Contents lists available at ScienceDirect



Journal of Controlled Release

journal homepage: www.elsevier.com/locate/jconrel

The small molecular Kinase Inhibitor (smKI) drug class is very promising and rapidly expanding. All of these

drugs are administered orally. The clear relationship between structure and function has led to drugs with a gen-

eral low intrinsic solubility. The majority of the commercial pharmaceutical formulations of the smKIs are phys-

ical mixtures that are limited by the low drug solubility of a salt form. This class of drugs is therefore characterized

by an impaired and variable bioavailability rendering them costly and their therapies suboptimal. New formulations are sparingly being reported in literature and patents. The presented data suggests that continued research

into formulation design can help to develop more efficient and cost-effective smKI formulation. Moreover, it may



© 2016 Elsevier B.V. All rights reserved.

CrossMark

Review article Inherent formulation issues of kinase inhibitors

M. Herbrink *, J.H.M. Schellens, J.H. Beijnen, B. Nuijen

Department of Pharmacy and Pharmacology, Netherlands Cancer Institute-Antoni van Leeuwenhoek, Louwesweg 6, 1006 BK Amsterdam, The Netherlands

ARTICLE INFO

ABSTRACT

Article history: Received 22 July 2016 Received in revised form 24 August 2016 Accepted 26 August 2016 Available online 28 August 2016

Keywords: Tyrosine Kinase Inhibitor Chemotherapy Formulation Bioavailability

Contents

1.	Introduction	118				
2. Bioavailability.						
3.	Physicochemistry	119				
	3.1. Essential structures	119				
	3.2. Solubility	121				
	3.3. pH-dependent solubility	121				
	3.4. Salt and free base polymorphs	121				
	3.5. Solvates	121				
4.	Formulations	121				
	4.1. 4.1. Commercial physical mixture formulations	122				
	4.2. Alternative formulations	123				
	4.2.1. Vemurafenib	123				
	4.2.2. Regorafenib	123				
	4.2.3. Nintedanib	123				
5.	New formulations in literature and patents	123				
	5.1. Solid dispersions	123				
	5.2. Crystalline stabilization	123				
	5.3. Cocrystals	123				
6.	Discussion	124				
7.	. Conclusions					
Refe	References					

also be of help in the future design of the formulations of new smKIs.

1. Introduction

Small molecular Kinase Inhibitors (smKIs) form a promising and rapidly expanding class of drugs [1,2]. The drugs target specific parts of Kinase receptor proteins that play an important part in the intracellular growth signaling pathways in tumor and immune cells [3–5]. After the first drug approval by the United States Food and

* Corresponding author. *E-mail address*: Maikel.Herbrink@slz.nl (M. Herbrink). Drug Administration (FDA) of Imatinib in 2001 [6,7], the number of drugs approved by both the FDA and European Medicines Agency (EMA) are now nearing 30, listed in Table 1. Many more smKIs are being investigated in clinical trials and are expected to be approved in the coming years. A range of small molecular inhibitors have proven to be useful in the therapy of certain types of cancer. Additionally, smKIs may be prescribed as alternatives when other therapeutic options have failed or are deemed inappropriate [8]. A few compounds are (also) applied in the therapy of immunomodulated diseases [9–12] and may even have a future in the therapy of diseases such as diabetes mellitus [13].

All of the smKIs are without exception administered orally. This has great advantages in terms of patient convenience and cost reduction [14–16]. It presents however serious difficulties for compounds with a low solubility and/or permeability. These drugs are hindered by a reduced and variable bioavailability. This may cause drug plasma concentrations to be ineffectively low or toxically high with all due consequences [17]. Understanding and controlling the parameters of solubility and permeability can therefore have a profound influence on patient plasma drug levels.

The smKIs have been designed using high-throughput screening and combinatorial chemistry from which the intricate structures and inherent solubility issues originate [18]. These means are used in the drug discovery of other drug classes as well, *e.g.* drugs acting on the central nervous system that also experience a problematic solubility [19].

Drug solubility and the dissolution process are affected by a plethora of factors with clinical implications. A number of these factors are inherent to the smKI structure and function. It follows that, as drug dissolution is most often the primary determinant for the smKI bioavailability, there is an apparent link between the high specificity of the smKIs and their impaired absorption into the systemic circulation.

This article will first present the overall problematic biopharmaceutical properties of the smKI drug group. Secondly, it will discuss the characteristic structural elements that are responsible for the solubility behavior of the smKIs. It will continue by reviewing the current commercial formulations along with alternative investigational formulations. This article aims to underline that although high specificity can, in many cases, place a challenging strain on drug solubility and bioavailability, different and innovative formulation techniques may present possible solutions to some of these issues. Literature offers several reviews that address (pre)formulation challenges for poorly soluble drugs in general [20–22]. This article focuses on smKIs in particular. To the authors' knowledge, this review is the first to combine literature and patent research on the solubility and formulation of smKI compounds from a pharmaceutical perspective.

2. Bioavailability

In the process of reaching the therapeutic target, the first step after oral administration and the disintegration of the dosage form is always the dissolution of the drug substance [75]. The second step, absorption, only takes place with the dissolved portion of the drug. Thus, poor drug solubility can be one of the main causes for a low and variable uptake of a drug into the systemic circulation, *i.e.* a low and variable bioavailability. This is generally true for the smKIs, as listed in Table 1. This reflects back in their BCS (the Biopharmaceutical Classification System)-classes of which most are II (solubility hindered bioavailability) or IV (solubility and permeability hindered bioavailability) [76,77].

Factors such as presystemic metabolism and mediated transport by transporter proteins may also play a part in reducing a drug's bioavailability. The combination of these factors is reviewed elsewhere [78,79].

3. Physicochemistry

3.1. Essential structures

In the past few decades the role of signal proteins in the homeostasis of tumors became more and more apparent [80,81]. The advancement of the diverse techniques and possibilities of molecular modelling have led to a therapeutic target-based drug discovery regime [82,83]. With it, structure-activity relations for inhibitory molecules for these proteins were assessed. Key in these relations is the binding of the lead drug molecule to the receptor and the inhibitory action thereon [84]. The latter can be viewed as a dependent of the first but does not necessarily result from the same molecular structure. The independence is illustrated by lenvatinib and sunitinib; they inhibit VEGFR2 by binding to the ATP-binding site with their core structure. Additional binding through a nearby structure in both compounds gives them their difference in residence time without influencing the inhibition itself [85].

The resulting collection of mainly lipophilic structures now forms the backbone of the majority of the KIs. Some molecular structures and scaffolds are found throughout the current marketed collection of smKIs. Even though the drugs inhibit a wide variety of proteins, the necessary structures to do so are similar. Table 2 lists the molecular structures of the free base smKIs ordered by primary target. Bold print indicates the proven binding moieties in the smKIs that are responsible for the inhibitory effect.

The bold printed structures in Table 2 have been shown in molecular docking and *in vitro* crystallization studies to be critical in receptor binding and inhibition [22–53]. These include highly lipophilic moieties such as (substituted) phenyls, aromatic amines, biaryl constructs and heterocyclic aromatics. Using these structures as scaffolds, a great number of studies have designed new smKIs by adding different side groups

Table 1

Approved smKIs (registered trademarks) by the FDA on March 20th 2016. Appointed BCS-classes are taken from registration documents.

Compound	Tradename	BCS ^a	Bioavailability(%)	Compound	Tradename	BSC ^a	Bioavailability(%)
Imatinib	Gleevec	Ι	98 [23,24]	Regorafenib	Stivarga	II	[25,26]
Gefitinib	Iressa	II	60 [27,28]	Tofacitinib	Xeljanz	III	74 [11,12]
Erlotinib	Tarceva	II	60 [29,30]	Bosutinib	Bosulif	IV	[31,32]
Sorafenib	Nexavar	II/IV	[33,34]	Axitinib	Inlyta	II	58 [35,36]
Dasatinib	Sprycel	II	[37,38]	Ibrutinib	Imbruvica	II	2.9 [39,40]
Sunitinib	Sutent	IV	[41]	Afatinib	Giotrif	I/III	[42,43]
Nilotinib	Tasigna	IV	30 ^b [44,45]	Dabrafenib	Tafinlar	II	95 [46,47]
Lapatinib	Tyverb	II	<25 ^b [48,49]	Trametinib	Mekinist	IV	72 [50,51]
Pazopanib	Votrient	II	14-39 [52,53]	Nintedanib	Vargatef	II/IV	5 [54,55]
Ruxolitinib	Jakavi	Ι	>95 [9,10]	Ceritinib	Zykadia	IV	25 ^b [56,57]
Crizotinib	Xalkori	IV	43 [58,59]	Alectinib	Alecensa	II/IV	37 [60]
Vemurafenib	Zelboraf	IV	[61,62]	Cobimetinib	Cotellic	I/III	46 [63,64]
Vandetanib	Caprelsa	II	[65,66]	Osimertinib	Tagrisso	III	[67,68]
Ponatinib	Iclusig	II	[69,70]	Lenvatinib	Lenvima	II/IV	[71,72]
Cabozantinib	Cometriq	II	[73,74]				

^a BCS, Biopharmaceutical Classification System.

^b Estimated/Based on mass balance.

Table 2

Effective moieties in molecular structures of smKI classes.

Primary target	Molecular structure(s)
BCR-Abl ^a [86–89]	$ \begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & $
EGFR ^b [90–92]	-N P ponatinib
	$\begin{array}{c} & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ \end{array}$
VEGFR ^b , PDGFR ^b [85,93–102]	afatinib osimertinib H_{HN} H_{HN}
	$ \begin{array}{c} \begin{array}{c} & & \\ & \\ & \\ & \\ & \\ & \\ \end{array} \end{array} \begin{array}{c} \\ & \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ $
	PHN V NH NH NH NH NH NH NH2 NH2 NH2 NH2 NH2 NH2 NH2 N
B-Raf ^b [103,104]	$C_{I} \xrightarrow{F}_{N} \xrightarrow{F}$
Jak ^b [105–108]	
ALK ^b [109–113]	ruxolitinib $F \xrightarrow{Cl}_{Cl} \xrightarrow{O}_{S_{20}} \xrightarrow{NH}_{N-N} \xrightarrow{Cl}_{N+N} \xrightarrow{NH}_{N-N} \xrightarrow{NH}_{N-N} \xrightarrow{N}_{N+N} \xrightarrow{N}_{N} N$
MEK ^b [114–116]	crizotinib cerumo alectinib

Table 2 (continued)



^a BCR-Abl, Breakpoint Cluster Region- Abelson murine lukemiaviral oncogene.

^b EGFR, Endothelial Growth Factor Receptor; VEGFR, Vascular Endothelial Growth Factor Receptor; PDGFR, Platelet Derived Grwoth Factor Receptor; B-raf, v-raf murine sarcoma viral oncogene homolog B1; Jak, Janus Kinase; ALK, Anaplastic Lymphoma Kinase; MEK, Mitogen activated protein Kinase.

[117–119]. Combining scaffolds to create a new multi-kinase-targeting smKI is also a commonly used strategy [120].

The lipophilic functional moieties and additional scaffold structures make up the larger part of the smKI molecular structure. To improve the overall aqueous solubility of a drug, drug development may include the evaluation of the possible addition of hydrophilic moieties to the core structure [121]. This addition of more hydrophilic structures may not be feasible due to the presence of hydrophobic residues that are essential for binding areas of the inhibitors however [122–125]. Adjustments to the KI chemical structures to improve the biopharmaceutical behavior can therefore only go so far. This has led to a continuous search for a delicate balance between pharmaceutical and pharmacological properties, most often resulting in the favor of the latter. As a consequence, the aqueous solubility of the free base smKI remains generally low. A recent study shows that this balance may not always lean towards poor solubility and that a combination of high inhibitor activity and hydrophilicity can be achieved for some KI structures [126].

Additionally, enhancing aqueous solubility can be achieved by constructing a pro-drug from the drug substance. Here, a solubility-improving side group is attached to the active drug, which is later removed by some form of metabolism *in vivo* after absorption [127]. Although this has not been used in the marketed smKIs, it is currently being examined for smKIs [128].

3.2. Solubility

The aqueous solubility of the smKIs is strongly influenced by environmental factors, as described in the following paragraphs. Additionally, the time points of solubility measurements determine whether an equilibrium or a dynamic value is assessed, which may differ significantly [51]. A single value can therefore not be appointed to the aqueous solubility. Reported values should be evaluated carefully, as the circumstances under which it was measured may not be adequately described.

3.3. pH-dependent solubility

The majority of the smKI scaffolds consists of nitrogen-based heterocyclic systems. An important consequence of the nitrogen-containing core inhibitory structures in combination with secondary amine moieties is an overall pH-dependent aqueous solubility.

The majority of the pK_a values of the ionizable nitrogen structures are sufficiently low to qualify them as relatively weak bases [129,130]. Table 3 lists the currently known values. The higher pK_a values commonly concern nitrogen groups that become neutral when protonated, whilst the lower pK_a values correspond to groups that attain a positive charge by protonation. For most compounds this means a relative high solubility in acidic media and a harshly reduced solubility in more alkaline environments [131–133]. Vemurafenib, Regorafenib and Trametinib are exceptions herein with an almost pH-independent solubility. This is due to their pK_a values that are either above or below the physiological pH-range.

3.4. Salt and free base polymorphs

The presence of ionizable groups in most of the smKI drug structure made the conversion to salts feasible [134]. The commercial formulations of many smKIs contain a salt in a designated stable polymorph form (Table 3). This led to a dramatic increase in solubility for some compounds, e.g. Imatinib, Tofacitinib and Afatinib, of which the free base forms are very poorly soluble (BCS II or IV). Their respective mesvlate, citrate and dimaleate salts upgraded them to BCS-I or III. For most of the other drugs salt formation did not significantly increase solubility and they remain BCS II or IV. The reasons for using particular salt or free base polymorphs in formulations are, however often not provided in registrations texts or patents. New salt and free base polymorphs with higher solubilities are being patented and reported in literature before and after market authorization is obtained. It seems that, at least in terms of solubility, better options were and are available. Comparisons in dissolution behavior and bioavailability between various salt forms are not yet accessible. The alternative salts may prove useful in future formulation improvements however.

3.5. Solvates

Solvates are crystalline solids that contain stoichiometric or nonstoichiometric proportions of a solvent within their crystal structure. When this solvent is water, the solvate is termed a hydrate. When no solvent or water is present, a compound is termed an anhydrate. Solvates, hydrates and solventless crystals can differ significantly in solubility [135]. Most of the smKIs are formulated as anhydrates. Data on dissolution performance of solvates and hydrates are only rarely published. Dasatinib, Nilotinib, Lapatinib and Bosutinib are present as monohydrate crystals in their respective formulations. After the approval of Sprycel® in 2006, a way to produce Dasatinib anhydrate was found and this form was shown to be 2.4 times more soluble than the monohydrate variant [136]. Trametinib is formulated as a stoichiometric DMSO-solvate. In rats this increased the bioavailability of Trametinib 30-fold compared to the unsolvated form. In aqueous environment, the dissolved Trametinib.DMSO slowly precipitates as the much less soluble unsolvated form [51].

4. Formulations

Partly as a consequence of the before-mentioned factors, the solubility of a drug salt polymorph can be described thermodynamically. The following combined equation illustrates this [137]:

$$-\log X = \frac{\Delta H_f}{2.303RT} \left(\frac{T_0 - T}{T_0 T} \right) + \frac{V\Phi}{2.303RT} (\delta_1 - \delta_2)^2 \tag{1}$$

X, dissolved molar fraction; ΔH_f , latent heat of fusion (heat absorbed during melting); R, gas constant; T, temperature; TO, melting point of solute; V, molar volume of liquid solute; Φ , volume fraction of the solvent; δ , solubility parameter (expression of cohesion between molecules).

Table 3

Reported solubilities of smKIs as pure drug substance at given pH values and their types of commercial formulation.

Compound	pK _a	рН	Solubility (mg/mL)	Dosage form	Formulation composition
Imatinib mesylate	Х	≤5.5	>1.6 ^m [24]	T/C	Patent expired
Gefitinib FB ^a	5.4; 7.2 [28]	1.0; 7.0	21 ^m ; <0.001 [28]	Т	L, MC, CS, P, SLS, MS
Erlotinib hydrochloride	5.4 [155]	2.0	0.4 ^m [155]	Т	L, MC, SSG, SLS, MS
Sorafenib tosylate	X	1.0; 4.5	0.034 ^m ; 0.013 [33]	Т	MC, CS, H, SLS, MS
Dasatinib FB·H ₂ O	3.1; 6.8; 10.8 [156]	2.6; 6.0	18.4 ^m ; 0.008 [37]	Т	L, MC, CS, HPC, MS
Sunitinib malate	9.0 [41]	1.2-6.8	>25 ^m [41]	С	MN, P, CS, MS
Nilotinib hydrochloride.H ₂ O	2.1; 5.4 [157]	1.0; 4.5	0.28 ^m ; <0.1 [45]	С	L, CP, PX, SC, MS
Lapatinib ditosilate.H ₂ O	5; 7.2 [158]	1.0; w	10^{-6} ; 0.007 ^m [49,159]	Т	MC, P, SSG, MS
Pazopanib hydrochloride	2.1; 6.4; 10.2 [160]	1.1	0.65 ^m [160]	Т	MC, P, SSG, MS
Ruxolitinib phosphate	4.3; 11.8 [161]	W	X (highly soluble) [161]	Т	L, MC, SSG, MS, SC, HPC, P
Crizotinib FB	5.6; 9.4 [59]	1.6; 8.2	>10 ^m ; <0.1 [162]	С	MC, SC, CHP, SSG, MS
Vandetanib FB	5.2; 9.4 [147]	6.8; w	0.35 ^m ; 0.008 [65,147]	Т	MC, DCP, CP, P, MS
Ponatinib hydrochloride	2.8; 7.8 [70]	1.7; 7.5	7.8 ^m ; 0.16 * 10 ⁻³ [70]	Т	L, MC, SSG, SC, MS
Cabozantinib malate	Х	2;>3	0.11 ^m ; x (very low) [163]	С	CS, SSG, SC, SA, MC
Tofacitinib citrate	5.1 [164]	1.0; w	>28 ^m ; 2.9 [164]	Т	MC, L, CS, MS
Bosutinib FB ⋅ H ₂ O	7.9 [32]	<5.0; >5.0	X (high); X (reduced) [32]	Т	MC, CS, PX, P, MS
Axitinib FB	4.8 [36]	1.1; >6.0	$1.841^{\rm m}$; 0.2×10^{-3} [36]	Т	MC, L, CS, MS
Ibrutinib FB	3.8 [40]	1.2; 5.5	2 ^m ; 0.003 [40]	С	MC, CS, MS, SLS
Afatinib dimaleate	5.0; 8.2 [165]	<6.0;>7.0	>50 ^m ; 0.04 [165,166]	Т	L, MC, CP, SC, MS
Dabrafenib mesylate	1.5; 2.2; 6.6 [47]	1.0; 4–8	X (VSS); X (PI) [47]	С	MC, MS, SC
Trametinib FB.DMSO	0.3 [167]	1.2; 6.8	0.0004; 0.011 ^m [51]	Т	MC, MN, H, CS, MS, SLS, SC
Ceritinib FB	4.1; 9.7 [168]	1.0; 6.8	$11^{\rm m}$; 0.2 * 10^{-3} [57]	С	MC, HPC, SSG, MS, SC
Alectinib hydrochloride	7.1 [169]	1.0; 6.8	0.0013; 0.0279 ^m [60]	С	L, HPC, SLS, MS, CMC
Cobimetinib fumarate	8.9 [170]	1.0; 6.8	48.21 ^m ; 0.78 [64]	Т	L, MC, CS, MS
Osimertinib mesylate	4.4; 9.5 [171]	1.2; 4.5	>3; >11 ^m [68]	Т	MN, H, SSF
Lenvatinib mesylate	5.1 [72]	<3.0; 3–7	X (VSS); < 0.096 [71]	С	MN, MC, CC, H, TC
Vemurafenib MPB ^b	7.9; 11.1 [62]	1.0; 6.8	<0.3 * 10 ⁻³ ; 0.5 * 10 ^{-3m} [62]	Т	Solid solution
Regorafenib FB	Х	Х	Х	Т	Solid dispersion
Nintedanib esilate	5.6; 9.4 [172]	1.0; ≥ 6.8	5 ^m ; 0.011 [55]	С	Lipophilic suspension

^a FB = Free Base.

^b MPB = MicroPrecipitated Bulk; M = reported maximum solubility; w = in water, pH not specified; x = value not reported; () = description given; VSS, Very slightly soluble; PI, practically insoluble; T, Tablet; C, Capsule; L, lactose; MC; Microcrystalline cellulose; CS, Crosscarmellose sodium; P, Povidone; SLS, Sodium Lauryl Sulphate; MS, Magnesium stearate; SSG. Sodium starch glycolate; H, Hypromellose; HPC, hydroxypropylcellulose; MN, Mannitol; CP, Crospovidone; PX, Poloxamer 188; SC, Silica colloidalis anhydrica; CHP, Calcium hydrogen phosphate; DCP, Dibasic calcium phosphate; SA, Stearic acid; CMC, Carboxymethylcellulose calcium; SSF, Sodium stearyl fumarate; CC, calcium carbonate; TC, Talc.

Eq. (1) clearly expresses the role of solid state crystallinity in a compound's solubility through the heat of fusion and the melting point of the drugs. The heat of fusion is the heat necessary to transform a compound from its solid to its liquid state at its melting point. Both the melting point and the heat of fusion are measures of the bond strength in the drug crystal. Stronger bonds between molecules in a crystal structure will increase both parameters and subsequently lower solubility [138]. The solubility parameters in Eq. (1) denote the significance of the molecular structure and the resulting intermolecular cohesive forces in drug and solvent in the solubility end term. The parameters are descriptors of the interaction between molecules of the drug and the solvent. If such interactions are alike in the separate drug and solvent [139]. The last term in Eq. (1) is low in such a case and the resulting solubility is relatively high.

In a physiological environment, a pharmaceutical solution may be regarded as dilute with a near-to-constant temperature [140,141]. In that situation, the molar volume approaches unity and the volume fraction may be disregarded [137]. This allows for the following simplification to a qualitative equation:

$$-\log X \approx \Delta H_f \left(\frac{T_0 - T}{T_0 T}\right) + (\delta_1 - \delta_2)^2 \tag{2}$$

Eq. (2) shows that a low heat of fusion and melting point are beneficial to a compound's solubility [142,143]. Additionally, the solubility of a drug can be further increased by creating more similarity between the solubility parameters of the drug and the solvent [144,145].

4.1. 4.1. Commercial physical mixture formulations

The commercial immediate release formulations of the smKIs are almost all designed as physical mixtures. They contain a crystal solid form.

Since crystallinity is a strong determinant of solubility, the heat of fusion and the melting temperature of crystal polymorphs are closely correlated with it. Although not known for all compounds, the melting points are relatively high [26,36,62,146,147]. Furthermore, the difference between the solubility parameters of the drugs and the physiological solvent is expected to be, based on structure, relatively large [148–150]. The outcome is a very poor solubility, as is presented in Table 3. The excipients present in the physical mixtures, such as polymers and surfactants, may have some influence on the composition of the physiological solvent in the direct environment of the dissolving drug. This may lower the difference in the last term of Eq. (2). In time this effect dissipates due to diffusion of both compounds and the drug will recrystallize [151]. Table 3 lists the excipients present in the commercial formulations. As far as could be assessed, most manufacturing processes entail dry or wet mixing of the formulation ingredients and optimization of the drug particle size [152]. The latter only increases dissolution speed as it does not alter the resultant solubility through any terms in Eq. (2). The accessibility of solubility data and dissolution curves is limited in literature and patents, so the effect of the formulation on the smKIs dissolution performance is largely unknown in the public domains. The proposed dissolution methods by the FDA for bioequivalence testing of smKI formulations offer clarification to some extent however [153]. The dissolution media are all more voluminous than the stomach. In many cases the pH is adjusted to highly acidic conditions. And for some drugs surfactants are added. Although these methods are designed for quality control purposes, they provide further evidence of poor drug solubility. The non-bioequivalent dissolution media are designed to enable appropriate drug solubility (>85%) for the comparison of individual drug product batches [153]. When acidification and addition of surfactants to the media are needed to achieve this, drug solubility in bioequivalent media is likely to be significantly low. The formulation of lenvatinib is an exceptional case because it was designed with special focus on its stability. Lenvatinib has the tendency to gelate and decompose under humid and heated storage conditions, unlike the other smKIs. Calcium carbonate and mannitol were chosen as filler/ disintegrant and diluent to keep the overall hygroscopicity of the formulation low. This combination of the carrier and the hypromellose capsule make water uptake by the formulation unlikely and thus prevents gelation and hydrolytic degradation, both of which are detrimental to the bioavailability [71,154]. This capsule, which is additionally kept in a protective blister, can be stored at room temperature and humidity [71].

4.2. Alternative formulations

Changing the solid state of the drug may radically alter the first term in Eq. (2). When a move is made from the highly structured crystal form to the less rigid amorphous form the solubility can increase. This is due to the fact that amorphous materials have a melting range instead of a melting point and lack a definite heat of fusion [173]. Amorphous forms of drugs may be prepared in various ways by incorporating them in a polymer matrix in order to retain their amorphous state [174,175]. Such a system is termed a solid dispersion [176]. The commercial formulations of Vemurafenib and Regorafenib (4.2.1. and 4.2.2.) are designed like this. Additional advantages of a solid dispersion are particle size reduction, increased wettability, reduced aggregation and agglomeration and a decrease in the difference in solubility parameters through the polymer changing the physiological solvent. All these factors combined may increase the solubility long enough to improve drug absorption.

During clinical studies of three smKIs it became apparent that a simple physical mixture did not suffice in creating an adequate plasma level. These cases are discussed below.

4.2.1. Vemurafenib

Crystalline free base vemurafenib is known to exist in several polymorphs and solvates [61,62]. The most thermodynamically stable form II is practically insoluble in water with pH-values ranging from 1.1 to 9. The first clinical study with Vemurafenib was performed with the more soluble crystalline form I in a micronized capsule formulation. Form I transformed to form II over time and the observed bioavailability was low [61,62]. Shah et al. describes the development of the amorphous Vemurafenib formulation that is now used in the marketed Zelboraf® [177]. The amorphous solid dispersion was prepared by a solvent-controlled coprecipitation process as a so-called microprecipitated bulk powder. The amorphous Vemurafenib is herein stabilized by a hypromellose acetate succinate matrix to prevent crystallization [178]. Compared to the crystalline formulation, the solid dispersion demonstrated a significantly improved solubility and a five-fold increase in exposure.

4.2.2. Regorafenib

Regorafenib as a monohydrate salt is poorly soluble in water at <0.1 mg/mL [179]. The possible consequences of the poor solubility for the bioavailability were recognized early on in the formulation development. Therefore, a series of physical mixtures and solid dispersions were tested. A solid dispersion of Regorafenib in PVP 25 with a composition of 1:4 was chosen after *in vitro* dissolution screening and the assessment of rat pharmacokinetics [180].

4.2.3. Nintedanib

The drug compound Nintedanib esilate is suspended in an oily base in its commercial formulation. Nintedanib is suspended in a mixture of medium-chain triglycerides (carrier) and hard fat (thickener) [181, 182]. A patent from 2009 describes the development of the formulation [183]. It stated that hydrolytic degradation seems to be problematic for the compound. In combination with the high drug load, a lipophilic carrier suspension in a hydrophilic capsule was deemed appropriate. The formulation had a higher bioavailability than the tested hydrophilic and lipophilic-surfactant systems in rats.

5. New formulations in literature and patents

Patents and exclusivities are still covering all of the smKls, except for Imatinib [184,185]. While registered and approved alternative formulations are a long way off, there is a limited body of research published and patented at the time of writing. This section will briefly discuss the most frequently reported and patented oral formulation types.

5.1. Solid dispersions

Producing and characterizing the amorphous form of the smKIs is described throughout the patent body. Using that amorphous form in a pharmaceutical formulation is less frequently reported. This can be due to a too unstable amorphous form, a non-superior solubility or simply because the terrain is still unexplored. Xspray microparticles is one of the very few that give a description of the effect of amorphization on the dissolution of some smKIs [186]. The inventors use supercritical fluid precipitation to produce solid dispersions of Axitinib, Crizotinib, Dasatinib, Erlotinib, Gefitinib, Lapatinib, Nilotinib, Pazopanib, Sorafenib and Vemurafenib with different polymers. The patent presents a significant solubilization of the investigated compounds by the incorporation into polymeric matrices. Godugu et al. describe a spray dried solid dispersion of Gefitinib that yields a 9-fold increase in rat AUC compared to free base Gefitinib [187]. The group of Truong found that a spray dried formulation of amorphous Sorafenib, a graft polymer and SLS increased the AUC by 1.8-fold in rats [188]. Nanologica patented a nanoporous formulation with loaded amorphous Dasatinib [189]. They report no dissolution data, but state that more Dasatinib is released in Simulated Intestinal Fluid from their formulation than form crystalline drug. Song et al. prepared various solid dispersion of Lapatinib and showed an increased solubility of the products in water with 0.2% SDS [190]. Unsolvated Trametinib in a spray dried formulation had better dissolution characteristics than the commercial formulation, as patented by Ratiopharm GmBH [191].

5.2. Crystalline stabilization

A small number of patents describe improved dissolution characteristics for set of smKIs by using excipients that stabilize crystalline polymorphs as solids or as solutes. Stabilization of an unstable, more soluble polymorph of Erlotinib hydrochloride with a hydrophilic polymer can lead to better dissolution profiles. This was shown by Synthon BV [192]. Liu et al. found that the solubility of Sorafenib can be markedly increased by formulating it with Polyvinylpyrrolidone vinyl acetate copolymer (PVP-VA). The drug-polymer interaction in solution provides a supersaturated state that nearly doubles the AUC in beagle dogs [193]. The development of sustained release dosage forms of the relatively soluble Tofacitinib citrate was carried out by Pfizer [194]. This may well be an example of the development route of smKIs once the solubility issues are under control.

5.3. Cocrystals

A cocrystal is defined as a homogenous crystalline material that is made up of two or more molecules in definite stoichiometric amounts held together by non-covalent forces [195]. The physicochemistry of the so-called non-covalent derivative may be very different from a drug salt form that is held together by ionic forces [196]. A patent filed in 2015 showed an increase in solubility of Gefitinib in cocrystalline form with certain carboxylic acids [197]. Basf Se prepared several cocrystals of Dasatinib and showed that especially the combination with methyl gallate increased the aqueous solubility to 42 µg/mL from 0.36 µg/mL of the dasatinib free base monohydrate [198]. The same was done for Nilotinib, which showed that the cocrystal with maleic acid had an almost 6-fold increase in solubility compared to the hydrochloride form [199]. Neither of the two are backed up with bioavailability data. Cocrystalline forms also improve dissolution of Lapatinib. Fabbrica Italiana Sintetici shows that a cocrystal of Lapatinib with Adipic acid has a higher solubility than Lapatinib ditosylate. In rats, however, this did not lead to an improvement of the bioavailability [200].

6. Discussion

The currently approved smKIs have important places in clinical practice for registered indications. A large fraction of these compounds is under investigation for additional indications. The new and upcoming smKIs target many of the same kinases and are designed in a similar fashion as the already marketed ones. Their general physicochemical properties might not differ significantly from those of the older smKIs. A low and variable bioavailability is an enormous problem and may also lead to additional expenses in the patient treatment. High drug loading may be necessary to reach certain plasma levels for drugs with a low bioavailability with due consequences. The narrow therapeutic window of the drugs reflect in serious side effects and reduced activity above and below certain plasma concentration thresholds, respectively [17,201]. A variable bioavailability therefore often requires plasma levels to be monitored over a given time as part of a therapeutic drug monitoring (TDM) regimen which in part takes the advantages of oral therapy away. This is illustrated by the necessity of dose adjustments of smKIs in 25% of the patients treated with these drugs in our own institute.

Poor solubility may also pose a problem for smKIs that do not bind to the active site of the kinase that they inhibit. Such smKIs can bind to a regulatory site of a kinase, *e.g.* Rebastinib. The molecular structure of Rebastinib contains similar groups as the presented smKIs in Table 2, which are necessary for binding [202]. Solubility data of this drug is not yet available in the public domain. However, based on its structure, rebastinib is expected to be poorly soluble.

Plain and relatively uncomplicated formulations may be troublesome and can have very costly consequences during early clinical development. The first clinical trial with vemurafenib is exemplary. During this trial, it was discovered that the formulated crystal polymorph exhibited a low bioavailability. A new formulation with amorphous vemurafenib was produced and the clinical trials had to be repeated [61]. Although such a case is probably an exception, it is worth noticing that this might have been prevented.

With the incidence of cancer on the rise and the value of smKI therapy thoroughly established, the challenges of further improving the therapy are gaining value [203]. With a disease that has such significant consequences for patients, therapeutic uncertainties are all the more undesired. Predicting and addressing these uncertainties, preferably in an early stage of development, seems more than appropriate. Simple and straightforward formulations are surely preferred in terms of cost effectiveness of development and ease-of-production. The experience with the smKI group teaches the most valued lesson that overall cost effectiveness and the ease-of-treatment may not at all be benefitted by such choices however. Speed and efficiency are becoming characteristics of drug development [204]. The future challenges will therefore lie in the implementation of thorough screening of formulations and dosage forms into the overall drug development process. To ensure efficiency, a useful translation from in vitro dissolution data to the in vivo setting is needed. As in vitro-in vivo predictability from simple dissolution setups is often troublesome, the adoption of methods such as the gastrointestinal model TIM may prove to be valuable [205].

The solubility-induced variable bioavailability and the accompanying risks and costs may largely benefit from formulations with improved performance. While most patents and exclusivities are still pending and new or bioequivalent formulations are not yet eligible for approval, formulation research increasingly highlights the opportunities for improved drug forms of approved and to-be-approved smKIs.

7. Conclusions

Due to the very distinct targeting of the smKIs, biocompatible physicochemistry is driven to the edge. This places a strain upon bioavailability and presents challenges to formulation scientists.

'Classical' physical mixtures may work to achieve a relatively high bioavailability for some compounds, namely Imatinib, Ruxolitinib and Dabrafenib. This is certainly not the case for the majority of smKIs however. Bearing patents and exclusivities in mind, the past experiences can lead to new and innovative formulations that may provide further improvement of the efficacy of anti cancer treatment.

References

- M. Hojjat-Farsangi, Targeting non-receptor tyrosine kinases using small molecule inhibitors: an overview of recent advances, J. Drug Target. 24 (2016) 192–211, http://dx.doi.org/10.3109/1061186X.2015.1068319.
- [2] A.K.A. Gaumann, F. Kiefer, J. Alfer, S.A. Lang, E.K. Geissler, G. Breier, Receptor tyrosine kinase inhibitors: are they real tumor killers? Int. J. Cancer 138 (2016) 540–554, http://dx.doi.org/10.1002/ijc.29499.
- [3] G. Mirone, A. Shukla, G. Marfe, Signaling mechanisms of resistance to EGFR- and anti-Angiogenic inhibitors cancer, Crit. Rev. Oncol. Hematol. 97 (2016) 85–95, http://dx.doi.org/10.1016/j.critrevonc.2015.08.012.
- [4] R.R. Kudchadkar, K.S.M. Smalley, L.F. Glass, J.S. Trimble, V.K. Sondak, Targeted therapy in melanoma, Clin. Dermatol. 31 (2012) 200–208, http://dx.doi.org/10.1016/j. clindermatol.2012.08.013.
- [5] S.W. Tas, C.X. Maracle, E. Balogh, Z. Szekanecz, Targeting of proangiogenic signalling pathways in chronic inflammation, Nat. Rev. Rheumatol. 12 (2015) 111–122, http://dx.doi.org/10.1038/nrrheum.2015.164.
- [6] US Food and Drug administration (FDA), CDER 2001 Report to the Nation, 2001.
- [7] US Food and Drug administration (FDA), FDA Prescribing Information Gleevec, 2001.
- [8] P. Wu, T.E. Nielsen, M.H. Clausen, Small-molecule kinase inhibitors: an analysis of FDA-approved drugs, Drug Discov. Today 21 (2016) 5–10, http://dx.doi.org/10. 1016/j.drudis.2015.07.008.
- [9] European Medicines Agency (EMA), Assessment Report Jakavi (EPAR), 2012.
- [10] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Jakafi, 2011.
- [11] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Xeljanz, 2012.
- [12] European Medicines Agency (EMA), Assessment report Xeljanz (EPAR), 2013.
- [13] A. Fountas, L-N. Diamantopoulos, A. Tsatsoulis, Tyrosine kinase inhibitors and diabetes: a novel treatment paradigm? Trends Endocrinol. Metab. 26 (2015) 643–656, http://dx.doi.org/10.1016/j.tem.2015.09.003.
- [14] R.A. Jibodh, J.S. Lagas, B. Nuijen, J.H. Beijnen, J.H.M. Schellens, Taxanes: old drugs, new oral formulations, Eur. J. Pharmacol. 717 (2013) 40–46, http://dx.doi.org/10. 1016/j.ejphar.2013.02.058.
- [15] L. Benjamin, F.-E. Cotté, C. Philippe, F. Mercier, T. Bachelot, G. Vidal-Trécan, Physicians' preferences for prescribing oral and intravenous anticancer drugs: a discrete choice experiment, Eur. J. Cancer 48 (2012) 912–920, http://dx.doi.org/10.1016/j. ejca.2011.09.019.
- [16] G. Liu, E. Franssen, M.I. Fitch, E. Warner, Patient preferences for oral versus intravenous palliative chemotherapy, J. Clin. Oncol. 15 (1997) 110–115 http://www.ncbi. nlm.nih.gov/pubmed/8996131.
- [17] N. Eckstein, L. Röper, B. Haas, H. Potthast, U. Hermes, C. Unkrig, F. Naumann-Winter, H. Enzmann, Clinical pharmacology of tyrosine kinase inhibitors becoming generic drugs: the regulatory perspective, J. Exp. Clin. Cancer Res. 33 (2014) 15, http://dx.doi.org/10.1186/1756-9966-33-15.
- [18] W.P. Janzen, Screening technologies for small molecule discovery: the state of the art, Chem. Biol. 21 (2014) 1162–1170, http://dx.doi.org/10.1016/j.chembiol.2014.07.015.
- [19] K. Nikolic, L. Mavridis, T. Djikic, J. Vucicevic, D. Agbaba, K. Yelekci, J.B.O. Mitchell, Drug design for CNS diseases: polypharmacological profiling of compounds using cheminformatic, 3D-QSAR and virtual screening methodologies, Front. Neurosci. 10 (2016), http://dx.doi.org/10.3389/fnins.2016.00265.
- [20] S. Stegemann, F. Leveiller, D. Franchi, H. de Jong, H. Lindén, When poor solubility becomes an issue: from early stage to proof of concept, Eur. J. Pharm. Sci. 31 (2007) 249–261, http://dx.doi.org/10.1016/j.ejps.2007.05.110.
- [21] Y. Kawabata, K. Wada, M. Nakatani, S. Yamada, S. Onoue, Formulation design for poorly water-soluble drugs based on biopharmaceutics classification system: basic approaches and practical applications, Int. J. Pharm. 420 (2011) 1–10, http://dx.doi.org/10.1016/j.ijpharm.2011.08.032.
- [22] A. Dahan, A. Beig, D. Lindley, J.M. Miller, The solubility-permeability interplay and oral drug formulation design: two heads are better than one, Adv. Drug Deliv. Rev. 101 (2016) 99–107, http://dx.doi.org/10.1016/j.addr.2016.04.018.
- [23] European Medicines Agency (EMA), Assessment Report Glivec (EPAR), 2004.
- [24] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Gleevec, 2001.
- [25] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Stivarga, 2012.
- [26] European Medicines Agency (EMA), Assessment Report Stivarga (EPAR), 2014.
- [27] European Medicines Agency (EMA), Assessment Report Iressa (EPAR), 2009.
- [28] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Iressa, 2003.

- [29] European Medicines Agency (EMA), Assessment Report Tarceva (EPAR), 2005.
- [30] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Tarceva, 2004.
- [31] European Medicines Agency (EMA), Assessment Report Bosulif (EPAR), 2014.
- [32] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Bosulif, 2012.
- [33] European Medicines Agency (EMA), Assessment Report Nexavar (EPAR), 2006.
- [34] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Nexavar, 2005.
- [35] European Medicines Agency (EMA), Assessment Report Inlyta (EPAR), 2012.
 [36] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Inlyta, 2012.
- [37] European Medicines Agency (EMA), Assessment Report Sprycel (EPAR), 2006.
- [38] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Sprycel, 2006.
- [39] European Medicines Agency (EMA), Assessment Report Imbruvica (EPAR), 2014.
 [40] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Imbruvica, 2013.
- [41] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Sutent, 2006.
- [42] European Medicines Agency (EMA), Assessment report Giotrif (EPAR), 2013.
- [43] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Giotrif, 2013.
- [44] European Medicines Agency (EMA), Assessment Report Tasigna (EPAR), 2009.
- [45] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Tasigna, 2007.
- [46] European Medicines Agency (EMA), Assessment Report Tafinlar (EPAR), 2013.
 [47] US Food and Drug administration (FDA), Clinical Pharmacology at
- [47] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Tafinlar, 2013.
- [48] European Medicines Agency (EMA), Assessment Report Tyverb (EPAR), 2009.
- [49] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Tykerb, 2007.
- [50] European Medicines Agency (EMA), Assessment Report Mekinist (EPAR), 2014.
 [51] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Mekinist, 2013.
- [52] European Medicines Agency (EMA), Assessment Report Votrient (EPAR), 2011.
- [53] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Votrient, 2009.
- [54] European Medicines Agency (EMA), Assessment report Vargatef (EPAR), 2015.
 [55] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Vargatef, 2014.
- [56] European Medicines Agency (EMA), Assessment Report Zykadia (EPAR), 2015.
- [57] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Zykadia, 2014.
- [58] European Medicines Agency (EMA), Assessment Report Xalkori (EPAR), 2013.
- [59] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Xalkori, 2011.
- [60] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Alecensa, 2015.
- [61] European Medicines Agency (EMA), Assessment Report Zelboraf (EPAR), 2011.[62] US Food and Drug administration (FDA), Clinical Pharmacology and
- Biopharmaceutics Review Zelboraf, 2011.
- [63] European Medicines Agency (EMA), Assessment Report Cotellic (EPAR), 2015.
- [64] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Cotellic, 2015.
- [65] European Medicines Agency (EMA), Assessment Report Caprelsa (EPAR), 2013.
- [66] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Caprelsa, 2011.
- [67] European Medicines Agency (EMA), Assessment Report Tagrisso (EPAR), 2016.
- [68] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Tagrisso, 2015.
- [69] European Medicines Agency (EMA), Assessment Report Iclusig (EPAR), 2013.
 [70] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Iclusig, 2012.
- [71] European Medicines Agency (EMA), Assessment Report Lenvima (EPAR), 2015.
- [72] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Lenvima, 2015.
- [73] European Medicines Agency (EMA), Assessment Report Cometriq (EPAR), 2014.
 [74] US Food and Drug administration (FDA), Clinical Pharmacology and Biopharmaceutics Review Cometriq, 2012.
- [75] M. Rowland, T.N. Tozer, Clinical Pharmacokinetics; Concepts and Applications, 3rd ed Lippincott, Williams & Wilkins, Philadelphia, 1994.
- [76] L.Z. Benet, The role of BCS (biopharmaceutics classification system) and BDDCS (biopharmaceutics drug disposition classification system) in drug development, J. Pharm. Sci. 102 (2013) 34–42, http://dx.doi.org/10.1002/jps.23359.
- [77] M. Herbrink, B. Nuijen, J.H.M. Schellens, J.H. Beijnen, Variability in bioavailability of small molecular tyrosine kinase inhibitors, Cancer Treat. Rev. 41 (2015) 412–422, http://dx.doi.org/10.1016/j.ctrv.2015.03.005.
- [78] M. Estudante, J.G. Morais, G. Soveral, L.Z. Benet, Intestinal drug transporters: an overview, Adv. Drug Deliv. Rev. 65 (2013) 1340–1356, http://dx.doi.org/10.1016/ j.addr.2012.09.042.
- [79] I. Pereira de Sousa, A. Bernkop-Schnürch, Pre-systemic metabolism of orally administered drugs and strategies to overcome it, J. Control. Release 192 (2014) 301–309, http://dx.doi.org/10.1016/j.jconrel.2014.08.004.
- [80] C.J. Tynan, V. Lo Schiavo, L. Zanetti-Domingues, S.R. Needham, S.K. Roberts, M. Hirsch, D.J. Rolfe, D. Korovesis, D.T. Clarke, M.L. Martin-Fernandez, A tale of the

epidermal growth factor receptor: the quest for structural resolution on cells, Methods 95 (2016) 86–93, http://dx.doi.org/10.1016/j.ymeth.2015.10.009.

- [81] P. Khanna, P.J. Chua, B.H. Bay, G.H. Baeg, The JAK/STAT signaling cascade in gastric carcinoma (review), Int. J. Oncol. 47 (2015) 1617–1626, http://dx.doi.org/10.3892/ iio.2015.3160.
- [82] M. Pinne, J.L. Raucy, Advantages of cell-based high-volume screening assays to assess nuclear receptor activation during drug discovery, Expert Opin. Drug Discovery 9 (2014) 669–686, http://dx.doi.org/10.1517/17460441.2014.913019.
- [83] P. Kohonen, R. Ceder, I. Smit, V. Hongisto, G. Myatt, B. Hardy, O. Spjuth, R. Grafström, Cancer biology, toxicology and alternative methods development go hand-in-hand, Basic Clin. Pharmacol. Toxicol. 115 (2014) 50–58, http://dx.doi. org/10.1111/bcot.12257.
- [84] R. Roskoski, Classification of small molecule protein kinase inhibitors based upon the structures of their drug-enzyme complexes, Pharmacol. Res. 103 (2016) 26–48, http://dx.doi.org/10.1016/j.phrs.2015.10.021.
- [85] K. Okamoto, M. Ikemori-Kawada, A. Jestel, K. von König, Y. Funahashi, T. Matsushima, A. Tsuruoka, A. Inoue, J. Matsui, Distinct binding mode of multikinase inhibitor lenvatinib revealed by biochemical characterization, ACS Med. Chem. Lett. 6 (2015) 89–94, http://dx.doi.org/10.1021/ml500394m.
- [86] L. Hu, Y. Zheng, Z. Li, Y. Wang, Y. Lv, X. Qin, C. Zeng, Design, synthesis, and biological activity of phenyl-pyrazole derivatives as BCR-ABL kinase inhibitors, Bioorg. Med. Chem. 23 (2015) 3147–3152, http://dx.doi.org/10.1016/i.bmc.2015.04.083.
- [87] N.M. Levinson, S.G. Boxer, Structural and spectroscopic analysis of the kinase inhibitor bosutinib and an isomer of bosutinib binding to the Abl tyrosine kinase domain, PLoS One 7 (2012), e29828, http://dx.doi.org/10.1371/journal.pone. 0029828.
- [88] J.S. Tokarski, The structure of dasatinib (BMS-354825) bound to activated ABL kinase domain elucidates its inhibitory activity against Imatinib-resistant ABL mutants, Cancer Res. 66 (2006) 5790–5797, http://dx.doi.org/10.1158/0008-5472. CAN-05-4187.
- [89] A. Tse, G.M. Verkhivker, Molecular determinants underlying binding specificities of the ABL kinase inhibitors: combining alanine scanning of binding hot spots with network analysis of residue interactions and coevolution, PLoS One 10 (2015), e0130203, http://dx.doi.org/10.1371/journal.pone.0130203.
- [90] R. Roskoski, ErbB/HER protein-tyrosine kinases: structures and small molecule inhibitors, Pharmacol. Res. 87 (2014) 42–59, http://dx.doi.org/10.1016/j.phrs.2014. 06.001.
- [91] Y. Yosaatmadja, S. Silva, J.M. Dickson, A.V. Patterson, J.B. Smaill, J.U. Flanagan, M.J. McKeage, C.J. Squire, Binding mode of the breakthrough inhibitor AZD9291 to epidermal growth factor receptor revealed, J. Struct. Biol. 192 (2015) 539–544, http:// dx.doi.org/10.1016/j.jsb.2015.10.018.
- [92] S. Ravez, O. Castillo-Aguilera, P. Depreux, L. Goossens, Quinazoline derivatives as anticancer drugs: a patent review (2011 – present), Expert Opin. Ther. Pat. 25 (2015) 789–804, http://dx.doi.org/10.1517/13543776.2015.1039512.
- [93] Z. Zhan, J. Ai, Q. Liu, Y. Ji, T. Chen, Y. Xu, M. Geng, W. Duan, Discovery of anilinopyrimidines as dual inhibitors of c-met and VEGFR-2: synthesis, SAR, and cellular activity, ACS Med. Chem. Lett. 5 (2014) 673–678, http://dx.doi.org/10. 1021/ml500066m.
- [94] G.J. Roth, R. Binder, F. Colbatzky, C. Dallinger, R. Schlenker-Herceg, F. Hilberg, S.-L. Wollin, R. Kaiser, Nintedanib: from discovery to the clinic, J. Med. Chem. 58 (2015) 1053–1063, http://dx.doi.org/10.1021/jm501562a.
- [95] J. Li, N. Zhou, K. Luo, W. Zhang, X. Li, C. Wu, J. Bao, In silico discovery of potential VEGFR-2 inhibitors from natural derivatives for anti-angiogenesis therapy, Int. J. Mol. Sci. 15 (2014) 15994–16011, http://dx.doi.org/10.3390/ijms150915994.
- [96] W.M. Eldehna, M. Fares, H.S. Ibrahim, M.H. Aly, S. Zada, M.M. Ali, S.M. Abou-Seri, H.A. Abdel-Aziz, D.A. Abou El Ella, Indoline ureas as potential anti-hepatocellular carcinoma agents targeting VEGFR-2: synthesis, in vitro biological evaluation and molecular docking, Eur. J. Med. Chem. 100 (2015) 89–97, http://dx.doi.org/10. 1016/j.ejmech.2015.05.040.
- [97] Y. Shan, C. Wang, L. Zhang, J. Wang, M. Wang, Y. Dong, Expanding the structural diversity of diarylureas as multi-target tyrosine kinase inhibitors, Bioorg. Med. Chem. 24 (2016) 750–758, http://dx.doi.org/10.1016/j.bmc.2015.12.038.
- [98] F. Meng, Molecular dynamics simulation of VEGFR2 with sorafenib and other ureasubstituted Aryloxy compounds, J. Theor. Chem. 2013 (2013) 1–7, http://dx.doi. org/10.1155/2013/739574.
- [99] T. Pemovska, E. Johnson, M. Kontro, G.A. Repasky, J. Chen, P. Wells, C.N. Cronin, M. McTigue, O. Kallioniemi, K. Porkka, B.W. Murray, K. Wennerberg, Axitinib effectively inhibits BCR-ABL1(T3151) with a distinct binding conformation, Nature 519 (2015) 102–105, http://dx.doi.org/10.1038/nature14119.
- [100] M. McTigue, B.W. Murray, J.H. Chen, Y.-L. Deng, J. Solowiej, R.S. Kania, Molecular conformations, interactions, and properties associated with drug efficiency and clinical performance among VEGFR TK inhibitors, Proc. Natl. Acad. Sci. 109 (2012) 18281–18289, http://dx.doi.org/10.1073/pnas.1207759109.
- [101] Y. Jia, J. Zhang, J. Feng, F. Xu, H. Pan, W. Xu, Design, synthesis and biological evaluation of pazopanib derivatives as antitumor agents, Chem. Biol. Drug Des. 83 (2014) 306-316, http://dx.doi.org/10.1111/cbdd.12243.
- [102] J. Zhang, X. Jiang, Y. Jiang, M. Guo, S. Zhang, J. Li, J. He, J. Liu, J. Wang, L. Ouyang, Recent advances in the development of dual VEGFR and c-met small molecule inhibitors as anticancer drugs, Eur. J. Med. Chem. 108 (2016) 495–504, http://dx.doi. org/10.1016/j.ejmech.2015.12.016.
- [103] M. Pellowka, D. Merk, Advances in personal medicine medicinal chemistry and pharmacology of vemurafenib and ivacaftor, Pharmazie 68 (2013) 484–491, http://dx.doi.org/10.1691/ph.2013.6526.
- [104] L. Ren, K.A. Ahrendt, J. Grina, E.R. Laird, A.J. Buckmelter, J.D. Hansen, B. Newhouse, D. Moreno, S. Wenglowsky, V. Dinkel, S.L. Gloor, G. Hastings, S. Rana, K. Rasor, T. Risom, H.L. Sturgis, W.C. Voegtli, S. Mathieu, The discovery of potent and selective

pyridopyrimidin-7-one based inhibitors of B-RafV600E kinase, Bioorg. Med. Chem. Lett. 22 (2012) 3387-3391, http://dx.doi.org/10.1016/j.bmcl.2012.04.015.

- [105] T. Zhou, S. Georgeon, R. Moser, D.J. Moore, A. Caflisch, O. Hantschel, Specificity and mechanism-of-action of the JAK2 tyrosine kinase inhibitors ruxolitinib and SAR302503 (TG101348), Leukemia 28 (2014) 404–407, http://dx.doi.org/10. 1038/leu.2013.205.
- [106] M.K. Kim, O. Bae, Y. Chong, Design, synthesis, and molecular docking study of Flavonol derivatives as selective JAK1 inhibitors, Bull. Kor. Chem. Soc. 35 (2014) 2581–2584, http://dx.doi.org/10.5012/bkcs.2014.35.8.2581.
- [107] Y. Duan, L. Chen, Y. Chen, X. Fan, c-Src Binds to the Cancer Drug Ruxolitinib with an Active Conformation, PLoS One 9 (2014), e106225, http://dx.doi.org/10.1371/ journal.pone.0106225.
- [108] N.K. Williams, R.S. Bamert, O. Patel, C. Wang, P.M. Walden, A.F. Wilks, E. Fantino, J. Rossjohn, I.S. Lucet, Dissecting specificity in the Janus kinases: the structures of JAK-specific inhibitors complexed to the JAK1 and JAK2 protein tyrosine kinase domains, J. Mol. Biol. 387 (2009) 219–232, http://dx.doi.org/10.1016/j.jmb.2009.01. 041.
- [109] A. Kumar, V. Shanthi, K. Ramanathan, Computational investigation and experimental validation of Crizotinib resistance conferred by C1156Y mutant anaplastic lymphoma kinase, Mol. Inform. 34 (2015) 105–114, http://dx.doi.org/10.1002/minf. 201400070.
- [110] J.J. Cui, M. Tran-Dubé, H. Shen, M. Nambu, P.-P. Kung, M. Pairish, L. Jia, J. Meng, L. Funk, I. Botrous, M. McTigue, N. Grodsky, K. Ryan, E. Padrique, G. Alton, S. Timofeevski, S. Yamazaki, Q. Li, H. Zou, J. Christensen, B. Mroczkowski, S. Bender, R.S. Kania, M.P. Edwards, Structure based drug design of crizotinib (PF-02341066), a potent and selective dual inhibitor of mesenchymal–epithelial transition factor (c-MET) kinase and anaplastic lymphoma kinase (ALK), J. Med. Chem. 54 (2011) 6342–6363, http://dx.doi.org/10.1021/jm2007613.
- [111] Z. Ni, T.-C. Zhang, Computationally unraveling how ceritinib overcomes drug-resistance mutations in ALK-rearranged lung cancer, J. Mol. Model. 21 (2015) 175, http://dx.doi.org/10.1007/s00894-015-2716-z.
- [112] X. Jiang, J. Zhou, J. Ai, Z. Song, X. Peng, L. Xing, Y. Xi, J. Guo, Q. Yao, J. Ding, M. Geng, A. Zhang, Novel tetracyclic benzo[b]carbazolones as highly potent and orally bioavailable ALK inhibitors: design, synthesis, and structure—activity relationship study, Eur. J. Med. Chem. 105 (2015) 39–56, http://dx.doi.org/10.1016/j.ejmech. 2015.10.005.
- [113] Z. Song, M. Wang, A. Zhang, Alectinib: a novel second generation anaplastic lymphoma kinase (ALK) inhibitor for overcoming clinically-acquired resistance, Acta Pharm. Sin. B 5 (2015) 34–37, http://dx.doi.org/10.1016/j.apsb.2014.12.007.
- [114] H. Yari, M.R. Ganjalikhany, H. Sadegh, In silico investigation of new binding pocket for mitogen activated kinase kinase (MEK): development of new promising inhibitors, Comput. Biol. Chem. 59 (2015) 185–198, http://dx.doi.org/10.1016/j. compbiolchem.2015.09.013.
- [115] I.V. Hartung, S. Hammer, M. Hitchcock, R. Neuhaus, A. Scholz, G. Siemeister, R. Bohlmann, R.C. Hillig, F. Pühler, Optimization of allosteric MEK inhibitors. Part 2: taming the sulfamide group balances compound distribution properties, Bioorg. Med. Chem. Lett. 26 (2016) 186–193, http://dx.doi.org/10.1016/j.bmcl.2015.11. 004.
- [116] K.D. Robarge, W. Lee, C. Eigenbrot, M. Ultsch, C. Wiesmann, R. Heald, S. Price, J. Hewitt, P. Jackson, P. Savy, B. Burton, E.F. Choo, J. Pang, J. Boggs, A. Yang, X. Yang, M. Baumgardner, Structure based design of novel 6,5 heterobicyclic mitogen-activated protein kinase kinase (MEK) inhibitors leading to the discovery of imidazo[1,5-a] pyrazine G-479, Bioorg. Med. Chem. Lett. 24 (2014) 4714–4723, http://dx.doi.org/10.1016/j.bmcl.2014.08.008.
- [117] T. Yogo, H. Nagamiya, M. Seto, S. Sasaki, H. Shih-Chung, Y. Ohba, N. Tokunaga, G.N. Lee, C.Y. Rhim, C.H. Yoon, S.Y. Cho, R. Skene, S. Yamamoto, Y. Satou, M. Kuno, T. Miyazaki, H. Nakagawa, A. Okabe, S. Marui, K. Aso, M. Yoshida, Structure-based design and synthesis of 3-amino-1,5-dihydro-4H-pyrazolopyridin-4-one derivatives as tyrosine kinase 2 inhibitors, J. Med. Chem. 59 (2016) 733–749, http://dx.doi. org/10.1021/acs.jmedchem.5b01857.
- [118] V. Simov, S.V. Deshmukh, C.J. Dinsmore, F. Elwood, R.B. Fernandez, Y. Garcia, C. Gibeau, H. Gunaydin, J. Jung, J.D. Katz, B. Kraybill, B. Lapointe, S.B. Patel, T. Siu, H. Su, J.R. Young, Structure-based design and development of (benz)imidazole pyridones as JAK1-selective kinase inhibitors, Bioorg. Med. Chem. Lett. 26 (2016) 1803–1808, http://dx.doi.org/10.1016/j.bmcl.2016.02.035.
- [119] M.A. Aziz, R.A.T. Serya, D.S. Lasheen, A.K. Abdel-Aziz, A. Esmat, A.M. Mansour, A.N.B. Singab, K.A.M. Abouzid, Discovery of potent VEGFR-2 inhibitors based on furopyrimidine and thienopyrimidne scaffolds as cancer targeting agents, Sci. Report. 6 (2016) 24460, http://dx.doi.org/10.1038/srep24460.
- [120] J. Han, S.J. Kaspersen, S. Nervik, K.G. Nørsett, E. Sundby, B.H. Hoff, Chiral 6-arylfuro[2,3-d]pyrimidin-4-amines as EGFR inhibitors, Eur. J. Med. Chem. 119 (2016) 278–299, http://dx.doi.org/10.1016/j.ejmech.2016.04.054.
- [121] G. Thomas, Medicinal Chemistry: An Introduction, 2nd ed John Wiley and sons, Chichester, 2007.
- [122] W. Xing, J. Ai, S. Jin, Z. Shi, X. Peng, L. Wang, Y. Ji, D. Lu, Y. Liu, M. Geng, Y. Hu, Enhancing the cellular anti-proliferation activity of pyridazinones as c-met inhibitors using docking analysis, Eur. J. Med. Chem. 95 (2015) 302–312, http://dx.doi.org/10. 1016/j.ejmech.2015.03.041.
- [123] X. Wu, S. Wan, G. Wang, H. Jin, Z. Li, Y. Tian, Z. Zhu, J. Zhang, Molecular dynamics simulation and free energy calculation studies of kinase inhibitors binding to active and inactive conformations of VEGFR-2, J. Mol. Graph. Model. 56 (2015) 103–112, http://dx.doi.org/10.1016/j.jmgm.2014.12.006.
- [124] E. Weisberg, H.G. Choi, A. Ray, R. Barrett, J. Zhang, T. Sim, W. Zhou, M. Seeliger, M. Cameron, M. Azam, J.A. Fletcher, M. Debiec-Rychter, M. Mayeda, D. Moreno, A.L. Kung, P.A. Janne, R. Khosravi-Far, J.V. Melo, P.W. Manley, S. Adamia, C. Wu, N. Gray, J.D. Griffin, Discovery of a small-molecule type II inhibitor of wild-type and

gatekeeper mutants of BCR-ABL, PDGFRalpha, Kit, and Src kinases: novel type II inhibitor of gatekeeper mutants, Blood 115 (2010) 4206–4216, http://dx.doi.org/10. 1182/blood-2009-11-251751.

- [125] E.P. Reddy, A.K. Aggarwal, The ins and outs of Bcr-Abl inhibition, Genes Cancer. 3 (2012) 447–454, http://dx.doi.org/10.1177/1947601912462126.
- [126] C. Fraser, J.C. Dawson, R. Dowling, D.R. Houston, J.T. Weiss, A.F. Munro, M. Muir, L. Harrington, S.P. Webster, M.C. Frame, V.G. Brunton, E.E. Patton, N.O. Carragher, A. Unciti-Broceta, Rapid discovery and structure-activity relationships of pyrazolopyrimidines that potently suppress breast cancer cell growth via SRC kinase inhibition with exceptional selectivity over ABL kinase, J. Med. Chem. 59 (2016) 4697–4710, http://dx.doi.org/10.1021/acs.jmedchem.6b00065.
- [127] J. Rautio, H. Kumpulainen, T. Heimbach, R. Oliyai, D. Oh, T. Järvinen, J. Savolainen, Prodrugs: design and clinical applications, Nat. Rev. Drug Discov. 7 (2008) 255–270, http://dx.doi.org/10.1038/nrd2468.
- [128] J.D. Oslob, S.A. Heumann, C.H. Yu, D.A. Allen, S. Baskaran, M. Bui, E. Delarosa, A.D. Fung, A. Hashash, J. Hau, S. Ivy, J.W. Jacobs, W. Lew, J. Maung, R.S. McDowell, S. Ritchie, M.J. Romanowski, J.A. Silverman, W. Yang, M. Zhong, T. Fuchs-Knotts, Water-soluble prodrugs of an aurora kinase inhibitor, Bioorg. Med. Chem. Lett. 19 (2009) 1409–1412, http://dx.doi.org/10.1016/j.bmcl.2009.01.043
- [129] P.S. Charifson, W.P. Walters, Acidic and basic drugs in medicinal chemistry: a perspective, J. Med. Chem. 57 (2014) 9701–9717, http://dx.doi.org/10.1021/ jm501000a.
- [130] G. Thomas, Medicinal Chemistry: An Introduction, Second Edition, 2nd ed. Wiley, West Sussex, 2007 49–51, http://dx.doi.org/10.1021/ed086p1036.
- [131] B.J. Krieg, S.M. Taghavi, G.L. Amidon, G.E. Amidon, In vivo predictive dissolution: comparing the effect of bicarbonate and phosphate buffer on the dissolution of weak acids and weak bases, J. Pharm. Sci. 104 (2015) 2894–2904, http://dx.doi. org/10.1002/jps.24460.
- [132] L. Zhang, F. Wu, S.C. Lee, H. Zhao, pH-dependent drug-drug interactions for weak base drugs: potential implications for new drug development, Clin. Pharmacol. Ther. 96 (2014) 266–277, http://dx.doi.org/10.1038/clpt.2014.87.
- [133] N.R. Budha, A. Frymoyer, G.S. Smelick, J.Y. Jin, M.R. Yago, M.J. Dresser, S.N. Holden, L.Z. Benet, J.A. Ware, Drug absorption interactions between oral targeted anticancer agents and PPIs: is pH-dependent solubility the Achilles heel of targeted therapy? Clin. Pharmacol. Ther. 92 (2012) 203–213, http://dx.doi.org/10.1038/clpt. 2012.73.
- [134] G.A. Stephenson, A. Aburub, T.A. Woods, Physical stability of salts of weak bases in the solid-state, J. Pharm. Sci. 100 (2011) 1607–1617, http://dx.doi.org/10.1002/jps. 22405.
- [135] R. Censi, P. Di Martino, Polymorph impact on the bioavailability and stability of poorly soluble drugs, Molecules 20 (2015) 18759–18776, http://dx.doi.org/10. 3390/molecules201018759.
- [136] S. Roy, R. Quiñones, A.J. Matzger, Structural and physicochemical aspects of dasatinib hydrate and anhydrate phases, Cryst. Growth Des. 12 (2012) 2122–2126, http://dx.doi.org/10.1021/cg300152p.
- [137] A. Martin, Non ideal solutions, Phys. Pharm. 4th ed. Lippincott, Williams & Wilkins, Baltimore 1993, pp. 223–225.
- [138] R. Pinal, Effect of molecular symmetry on melting temperature and solubility, Org. Biomol. Chem. 2 (2004) 2692–2699, http://dx.doi.org/10.1039/B407105K.
- [139] C.M. Hansen, 50 Years with solubility parameters—past and future, Prog. Org. Coat. 51 (2004) 77–84, http://dx.doi.org/10.1016/j.porgcoat.2004.05.004.
- [140] I.B. Mekjavic, Contribution of thermal and nonthermal factors to the regulation of body temperature in humans, J. Appl. Physiol. 100 (2006) 2065–2072, http://dx. doi.org/10.1152/japplphysiol.01118.2005.
- [141] G. Kelly, Body temperature variability (part 1): a review of the history of body temperature and its variability due to site selection, biological rhythms, fitness, and aging, Altern. Med. Rev. 11 (2006) 278–293 http://www.ncbi.nlm.nih.gov/ pubmed/17176167.
- [142] F.S. Mortimer, Melting point, latent heat of fusion and solubility, J. Am. Chem. Soc. 44 (1922) 1416–1429, http://dx.doi.org/10.1021/ja01428a002.
- [143] E. Thomas, J. Rubino, Solubility, melting point and salting-out relationships in a group of secondary amine hydrochloride salts, Int. J. Pharm. 130 (1996) 179–185, http://dx.doi.org/10.1016/0378-5173(95)04269-5.
- [144] B. Hancock, The use of solubility parameters in pharmaceutical dosage form design, Int. J. Pharm. 148 (1997) 1–21, http://dx.doi.org/10.1016/S0378-5173(96)04828-4.
- [145] C.M. Hansen, The three Dimensional Solubility Parameter and Solvent Diffusion CoefficientDanmarks Tekniske Hojskole 1967.
- [146] H.P. Pharmaceutical, Solvate Form M of Trametinib Dimethyl Sulfoxide and Methods of Making and Using thereof, US9181243B2, 2013.
- [147] Australian Therapeutic Goods Administration, Australian Public Asessment Report (AusPAR): Vandetanib, 2013.
- [148] T.A. Albahri, Accurate prediction of the solubility parameter of pure compounds from their molecular structures, Fluid Phase Equilib. 379 (2014) 96–103, http:// dx.doi.org/10.1016/j.fluid.2014.07.016.
- [149] F. Gharagheizi, A. Eslamimanesh, F. Farjood, A.H. Mohammadi, D. Richon, Solubility parameters of nonelectrolyte organic compounds: determination using quantitative structure-property relationship strategy, Ind. Eng. Chem. Res. 50 (2011) 11382–11395, http://dx.doi.org/10.1021/ie200962w.
- [150] C.M. Hansen, Hansen Solubility Parameters: A User's Handbook, 1st ed CRC Press, Boca Raton, Florida, 1999.
- [151] J. Goole, D.J. Lindley, W. Roth, S.M. Carl, K. Amighi, J.-M. Kauffmann, G.T. Knipp, The effects of excipients on transporter mediated absorption, Int. J. Pharm. 393 (2010) 17–31, http://dx.doi.org/10.1016/j.ijpharm.2010.04.019.
- [152] European Medicines Agency (EMA), European Public Assessment Report (EPAR), 2001–2016.

- [153] US Food and Drug administration (FDA), Dissolution methods, August 30, 2016 www.accessdata.fda.gov/scripts/cder/dissolution/, accessed March 1, 2016.
- E. R&D, Pharmaceutical composition of Lenvatinib, US8969379, 2005. [154] [155] US Food and Drug administration (FDA) Chemistry Review Tarceva 2004
- [156] Australian Therapeutic Goods Administration, Australian Public Assessment Report
- (AUSPAR) Dasatinib, 2011. [157]
- US Food and Drug administration (FDA), Chemistry Review Tasigna, 2007. [158] US Food and Drug administration (FDA), Chemistry Review Tykerb, 2007.
- [159] Australian Therapeutic Goods Administration, Australian Public Assessment Report (AUSPAR) Lapatinib, 2012.
- Australian Therapeutic Goods Administration, Australian Public Assessment Report [160] (AUSPAR) Pazopanib, 2013.
- Australian Therapeutic Goods Administration, Australian Public Assessment Report [161] (AUSPAR) Ruxolitinib, 2014.
- US Food and Drug administration (FDA), Prescribing information Xalkori, 2011. [162]
- [163] L. Nguyen, J. Holland, R. Mamelok, M. Laberge, J. Grenier, D. Swearingen, D. Armas, S. Lacy, Evaluation of the effect of food and gastric pH on the single-dose pharmacokinetics of cabozantinib in healthy adult subjects, J. Clin. Pharmacol. 55 (2015) 1293-1302, http://dx.doi.org/10.1002/jcph.526.
- [164] Australian Therapeutic Goods Administration, Australian Public Assessment Report (AUSPAR) Tofacitinib. 2015.
- [165] Australian Therapeutic Goods Administration, Australian Public Assessment Report (AUSPAR) Afatinib, 2014.
- [166] Australian Therapeutic Goods Administration, Product Information Afatinib, 2014. [167] Australian Therapeutic Goods Administration, Australian Public Assessment Report
- (ALISPAR) Trametinib 2014
- US Food and Drug administration (FDA), Prescribing information Zykadia, 2014. [168]
- [169] US Food and Drug administration (FDA), Prescribing information Alecensa, 2014.
- [170] US Food and Drug administration (FDA), Chemistry Review Cotellic, 2015.
- [171] US Food and Drug administration (FDA), Chemistry Review Tagrisso, 2015.
- [172] Boehringer-Ingelheim, Product monograph Ofev, 2015.
- S. Gaisford, M. Saunders, Essentials of Pharmaceutical Preformulation, 1st ed [173] Wiley-Blackwell, Chichester, 2013.
- [174] D.K. Mishra, V. Dhote, A. Bhargava, D.K. Jain, P.K. Mishra, Amorphous solid dispersion technique for improved drug delivery: basics to clinical applications, Drug Deliv. Transl. Res. 5 (2015) 552-565, http://dx.doi.org/10.1007/s13346-015-0256-9.
- [175] K.T. Savjani, A.K. Gajjar, J.K. Savjani, Drug solubility: importance and enhancement techniques, ISRN Pharm. 2012 (2012) 1-10, http://dx.doi.org/10.5402/2012/ 195727
- [176] W.L. Chiou, S. Riegelman, Pharmaceutical applications of solid dispersion systems, J. Pharm. Sci. 60 (1971) 1281-1302 http://www.ncbi.nlm.nih.gov/pubmed/ 4935981
- [177] N. Shah, R.M. Iyer, H.-J. Mair, D. Choi, H. Tian, R. Diodone, K. Fahnrich, A. Pabst-Ravot, K. Tang, E. Scheubel, J.F. Grippo, S.A. Moreira, Z. Go, J. Mouskountakis, T. Louie, P.N. Ibrahim, H. Sandhu, L. Rubia, H. Chokshi, D. Singhal, W. Malick, Improved human bioavailability of vemurafenib, a practically insoluble drug, using an amorphous polymer-stabilized solid dispersion prepared by a solvent-controlled coprecipitation process, J. Pharm. Sci. 102 (2013) 967-981, http://dx.doi. org/10.1002/ips.23425
- [178] L. Roche, Propane-i-sulfonic Acid {3-[5-(4-chloro-phenyl)-1 h-pyrrolo [2,3-b] pyridine-3-carbonyl]-2, 4-difluoro-pheny l}-amide Compositions and Uses Thereof, WO2010114928, 2010.
- [179] Bayer, Coated Pharmaceutical Composition Containing Regorafenib, US 20140065212, 2013.
- [180] Bayer, New Pharmaceutical Compositions Comprising 4-(4-(3-(4-chloro-3trifluoromethyl-phenyl)-ureido)-3-fluoro-phenoxy)-pyridine-2-carboxylic Acid for the Treatment of Hyper-proliferative Disorders, WO 2006026500 A1, 2005.
- [181] B. Ingelheim, Pharmaceutical Combination, US20110178099A1, 2009.
- [182] A. Larsen, Method for Treating Colorectal Cancer, WO2010081817A1, 2010.

- [183] B. Ingelheim, Capsule Pharmaceutical Dosage from Comprising a Suspension Formulation of an Indoline Derivative, wo2009147212A1, 2008.
- [184] US Food and Drug administration (FDA), FDA Orange Book Imatinib, 2016.
- [185] European Medicines Agency (EMA), European Public Assessment Report Imatinib, 2016
- [186] Xspray Microparticles AB, A Method for Producing Stable, Amorphous Nanoparticles Comprising at Least one Protein Kinase Inhibitor and at Least one Polymeric Stabilizing and Matrix Forming Component, WO 2013105894 A1, 2012.
- [187] C. Godugu, R. Doddapaneni, A.R. Patel, R. Singh, R. Mercer, M. Singh, Novel gefitinib formulation with improved oral bioavailability in treatment of A431 skin carcinoma, Pharm. Res. 33 (2016) 137-154, http://dx.doi.org/10.1007/s11095-015-1771-
- [188] D.H. Truong, T.H. Tran, T. Ramasamy, J.Y. Choi, H.-G. Choi, C.S. Yong, J.O. Kim, Preparation and characterization of solid dispersion using a novel amphiphilic copolymer to enhance dissolution and oral bioavailability of sorafenib, Powder Technol. 283 (2015) 260-265, http://dx.doi.org/10.1016/j.powtec.2015.04.044.
- [189] N. Ab, Super-saturated Delivery Vehicles for poorly Water-Soluble Pharmaceutical and Cosmetic Active Ingredients, EP 2616050 A1, 2011.
- [190] Y. Song, X. Yang, X. Chen, H. Nie, S. Byrn, J.W. Lubach, Investigation of drug-excipient interactions in lapatinib amorphous solid dispersions using solid-state NMR spectroscopy, Mol. Pharm. 12 (2015) 857-866, http://dx.doi.org/10.1021/ mp500692a.
- [191] G.B.H. Ratiopharm, Pharmaceutical Composition Comprising Trametinib, EP 2913048 A1, 2014.
- [192] B.V. Synthon, Pharmaceutical composition comprising Erlotinib hydrochloride, WO 2014118112 (2014)
- [193] C. Liu, Z. Chen, Y. Chen, J. Lu, Y. Li, S. Wang, G. Wu, F. Qian, Improving oral bioavailability of sorafenib by optimizing the "spring" and "parachute" based on molecular interaction mechanisms, Mol. Pharm. 13 (2016) 599-608, http://dx.doi.org/10. 1021/acs.molpharmaceut.5b00837.
- [194] P. Inc, Tofacitinib Oral Sustained Release Dosage Forms, WO 20140271842 A1, 2014
- [195] C.B. Aakeröy, M.E. Fasulo, J. Desper, Cocrystal or salt: does it really matter? Mol. Pharm. 4 (2007) 317-322, http://dx.doi.org/10.1021/mp060126o.
- [196] E. Stoler, J. Warner, Non-covalent derivatives: cocrystals and eutectics, Molecules 20 (2015) 14833-14848, http://dx.doi.org/10.3390/molecules200814833.
- [197] Council of Scientific & Indistrial research, Pharmaceutical cocrystals of gefitinib, WO 2015170345 A1, 2015.
- [198] S.E. Basf, Multicomponent crystals comprising dasatinib and selected cocrystal formers, WO 2013186726 A2, 2013.
- [199] S.E. Basf, Multicomponent crystalline system comprising Nilotinib and selected cocrystal formers, WO 2014060449 A1, 2013.
- [200] FIS, Co-crystals of Lapatinib Monoacid Salts, WO 2015162007, 2015.
- A. Arora, E.M. Scholar, Role of tyrosine kinase inhibitors in cancer therapy, J. [201] Pharmacol. Exp. Ther. 315 (2005) 971-979, http://dx.doi.org/10.1124/jpet.105. 084145
- [202] W.W. Chan, S.C. Wise, M.D. Kaufman, Y.M. Ahn, C.L. Ensinger, T. Haack, M.M. Hood, J. Jones, J.W. Lord, W.P. Lu, D. Miller, W.C. Patt, B.D. Smith, P.A. Petillo, T.J. Rutkoski, H. Telikepalli, L. Vogeti, T. Yao, L. Chun, R. Clark, P. Evangelista, L.C. Gavrilescu, K. Lazarides, V.M. Zaleskas, L.J. Stewart, R.A. Van Etten, D.L. Flynn, Conformational control inhibition of the BCR-ABL1 tyrosine kinase, including the gatekeeper T315I mutant, by the switch-control inhibitor DCC-2036, Cancer Cell 19 (2011) 556-568, http://dx.doi.org/10.1016/j.ccr.2011.03.003.
- [203] WHO, Cancer Factsheet no 297, 2015.
- [204] US Food and Drug administration (FDA), Fast Track, Breakthrough, Therapy, Accelerated Approval, Priority review, fda.gov/forpatients/approvals/fast/ucm20041766. htm 2015 accessed August 19, 2016.
- M. Minekus, The TNO gastro-intestinal model (TIM), Impact Food Bioact. Heal. [205] Springer International Publishing, Cham 2015, pp. 37-46, http://dx.doi.org/10. 1007/978-3-319-16104-4 5.