirm this observation. Recently, it has been shown that the loss of HLA class I antigen expression as a result of beta-2 microglobulin mutations in LS tumors, leads to less Treg infiltration [47]. Therefore, more research is needed to elucidate the role of Tregs and the exact location of these cells within the tumor [45–51]. Knowledge about the presence of Tregs in tumors and blood of patients with CMMRD is not yet available. Studies on the numbers and localization of CTLs, Tregs, myeloid-derived suppressor cells, and specific dendritic cell subsets could be very informative. The presence of these immune cells and the beta-2 microglobulin status may provide key information about the immunogenicity of CMMRD-associated tumors and about a proposed immunosuppressive tumor microenvironment.

Opportunities for cancer immunotherapy

The immune system encompasses inhibitory mechanisms to prevent excessive and therefore damaging immune responses. These inhibitory mechanisms are necessary for balanced immunity in normal homeostasis. However, in the presence of a growing malignancy the balance is disrupted and skewed towards excessive inhibition of immune reactivity due to tumor-induced immune suppression and enhanced immunologic tolerance [52]. Currently, immunotherapies are directed against inhibiting receptors, such as programmed death 1 (PD-1) protein, a T cell co-inhibitory receptor, and one of its ligands programmed death-ligand 1 (PD-L1). PD-L1 is typically expressed on the surface of tumor cells and inhibits activation of T cells through its receptor PD-1. In many tumors the expression of PD-L1 is aberrantly up-regulated and can provide inhibitory signals to activated T cells. PD-L1 expressing tumor cells can evade immune detection and prevent effective host antitumor immunity [42,53,54]. Antibodies that interfere with this pathway by blocking PD-1 or its ligand, have led to significant clinical responses in patients with many different types of cancer [54–58]. Several studies have shown that tumors with a high mutation load

and therefore a higher chance of expressing neoantigens were most likely to respond to immune checkpoint blockers [59,60]. A more diverse repertoire of neoantigens increases the chance of a tumor-specific T cell response [61]. MMR deficient tumors are more responsive to PD-1 blockades than MMR proficient tumors [54]. MMR deficient tumors harbor 10–100 times more somatic mutations and therefore more neoantigens are present which are able to elicit an immune response. Tumors of MMR deficient patients also showed a high number of tumor-infiltrating lymphocytes which is correlated with a better prognosis [54,62]. This data strongly suggest that treatment with immune checkpoint blockade can be very attractive for MMR deficient tumors independent of the underlying tumor type [61]. Likewise, immunotherapy with checkpoint inhibitors could hold the key to an effective cancer treatment and prevention strategy of CMMRD syndrome.

Children with GBM have a poor prognosis and sometimes have underlying germline mutations in *TP53* (Li-Fraumeni syndrome) or the MMR genes [9]. Primary management consists of surgical resection followed by radiation therapy and chemotherapy. Pediatric CMMRD-related GBM have similar outcomes to sporadic childhood GBMs [63]. The mean time from relapse to death is less than three months in CMMRD GBM. In a recent study, two siblings with CMMRD and relapsed GBM were treated with anti-PD-1-directed immune checkpoint inhibitor. Clinically significant responses and a profound radiologic response were observed. No severe treatment-related side effects were observed, except the presentation of seizures, hyponatremia or apparent disease flaring. However, these symptoms are possibly GBM-related and not associated with anti-PD-1 treatment. The results of this study may have implications for treatment of GBM in general and other ultra-hypermutated cancers [59]. Other recent data have demonstrated that checkpoint inhibitors are effective in the treatment of Hodgkin lymphomas and also appear efficient against some non-Hodgkin lymphomas [64,65]. Non-Hodgkin lymphomas are over-represented among hematological tumors in CMMRD patients, and treatment of Non-Hodgkin lymphoma patients is improving through the development of targeted therapies [9]. In a subset of Hodgkin lymphomas and in genetically related non-Hodgkin lymphomas, genetic amplification of the loci encoding the PD-1 ligands have been discovered. The overexpression of PD-1 on lymphoma cells may play a critical role in immune evasion by these cancers and makes it therefore an attractive target for checkpoint inhibitor therapy [66,67]. Further studies are required to investigate whether other tumors in patients with CMMRD will benefit from checkpoint inhibitor therapy.

Cellular immunotherapies, such as vaccines can also be used as a strategy to elicit a potent immune response against cancer antigens. However, the overall clinical efficacy of therapeutic cancer vaccines is disappointing until now [68,69]. This is probably due to the difficulty of identifying tumor-specific target antigens and overcoming the immunosuppressive tumor microenvironment. Tumor-specific antigens should be uniquely expressed by the tumor or overexpressed on the tumors as compared to normal cells. In the past years, multiple tumor vaccine strategies have been developed, such as tumor cell vaccines, tumor-associated antigen vaccines, and dendritic cell vaccines [68,70,71]. Anti-cancer vaccines, especially dendritic cell vaccination, might be effective when combined with immune-checkpoint inhibitors [72].

MSI in LS is associated with lymphocyte infiltration and a comparatively favorable prognosis. Tumors with MSI give rise to the generation of potentially immunogenic frameshift-derived neoantigens. For example, the *OGT* gene is commonly mutated in MSI colorectal tumors, and neoantigens derived from this protein stimulate CTLs that recognize this neoantigen [21]. Also TGF β RII derived neoantigens are highly immunogenic and are applicable as

target for tumor infiltrating CD4⁺ T cells in MSI tumors [62]. Vaccination with neoantigens is a promising approach for the treatment of LS patients, but also a promising approach for tumor prevention in healthy LS mutation carriers [22,73]. Neoantigen-loaded vaccination strategies are, in contrast to checkpoint inhibitors, antigen specific and should therefore elicit a specific effector- and memory cell response [70]. Several clinical studies are investigating the response of anti-cancer vaccines in LS patients. Currently, in a clinical phase I/II trial neoantigen-loaded dendritic cells vaccinations are studied in CRC patients with MSI or healthy germline MMR-gene mutation carriers. The primary objective is safety and feasibility, and secondary endpoints cover induction or enhancement of an immune response (NCT01885702). In another ongoing clinical phase I/IIa trial frameshift-derived neoantigen vaccination against *AIM2*, *HT001*, and *TAF1B* is studied (NCT01461148). Preliminary data suggest that neoantigen vaccination is safe and well tolerated [74]. Strong immune responses against neoantigens in all vaccinated patients were observed so far. LS mutation carriers already show neoantigen-specific immune responses [75]. Therefore, immune surveillance mechanisms with specific T cell responses may play an important role in preventing MSI tumor development in LS mutation carriers. Neoantigen vaccination strategies may be applied in future standard of care as an adjuvant therapy in MSI CRC patients or even as a preventive vaccine in MMR germline mutation carriers. Moreover, in patients with CMMRD adjuvant or preventive neoantigen-based vaccinations could be the basis for an effective treatment regimen for the induction of anti-cancer immune responses [74]. The number of CMMRD patients is limited, which makes it difficult to identify commonly mutated coding microsatellites. However, there might be overlap in the genes that are mutated early in tumorigenesis or neoantigens that are shared among tumors of a particular organ [21]. One must be cautious for selecting neoantigens for vaccination because every cell in CMMRD patients is MMR deficient, and when the neoantigen is present in healthy cells it might result in immune-related adverse events and might trigger autoimmune diseases [76,77]. Therefore, selection of cell growth- or apoptosis-related neoantigens might be the safest way to go.

Conclusion and future perspectives

CMMRD is a cancer predisposition syndrome with a high mortality rate. Currently, no curative treatment options are available for patients with CMMRD. CMMRD-derived tumors are ultra-hypermutated, leading to a high neoantigen load. This makes these tumors attractive for treatment with checkpoint inhibitors or neoantigen-based vaccination strategies. However, as described, several immunosuppressive hurdles need to be overcome. It has been hypothesized that the potential of immunotherapies might increase in case of combination therapy. (Neo)antigen-specific vaccination and checkpoint inhibition could act complementary, as a vaccine activates the immune system in a (neo)antigen-specific manner and concomitant or subsequent treatment with immune checkpoint inhibitors could boost the induced response to overcome immunosuppression. In addition, it has been shown in mice that antigen cross-presentation by DCs is even necessary for immune checkpoint inhibitor-induced immune responses to tumor-associated (neo)antigens [78]. A number of studies have analyzed the effect of anti-CTLA4 treatment after vaccination with DCs loaded with shared antigens. Pierret and colleagues suggested that anti-CTLA-4 treatment of advanced melanoma may be more effective after prior treatment with DC vaccination [79]. Another study suggested that anti-CTLA-4 treatment enhanced the T cell responses induced by prior DNA or protein vaccination [80]. In addition, in small clinical trials in melanoma patients and prostate

cancer patients, the combination of anti-CTLA-4 antibodies and DC-based immunotherapy [81,82] or other forms of antigen-specific immunotherapy [83,84] seemed more effective than treatment with either agent alone. Future research is needed to investigate the clinical effects and potential risks of these therapies in metastatic, adjuvant, and even preventive settings in patients with CMMRD.

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Conflict of interest

All authors declare that they have no competing financial interests.

References

- [1] F. Kastanos, S. Syngal, Inherited colorectal cancer syndromes, *Cancer J.* 17 (6) (2011) 405–415.
- [2] H. Kawakami, A. Zaanan, F.A. Sinicrope, Implications of mismatch repair-deficient status on management of early stage colorectal cancer, *J. Gastrointest. Oncol.* 6 (6) (2015) 676–684.
- [3] D.D. Buchanan, C. Rosty, M. Clendenning, A.B. Spurdle, A.K. Win, Clinical problems of colorectal cancer and endometrial cancer cases with unknown cause of tumor mismatch repair deficiency (suspected Lynch syndrome), *Appl. Clin. Genet.* 7 (2014) 183–193.
- [4] P. Peltomaki, Update on Lynch syndrome genomics, *Fam. Cancer* 15 (3) (2016) 385–393.
- [5] G.M. Li, Mechanisms and functions of DNA mismatch repair, *Cell Res.* 18 (1) (2008) 85–98.
- [6] K. Wimmer, C.P. Kratz, Constitutional mismatch repair-deficiency syndrome, *Haematologica* 95 (5) (2010) 699–701.
- [7] R.H. Sijmons, R.M. Hofstra, Review: clinical aspects of hereditary DNA Mismatch repair gene mutations, *DNA Repair (Amst)* 38 (2016) 155–162.
- [8] H.N. Baris, I. Barnes-Kedar, H. Toledano, M. Halpern, D. Hershkovitz, A. Lossos, et al., Constitutional mismatch repair deficiency in Israel: high proportion of founder mutations in MMR genes and consanguinity, *Pediatr. Blood Cancer* 63 (3) (2016) 418–427.
- [9] K. Wimmer, C.P. Kratz, H.F. Vasen, O. Caron, C. Colas, N. Entz-Werle, et al., Diagnostic criteria for constitutional mismatch repair deficiency syndrome: suggestions of the European consortium 'care for CMMRD' (C4CMMRD), *J. Med. Genet.* 51 (6) (2014) 355–365.
- [10] H.F. Vasen, Z. Ghorbanoghli, F. Bourdeau, O. Cabaret, O. Caron, A. Duval, et al., Guidelines for surveillance of individuals with constitutional mismatch repair-deficiency proposed by the European Consortium "Care for CMMRD-D" (C4CMMRD-D), *J. Med. Genet.* 51 (5) (2014) 283–293.
- [11] M. Kloor, S. Michel, M. von Knebel Doeberitz, Immune evasion of microsatellite unstable colorectal cancers, *Int. J. Cancer* 127 (5) (2010) 1001–1010.
- [12] M. Kloor, L. Staffa, A. Ahadova, M. von Knebel Doeberitz, Clinical significance of microsatellite instability in colorectal cancer, *Langenbecks Arch. Surg.* 399 (1) (2014) 23–31.
- [13] V. Lee, D.T. Le, Efficacy of PD-1 blockade in tumors with MMR deficiency, *Immunotherapy* 8 (1) (2016) 1–3.
- [14] L. Setaffy, C. Langner, Microsatellite instability in colorectal cancer: clinicopathological significance, *Pol. J. Pathol.* 66 (3) (2015) 203–218.
- [15] M. Schlesner, R. Eils, Hypermutation takes the driver's seat, *Genome Med.* 7 (1) (2015) 31.
- [16] S. Bobisse, P.G. Foukas, G. Coukos, A. Harari, Neoantigen-based cancer immunotherapy, *Ann. Transl. Med.* 4 (14) (2016) 262.
- [17] A. Shlien, B.B. Campbell, R. de Borja, L.B. Alexandrov, D. Merico, D. Wedge, et al., Combined hereditary and somatic mutations of replication error repair genes result in rapid onset of ultra-hypermutated cancers, *Nat. Genet.* 47 (3) (2015) 257–262.
- [18] A. Vasaturo, A. Halilovic, K.F. Bol, D.I. Verweij, W.A. Blokx, C.J. Punt, et al., T-cell landscape in a primary melanoma predicts the survival of patients with metastatic disease after their treatment with dendritic cell vaccines, *Cancer Res.* 76 (12) (2016) 3496–3506.
- [19] J. Galon, A. Costes, F. Sanchez-Cabo, A. Kirilovsky, B. Mlecnik, C. Lagorce-Pagès, et al., Type, density, and location of immune cells within human colorectal tumors predict clinical outcome, *Science* 313 (5795) (2006) 1960–1964.
- [20] P. Maby, D. Tougeron, M. Hamieh, B. Mlecnik, H. Kora, G. Bindea, et al., Correlation between density of CD8+ T-cell infiltrate in microsatellite unstable colorectal cancers and frameshift mutations: a rationale for personalized immunotherapy, *Cancer Res.* 75 (17) (2015) 3446–3455.
- [21] E. Ripberger, M. Linnebacher, Y. Schwitalle, J. Gebert, M. von Knebel Doeberitz, Identification of an HLA-A0201-restricted CTL epitope generated by a tumor-specific frameshift mutation in a coding microsatellite of the OGT gene, *J. Clin. Immunol.* 23 (5) (2003) 415–423.
- [22] H. Westdorp, F.L. Fennemann, R.D. Weren, T.M. Bisseling, M.J. Ligtenberg, C.G. Figdor, et al., Opportunities for immunotherapy in microsatellite instable colorectal cancer, *Cancer Immunol. Immunother.* 65 (10) (2016) 1249–1259.
- [23] M.S. Daniels, K.H. Lu, Clearer picture of PMS2-associated lynch syndrome is emerging, *J. Clin. Oncol.* 33 (4) (2015) 299–300.
- [24] L. Senter, M. Clendenning, K. Sotamaa, H. Hampel, J. Green, J.D. Potter, et al., The clinical phenotype of Lynch syndrome due to germ-line PMS2 mutations, *Gastroenterology* 135 (2) (2008) 419–428.
- [25] C.A. Durno, M. Aronson, U. Tabori, D. Malkin, S. Gallinger, H.S. Chan, Oncologic surveillance for subjects with biallelic mismatch repair gene mutations: 10 year follow-up of a kindred, *Pediatr. Blood Cancer* 59 (4) (2012) 652–656.
- [26] J. Burn, A.M. Gerdes, F. Macrae, J.P. Mecklin, G. Moeslein, S. Olschwang, et al., Long-term effect of aspirin on cancer risk in carriers of hereditary colorectal cancer: an analysis from the CAPP2 randomised controlled trial, *Lancet* 378 (9809) (2011) 2081–2087.
- [27] D. Ait Ouakrim, S.G. Dashti, R. Chau, D.D. Buchanan, M. Clendenning, C. Rosty, et al., Aspirin, ibuprofen, and the risk of colorectal cancer in lynch syndrome, *J. Natl. Cancer Inst.* 107 (9) (2015).
- [28] A.T. Chan, S. Ogino, C.S. Fuchs, Aspirin and the risk of colorectal cancer in relation to the expression of COX-2, *N. Engl. J. Med.* 356 (21) (2007) 2131–2142.
- [29] S.P. Fink, M. Yamauchi, R. Nishihara, S. Jung, A. Kuchiba, K. Wu, et al., Aspirin and the risk of colorectal cancer in relation to the expression of 15-hydroxyprostaglandin dehydrogenase (HPGD), *Sci. Transl. Med.* 6 (233) (2014) 233re2.
- [30] J.R. Brown, R.N. DuBois, COX-2: a molecular target for colorectal cancer prevention, *J. Clin. Oncol.* 23 (12) (2005) 2840–2855.
- [31] Y. Cao, R. Nishihara, Z.R. Qian, M. Song, K. Mima, K. Inamura, et al., Regular aspirin use associates with lower risk of colorectal cancers with low numbers of tumor-infiltrating lymphocytes, *Gastroenterology* 151 (5) (2016) 879–892 e4.
- [32] S. Zelenay, A.G. van der Veen, J.P. Böttcher, K.J. Snelgrove, N. Rogers, S.E. Acton, et al., Cyclooxygenase-dependent tumor growth through evasion of immunity, *Cell* 162 (6) (2015) 1257–1270.
- [33] R. Elhasid, R. Dvir, H. Rosenfeld Keidar, S. Ben Shachar, M. Bitan, I. Solar, et al., Management of acute myeloblastic leukemia in a child with biallelic mismatch repair deficiency, *J. Pediatr. Hematol. Oncol.* 37 (8) (2015) e490–e493.
- [34] R.H. Scott, S. Mansour, K. Pritchard-Jones, D. Kumar, F. MacSweeney, N. Rahman, Medulloblastoma, acute myelocytic leukemia and colonic carcinomas in a child with biallelic MSH6 mutations, *Nat. Clin. Pract. Oncol.* 4 (2) (2007) 130–134.
- [35] A. Fedier, D. Fink, Mutations in DNA mismatch repair genes: implications for DNA damage signaling and drug sensitivity (review), *Int. J. Oncol.* 24 (4) (2004) 1039–1047.
- [36] D.P. Cahill, K.K. Levine, R.A. Betensky, P.J. Codd, C.A. Romany, L.B. Reavie, et al., Loss of the mismatch repair protein MSH6 in human glioblastomas is associated with tumor progression during temozolomide treatment, *Clin. Cancer Res.* 13 (7) (2007) 2038–2045.
- [37] J.L. McFaline-Figueroa, C.J. Braun, M. Stanciu, Z.D. Nagel, P. Mazzucato, D. Sangaraju, et al., Minor changes in expression of the mismatch repair protein MSH2 exert a major impact on glioblastoma response to temozolomide, *Cancer Res.* 75 (15) (2015) 3127–3138.
- [38] C. Hunter, R. Smith, D.P. Cahill, P. Stephens, C. Stevens, J. Teague, et al., A hypermutation phenotype and somatic MSH6 mutations in recurrent human malignant gliomas after alkylator chemotherapy, *Cancer Res.* 66 (8) (2006) 3987–3991.
- [39] K. Wimmer, J. Etzler, Constitutional mismatch repair-deficiency syndrome: have we so far seen only the tip of an iceberg? *Hum. Genet.* 124 (2) (2008) 105–122.
- [40] L. Stojic, R. Brun, J. Jiricny, Mismatch repair and DNA damage signalling, *DNA Repair (Amst)* 3 (8–9) (2004) 1091–1101.
- [41] J. Proske, L. Walter, E. Bumès, M. Hutterer, A. Vollmann-Zwerenz, I.Y. Eyüpoğlu, et al., Adaptive immune response to and survival effect of temozolomide- and valproic acid-induced autophagy in glioblastoma, *Anticancer Res.* 36 (3) (2016) 899–905.
- [42] W. Ma, B.M. Gilligan, J. Yuan, T. Li, Current status and perspectives in translational biomarker research for PD-1/PD-L1 immune checkpoint blockade therapy, *J. Hematol. Oncol.* 9 (1) (2016) 47.
- [43] J.M. Zaretsky, A. Garcia-Diaz, D.S. Shin, H. Escuin-Ordinas, W. Hugo, S. Hu-Lieskovan, et al., Mutations associated with acquired resistance to PD-1 blockade in melanoma, *N. Engl. J. Med.* 375 (9) (2016) 819–829.
- [44] F.M. Marincola, E.M. Jaffee, D.J. Hicklin, S. Ferrone, Escape of human solid tumors from T-cell recognition: molecular mechanisms and functional significance, *Adv. Immunol.* 74 (2000) 181–273.
- [45] F.A. Sinicrope, R.L. Rego, S.M. Ansell, K.L. Knutson, N.R. Foster, D.J. Sargent, Intraepithelial effector (CD3+) / regulatory (FoxP3+) T-cell ratio predicts a clinical outcome of human colon carcinoma, *Gastroenterology* 137 (4) (2009) 1270–1279.

- [46] H. Suzuki, N. Chikazawa, T. Tasaka, J. Wada, A. Yamasaki, Y. Kitaura, et al., Intratumoral CD8(+) T/FOXP3 (+) cell ratio is a predictive marker for survival in patients with colorectal cancer, *Cancer Immunol. Immunother.* 59 (5) (2010) 653–661.
- [47] F. Echterdiek, J. Janikovits, L. Staffa, M. Müller, B. Lahrman, M. Fröhshütz, et al., Low density of FOXP3-positive T cells in normal colonic mucosa is related to the presence of beta2-microglobulin mutations in Lynch syndrome-associated colorectal cancer, *Oncoimmunology* 5 (2) (2016), e1075692.
- [48] K. Bauer, N. Nelius, M. Reuschenbach, M. Koch, J. Weitz, G. Steinert, et al., T cell responses against microsatellite instability-induced frameshift peptides and influence of regulatory T cells in colorectal cancer, *Cancer Immunol. Immunother.* 62 (1) (2013) 27–37.
- [49] S. Michel, A. Benner, M. Tariverdian, N. Wentzensen, P. Hoefler, T. Pommerenke, et al., High density of FOXP3-positive T cells infiltrating colorectal cancers with microsatellite instability, *Br. J. Cancer* 99 (11) (2008) 1867–1873.
- [50] N. Komatsu, K. Okamoto, S. Sawa, T. Nakashima, M. Oh-hora, T. Kodama, et al., Pathogenic conversion of Foxp3+ T cells into TH17 cells in autoimmune arthritis, *Nat. Med.* 20 (1) (2014) 62–68.
- [51] S. Le Gouvello, S. Bastuji-Garin, N. Aloulou, H. Mansour, M.T. Chaumette, F. Berrehar, et al., High prevalence of Foxp3 and IL17 in MMR-proficient colorectal carcinomas, *Gut* 57 (6) (2008) 772–779.
- [52] M.L. Palomba, Active immunotherapy: current state of the art in vaccine approaches for NHL, *Curr. Oncol. Rep.* 14 (5) (2012) 433–440.
- [53] D.J. Andorsky, R.E. Yamada, J. Said, G.S. Pinkus, D.J. Betting, J.M. Timmerman, Programmed death ligand 1 is expressed by non-hodgkin lymphomas and inhibits the activity of tumor-associated T cells, *Clin. Cancer Res.* 17 (13) (2011) 4232–4244.
- [54] D.T. Le, J.N. Uram, H. Wang, B.R. Bartlett, H. Kemberling, A.D. Eyring, et al., PD-1 blockade in tumors with mismatch-repair deficiency, *N. Engl. J. Med.* 372 (26) (2015) 2509–2520.
- [55] S.L. Topalian, F.S. Hodi, J.R. Brahmer, S.N. Gettinger, D.C. Smith, D.F. McDermott, et al., Safety, activity, and immune correlates of anti-PD-1 antibody in cancer, *N. Engl. J. Med.* 366 (26) (2012) 2443–2454.
- [56] R.J. Motzer, B. Escudier, D.F. McDermott, S. George, H.J. Hammers, S. Srinivas, et al., Nivolumab versus everolimus in advanced renal-cell carcinoma, *N. Engl. J. Med.* 373 (19) (2015) 1803–1813.
- [57] J. Brahmer, K.L. Reckamp, P. Baas, L. Crinò, W.E. Eberhardt, E. Poddubskaia, et al., Nivolumab versus docetaxel in advanced squamous-cell non-small-cell lung cancer, *N. Engl. J. Med.* 373 (2) (2015) 123–135.
- [58] J. Larkin, V. Chiarion-Sileni, R. Gonzalez, J.J. Grob, C.L. Cowey, C.D. Lao, et al., Combined nivolumab and ipilimumab or monotherapy in untreated melanoma, *N. Engl. J. Med.* 373 (1) (2015) 23–34.
- [59] E. Bouffet, V. Larouche, B.B. Campbell, D. Merico, R. de Borja, M. Aronson, et al., Immune checkpoint inhibition for hypermutant glioblastoma multiforme resulting from germline biallelic mismatch repair deficiency, *J. Clin. Oncol.* 34 (19) (2016) 2206–2211.
- [60] M.M. Gubin, R.D. Schreiber, *CANCER*. The odds of immunotherapy success, *Science* 350 (6257) (2015) 158–159.
- [61] S. Kelderman, T.N. Schumacher, P. Kvistborg, Mismatch repair-deficient cancers are targets for anti-PD-1 therapy, *Cancer Cell* 28 (1) (2015) 11–13.
- [62] I. Saeterdal, J. Bjørheim, K. Lislerud, M.K. Gjertsen, I.K. Bukholm, O.C. Olsen, et al., Frameshift-mutation-derived peptides as tumor-specific antigens in inherited and spontaneous colorectal cancer, *Proc. Natl. Acad. Sci. U. S. A.* 98 (23) (2001) 13255–13260.
- [63] N. Amayiri, U. Tabori, B. Campbell, D. Bakry, M. Aronson, C. Durno, et al., High frequency of mismatch repair deficiency among pediatric high grade gliomas in Jordan, *Int. J. Cancer* 138 (2) (2016) 380–385.
- [64] E. Matsuki, A. Younes, Checkpoint inhibitors and other immune therapies for hodgkin and non-hodgkin lymphoma, *Curr. Treat. Options Oncol.* 17 (6) (2016) 31.
- [65] S.M. Ansell, A.M. Lesokhin, I. Borrello, A. Halwani, E.C. Scott, M. Gutierrez, et al., PD-1 blockade with nivolumab in relapsed or refractory Hodgkin's lymphoma, *N. Engl. J. Med.* 372 (4) (2015) 311–319.
- [66] C.E. Sorge, J.K. McDaniel, A.C. Xavier, Targeted therapies for the treatment of pediatric non-hodgkin lymphomas: present and future, *Pharm. (Basel)* 9 (2) (2016).
- [67] J. Kline, M.R. Bishop, Update on checkpoint blockade therapy for lymphoma, *J. Immunother. Cancer* 3 (2015) 33.
- [68] K.F. Bol, G. Schreiber, W.R. Gerritsen, I.J.M. de Vries, C.G. Figdor, Dendritic cell-based immunotherapy: state of the art and beyond, *Clin. Cancer Res.* 22 (8) (2016) 1897–1906.
- [69] S. Anguille, E.L. Smits, E. Lion, V.F. van Tendeloo, Z.N. Berneman, Clinical use of dendritic cells for cancer therapy, *Lancet Oncol.* 15 (7) (2014) e257–e267.
- [70] M. Tagliamonte, A. Petrizzo, M.L. Tornesello, F.M. Buonaguro, L. Buonaguro, Antigen-specific vaccines for cancer treatment, *Hum. Vaccin Immunother.* 10 (11) (2014) 3332–3346.
- [71] P. Kantoff, C.S. Higano, N.D. Shore, E.R. Berger, E.J. Small, D.F. Penson, et al., Sipuleucel-T immunotherapy for castration-resistant prostate cancer, *N. Engl. J. Med.* 363 (5) (2010) 411–422.
- [72] S. Boudewijns, M. Bloemendal, W.R. Gerritsen, I.J.M. de Vries, G. Schreiber, Dendritic cell vaccination in melanoma patients: from promising results to future perspectives, *Hum. Vaccin. Immunother.* 12 (10) (2016) 2523–2528.
- [73] M. von Knebel Doeberitz, M. Kloor, Towards a vaccine to prevent cancer in Lynch syndrome patients, *Fam. Cancer* 12 (2) (2013) 307–312.
- [74] M. Kloor, The immune biology of microsatellite-unstable cancer, *Trends Cancer* 2 (3) (2016) 121–133.
- [75] Y. Schwitalle, M. Kloor, S. Eiermann, M. Linnebacher, P. Kienle, H.P. Knaebel, et al., Immune response against frameshift-induced neopeptides in HNPCC patients and healthy HNPCC mutation carriers, *Gastroenterology* 134 (4) (2008) 988–997.
- [76] N. Rahner, G. Höfle, C. Högenauer, C. Lackner, V. Steinke, M. Sengteller, et al., Compound heterozygosity for two MSH6 mutations in a patient with early onset colorectal cancer, vitiligo and systemic lupus erythematosus, *Am. J. Med. Genet. A* 146A (10) (2008) 1314–1319.
- [77] J. Plaschke, M. Linnebacher, M. Kloor, J. Gebert, F.W. Cremer, S. Tinschert, et al., Compound heterozygosity for two MSH6 mutations in a patient with early onset of HNPCC-associated cancers, but without hematological malignancy and brain tumor, *Eur. J. Hum. Genet.* 14 (5) (2006) 561–566.
- [78] A.R. Sanchez-Paulete, F.J. Cueto, M. Martínez-López, S. Labiano, A. Morales-Kastresana, M.E. Rodríguez-Ruiz, et al., Cancer immunotherapy with immunomodulatory anti-CD137 and anti-PD-1 monoclonal antibodies requires BATF3-dependent dendritic cells, *Cancer Discov.* 6 (1) (2016) 71–79.
- [79] L. Pierret, S. Wilgenhof, J. Corthals, T. Roelandt, K. Thielemans, B. Neyns, Correlation between prior therapeutic dendritic cell vaccination and the outcome of patients with metastatic melanoma treated with ipilimumab, *J. Clin. Oncol.* 27 (2009) abstr e20006.
- [80] J. Yuan, B. Ginsberg, D. Page, Y. Li, T. Rasalan, H.F. Gallardo, et al., CTLA-4 blockade increases antigen-specific CD8(+) T cells in prevaccinated patients with melanoma: three cases, *Cancer Immunol. Immunother.* 60 (8) (2011) 1137–1146.
- [81] S. Wilgenhof, J. Corthals, C. Heirman, N. van Baren, S. Lucas, P. Kvistborg, et al., Phase II study of autologous monocyte-derived mRNA electroporated dendritic cells (TriMixDC-MEL) plus ipilimumab in patients with pretreated advanced melanoma, *J. Clin. Oncol.* 34 (12) (2016) 1330–1338.
- [82] A. Ribas, B. Comin-Anduix, B. Chmielowski, J. Jalil, P. de la Rocha, T.A. McCannel, et al., Dendritic cell vaccination combined with CTLA4 blockade in patients with metastatic melanoma, *Clin. Cancer Res.* 15 (19) (2009) 6267–6276.
- [83] A.J. van den Eertwegh, J. Versluis, H.P. van den Berg, S.J. Santegoets, R.J. van Moorselaar, T.M. van der Sluis, et al., Combined immunotherapy with granulocyte-macrophage colony-stimulating factor-transduced allogeneic prostate cancer cells and ipilimumab in patients with metastatic castration-resistant prostate cancer: a phase 1 dose-escalation trial, *Lancet Oncol.* 13 (5) (2012) 509–517.
- [84] R.A. Madan, M. Mohebtash, P.M. Arlen, M. Vergati, M. Rauckhorst, S.M. Steinberg, et al., Ipilimumab and a poxviral vaccine targeting prostate-specific antigen in metastatic castration-resistant prostate cancer: a phase 1 dose-escalation trial, *Lancet Oncol.* 13 (5) (2012) 501–508.