

# Thermodynamic Potential for the Abiotic Synthesis of Adenine, Cytosine, Guanine, Thymine, Uracil, Ribose, and Deoxyribose in Hydrothermal Systems

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Received: 6 March 2008 / Accepted: 16 April 2008  
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**Abstract** The thermodynamic potential for the abiotic synthesis of the five common nucleobases (adenine, cytosine, guanine, thymine, and uracil) and two monosaccharides (ribose and deoxyribose) from formaldehyde and hydrogen cyanide has been quantified under temperature, pressure, and bulk composition conditions that are representative of hydrothermal systems. The activities of the precursor molecules (formaldehyde and hydrogen cyanide) required to evaluate the thermodynamics of biomolecule synthesis were computed using the concentrations of aqueous  $N_2$ , CO,  $CO_2$  and  $H_2$  reported in the modern Rainbow hydrothermal system. The concentrations of precursor molecules that can be synthesized are strongly dependent on temperature with larger concentrations prevailing at lower temperatures. Similarly, the thermodynamic drive to synthesize nucleobases, ribose and deoxyribose varies considerably as a function of temperature: all of the biomolecules considered in this study are thermodynamically favored to be synthesized throughout the temperature range from 0°C to between 150°C and 250°C, depending on the biomolecule. Furthermore, activity diagrams have been generated to illustrate that activities in the range of  $10^{-2}$ – $10^{-6}$  for nucleobases, ribose and deoxyribose can be in equilibrium with a range of precursor molecule activities at 150°C and 500 bars. The results presented here support the notion that hydrothermal systems could have played a fundamental role in the origin of life, and can be used to plan and constrain experimental investigation of the abiotic synthesis of nucleic-acid related biomolecules.

**Keywords** Nucleobases · Ribose · Deoxyribose · Hydrothermal systems · Thermodynamics

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## Introduction

In recent years many theoretical and experimental studies have tested the hypothesis that ancient deep sea hydrothermal systems provided an environment conducive to the abiotic synthesis of biomolecules that are essential for the emergence of life (Bock and Goode 1996; Hazen et al. 2002; Holm 1992; Holm and Andersson 1995, 2005; Holm et al. 2006; McCollom and Seewald 2007; Nisbet and Fowler 1996; Shock 1990, 1992b, 1996; Shock et al. 2000; Simoneit 2004; Woese 1998; Woese et al. 1990). As a result, abiotic synthesis studies involving amino acids (Amend and Shock 1998, 2000; Hennet et al. 1992; Marshall 1994; Shock and Schulte 1990), lipid-like compounds (McCollom et al. 1999; Rushdi and Simoneit 2001), aldehydes (Shock and Schulte 1993), carboxylic acids (McCollom and Seewald 2001, 2003a, b), alcohols, ketones (Shock and Schulte 1995, 1998), polycyclic, alkylated and hydroxylated aromatic hydrocarbons (McCollom 2003; Williams et al. 2005; Zolotov and Shock 1999), thiols (Rushdi and Simoneit 2005; Schulte and Rogers 2004) and other organic compounds (Foustoukos and Seyfried 2004; McCollom and Simoneit 1999; Rushdi and Simoneit 2004; Shock and McKinnon 1993) have been carried out under hydrothermal conditions using experimental and/or computational–thermodynamic techniques. However, the common organic monomers constituting nucleic acids, that is, adenine, cytosine, guanine, cytosine, thymine, uracil, ribose, and deoxyribose, have received far less attention. A notable exception is the recent work carried out by Franiatte and coworkers in which the stability of aqueous adenine was measured at 300°C under controlled fugacities of H<sub>2</sub>, CO<sub>2</sub>, and N<sub>2</sub> (Franiatte et al. 2008). Exploring potential extraterrestrial sources of nucleobases, Saldino and coworkers have synthesized adenine cytosine, thymine and uracil in one-pot experiments from 0.12 mM formamide (CH<sub>3</sub>NO) at 160°C in the presence of cosmic dust analogues (amorphous olivines), TiO<sub>2</sub> and montmorillonites (Saladino et al. 2003, 2004, 2005b) (for a review of experimental abiotic nucleobase synthesis see Saladino et al. 2005a). Similarly, several other experiments, carried out using high concentrations of precursor molecules, organic solvent extractions and a variety of catalysts under unspecified oxidation states, have produced nucleobases in gas, solid and aqueous phases (Ferris et al. 1968; Hayatsu et al. 1968; Hill and Orgel 2002; Miyakawa et al. 2000; Wakamatsu et al. 1966). But because the conditions under which these experiments were performed do not reflect those of any known current or past hydrothermal systems, their relevance to natural systems is unclear. The purpose of the present study is to explore the thermodynamic potential for the abiotic synthesis of the five common nucleobases (adenine, guanine, cytosine, thymine and uracil) and two sugars (ribose & deoxyribose) that make up nucleic acids (DNA & RNA) from two precursor molecules, formaldehyde (CH<sub>2</sub>O) and hydrogen cyanide (HCN), over a range of pressures, temperatures, and bulk compositions that are characteristic of hydrothermal systems. This is accomplished by first quantifying the thermodynamic potential to synthesize the precursor molecules from N<sub>2</sub>, H<sub>2</sub>, CO<sub>2</sub> and CO, and then calculating the energetics of biomolecular synthesis from the two precursor compounds.

## Precursor Molecules

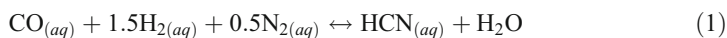
The successful experimental synthesis of organic compounds from the condensation of CH<sub>2</sub>O and HCN, usually referred to as Strecker synthesis, has led to the hypothesis that reactions involving these precursor molecules were responsible for the abiotic synthesis of biomolecules on the early Earth (Miller 1957; Ferris et al. 1978; Schulte and Shock 1993,

1995). Oró and coworkers were among the first to contribute to this idea by synthesizing adenine,  $C_5H_5N_5$ , from HCN (Oró 1960; Oró and Kimball 1961). In addition, it has long been established that carbohydrates of varying carbon number, such as ribose,  $C_5H_{10}O_5$ , can be made abiotically from formaldehyde according to the formose reaction (Butlerov 1861). More recently, Hennet et al. (1992) synthesized several amino acids from  $CH_2O$  and HCN at 150°C. The prevalence of  $CH_2O$  and HCN in ancient or modern hydrothermal systems is not known, but the amount of these compounds that could have existed in prebiotic hydrothermal systems can be estimated by taking into account reactions among simple sources of N, C, H and O in these environments that can be relatively well-constrained.

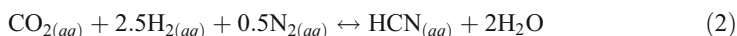
## Hydrogen Cyanide

HCN in hydrothermal systems could come from reactions between  $H_2$ ,  $N_2$ , and  $CO_2$  and CO (Schulte and Shock 1995; Shock 1992a), all of which are considered to have been common species on the early Earth (Kasting 1993). The activities of aqueous HCN in equilibrium with  $N_2$ ,  $H_2$  and CO and  $N_2$ ,  $H_2$  and  $CO_2$  from 0°C to 350°C, at pressures that are either equal to those corresponding to the liquid/vapor saturation curve for  $H_2O$  (P<sub>sat</sub>) or at 500 bars are shown in Fig. 1a,b, respectively. The phase diagram for  $H_2O$  shown in Fig. 2 displays the values of pressure used in the P<sub>sat</sub> calculations as a function of temperature. It can be seen in Fig. 1a,b that the activity of HCN in equilibrium with  $N_2$ ,  $H_2$  and CO or  $CO_2$  decreases substantially as temperature increases and that pressure has little effect on the magnitude of this variation.

The curves in Fig. 1a,b were generated by first calculating the equilibrium constant,  $K$ , for



and



Statements of the law of mass action for these reactions, e.g.,

$$K_1 = \frac{a_{HCN_{(aq)}} a_{H_2O}}{a_{CO_{(aq)}} a_{H_2(aq)}^{1.5} a_{N_2(aq)}^{0.5}} \quad (3)$$

were then rearranged to solve for the equilibrium activity of the species of interest:

$$\log a_{HCN_{(aq)}} = \log K_1 - \log a_{H_2O} + \log a_{CO_{(aq)}} + 1.5 \log a_{H_2(aq)} + 0.5 \log a_{N_2(aq)} \quad (4)$$

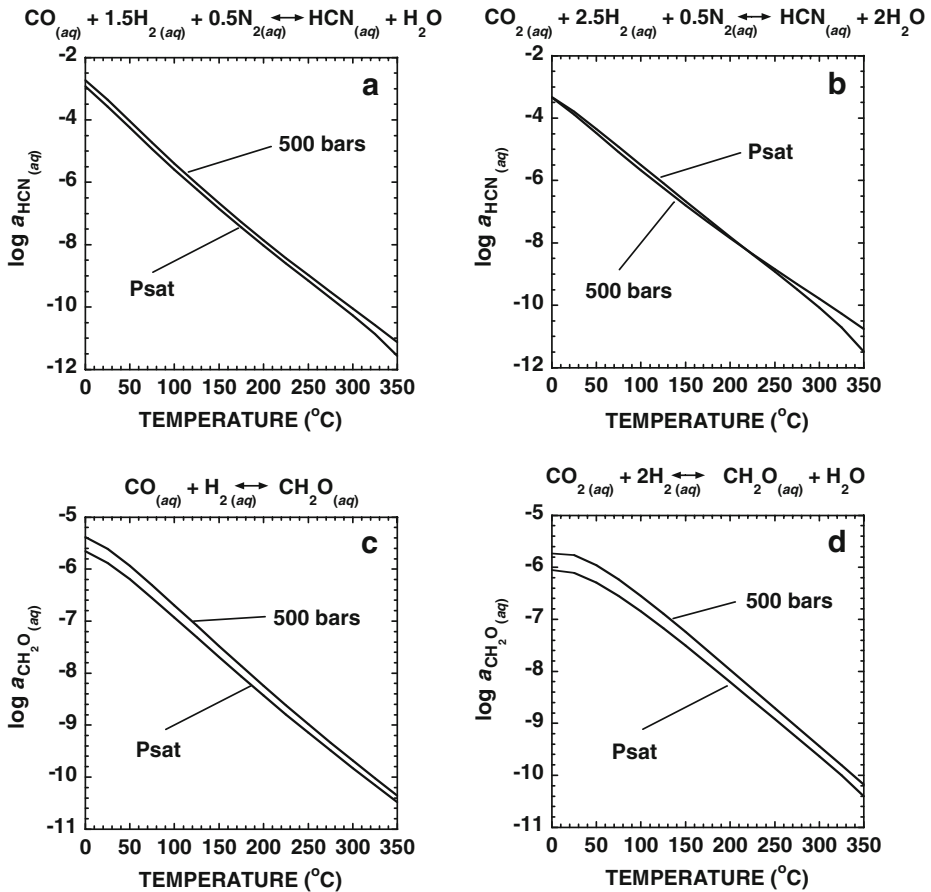
Similarly, the logarithmic equivalent for Reaction (2) can be written:

$$\log a_{HCN_{(aq)}} = \log K_2 - \log a_{H_2O} + \log a_{CO_{2(aq)}} + 2.5 \log a_{H_2(aq)} + 0.5 \log a_{N_2(aq)} \quad (5)$$

where  $K_1$  and  $K_2$  refer to the equilibrium constants of Reactions (1) and (2), respectively. Values of  $K$  for these and the following reactions were calculated using

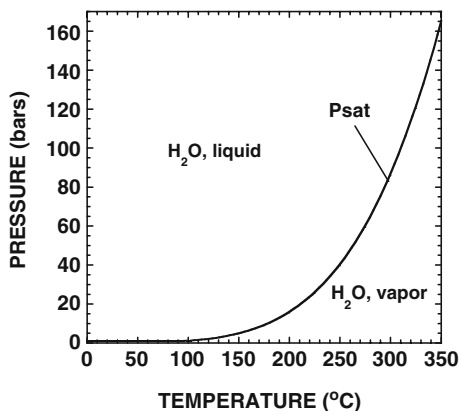
$$K = e^{-\Delta G_r^0/RT} \quad (6)$$

where  $R$  stands for the gas constant,  $T$  represents the absolute temperature and  $\Delta G_r^0$  denotes the standard molal Gibbs energy of reaction. Values of  $\Delta G_r^0$  were computed with the SUPCRT92 software package (Johnson et al. 1992) which relies on the revised Helgeson–



**Fig. 1** Temperature-activity diagrams depicting the activities of  $\text{CH}_2\text{O}$  and  $\text{HCN}$  that are in equilibrium with the reactants and products shown in the reactions above each panel. The concentrations of  $\text{CO}$ ,  $\text{CO}_2$ ,  $\text{N}_2$ , and  $\text{H}_2$  used to generate these figures are given in Table 1

Kirkham–Flowers (HKF) equations of state (Shock and Helgeson 1990; Tanger and Helgeson 1988) to calculate the thermodynamic properties of chemical reactions at elevated temperatures and pressures. The standard state adopted in the present study for aqueous species other than  $\text{H}_2\text{O}$  corresponds to unit activity of the species in a hypothetical one molal solution referenced to infinite dilution at any pressure and temperature (see LaRowe and Helgeson 2007). The activity of liquid  $\text{H}_2\text{O}$  was taken to be 1. The activities of  $\text{N}_2$ ,  $\text{H}_2$ ,  $\text{CO}$  and  $\text{CO}_2$  were specified by multiplying the measured concentrations of these species (Table 1) in the presently-active Rainbow hydrothermal system, located at  $36^{\circ}14' \text{ N}$  on the Mid-Atlantic Ridge at a depth of 2,300 m (Charlou and colleagues reported fluids emanating at  $365^{\circ}\text{C}$  at  $\text{pH}=2.8$  (Charlou et al. 2002)), with activity coefficients,  $\gamma$ , that were computed using the HCh software package (Shvarov and Bastrakov 1999). Because the ionic strength used in the activity coefficient calculations is that of modern seawater, values of  $\gamma$  for neutral species were generally slightly less than one ( $\sim 0.975$ ). This approach was used principally because the composition of ancient hydrothermal systems is unknown, and, at the very least, the modern concentrations represent a proxy for characterizing geochemical solutions that may have prevailed in the past. In addition, this is the rare

**Fig. 2** Liquid–vapor phase diagram for H<sub>2</sub>O

hydrothermal systems for which concentrations of CO, CO<sub>2</sub>, H<sub>2</sub> and N<sub>2</sub> have been reported in the literature. To specify the composition of ancient hydrothermal fluids, Schulte and Shock (1995) followed a different approach that used instead fugacities of CO, CO<sub>2</sub>, and N<sub>2</sub> constrained from estimated ancient atmospheric concentrations of the same species made by Kasting (1993). However, a cursory examination of modern hydrothermal vent compositions (Charlou et al. 2002; Von Damm 1995) reveals that the concentrations of CO, CO<sub>2</sub>, and N<sub>2</sub> are highly variable from one system to another and do not correlate with their concentrations in the modern atmosphere. The assumption that the concentrations of the species in ancient and modern hydrothermal fluids are similar is reasonable because the composition of hydrothermal fluids are primarily determined by their interactions with mantle-derived rocks, whose composition (i.e., oxidation state) has not changed significantly in the last four billion years (Delano 2001).

### Formaldehyde

Formaldehyde, CH<sub>2</sub>O, is often cited as a common prebiotic molecule (Orgel 2004; Shapiro 1988), but the concentration of this compound on the early earth is not known. In order to quantify possible concentrations of aqueous CH<sub>2</sub>O in hydrothermal systems, as with the HCN, equilibrium activities of this species have been calculated using the H<sub>2</sub>, CO and CO<sub>2</sub> concentrations from the Rainbow site (Fig. 1c,d). Although the equilibrium activity of CH<sub>2</sub>O follows the same qualitative trend with temperature as the activity of HCN shown in Fig. 1a,b, these calculations show that higher pressure (500 bars) favors higher equilibrium activities of CH<sub>2</sub>O relative to lower pressure conditions (Psat). However, even under the most favorable conditions of low temperatures and high pressure, the equilibrium activity of CH<sub>2</sub>O, with respect to H<sub>2</sub> and CO and CO<sub>2</sub>, is much less than HCN and 10<sup>-5</sup>. These results reflect the conditions in one modern hydrothermal system only so if the concentrations of H<sub>2</sub>, CO

**Table 1** Concentrations of selected species from Rainbow hydrothermal field (Charlou et al. 2002)

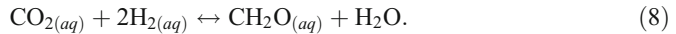
Species	Concentration (mM)
H <sub>2</sub>	16
N <sub>2</sub>	3
CO <sub>2</sub>	16
CO	5×10 <sup>-6</sup>

and  $\text{CO}_2$  would have been higher in ancient hydrothermal systems, the activity of  $\text{CH}_2\text{O}$  in equilibrium with these reactants could also have been greater. Kasting (1993) estimated that during the first several hundred million years of Earth's history the atmosphere contained  $\text{CO} + \text{CO}_2$  equal to 10 bars, a result which suggests that the concentration of precursor carbon molecules in hydrothermal systems could have been different from those today.

The curves in Fig. 1c,d were generated following the strategy outlined above for HCN, but in accordance with



and



The equilibrium activities of HCN and  $\text{CH}_2\text{O}$  shown in Fig. 1 are used in the following section to calculate the thermodynamic drive for the abiotic synthesis of nucleobases, ribose and deoxyribose.

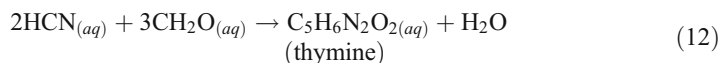
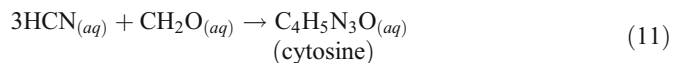
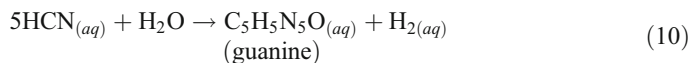
### Nucleobase, Ribose and Deoxyribose Synthesis

In this section, the thermodynamic drive to synthesize the five common nucleobases, ribose and deoxyribose is quantified by calculating the Gibbs energy of reaction for each of these compounds from the precursor molecules HCN and  $\text{CH}_2\text{O}$  as a function of temperature and temperature.

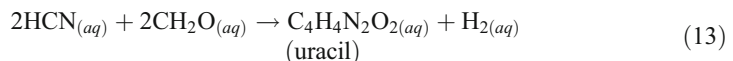
#### Nucleobases

The nucleobases considered in this study are the common purines (adenine and guanine) and pyrimidines (cytosine, thymine, and uracil) found in DNA and RNA. Experimental attempts to synthesize these compounds abiotically have been less successful than for amino acids, especially guanine and uracil (Saladino et al. 2005a).

The overall Gibbs energy of reaction,  $\Delta G_r$ , for the synthesis of the five common nucleobases from aqueous HCN and  $\text{CH}_2\text{O}$  are calculated for



and



and are shown as a function of temperature at Psat and 500 bars in Fig. 3a–e. It can be seen in this figure that the values of  $\Delta G_r$  are negative for Reactions 9–13 from 0°C to between 175°C and 240°C, depending on the nucleobase. Above these temperatures, Reactions 9–13 are no longer favored. The small difference in values of  $\Delta G_r$  between the Psat and 500-bar curves indicate that pressure has little effect on these reactions. The dashed lines in Fig. 3a–e represent values of  $\Delta G_r^0$  for the respective reactions. These lines are shown to illustrate the quantitative difference between the energetics of chemical reactions when the activities of the reactants and products are ( $\Delta G_r$ ) and not ( $\Delta G_r^0$ ) taken into account.

Values of the Gibbs energy of reaction were calculated using

$$\Delta G_r = -RT \ln \left( \frac{K}{Q} \right) \quad (14)$$

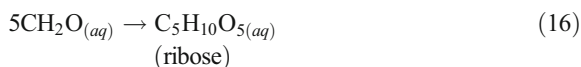
where  $Q$  refers to the activity product, which is defined by

$$Q = \Pi a_i^{\nu_i} \quad (15)$$

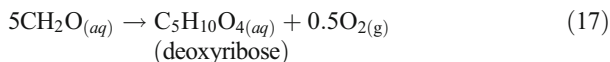
where the symbols  $a_i$  and  $\nu_i$  denote the activity and stoichiometric reaction coefficient of the  $i$ th species, respectively. The standard molal thermodynamic properties and revised HKF equation of state parameters for the nucleobases required to calculate values of  $K$  were taken from LaRowe and Helgeson (2006). The values of  $a_i$  for HCN and CH<sub>2</sub>O were taken from the results shown in Fig. 1b,d. The  $\Delta G_r$  curves, calculated using CO as a carbon source, are not included here because they are nearly identical to the CO<sub>2</sub>-derived calculations. The activities of H<sub>2</sub> required to evaluate Eq. 15 for reactions 10 & 13 were taken from the Rainbow hydrothermal system (Table 1). The activities of the nucleobases were taken to be 10<sup>-12</sup>, after (McCollom and Amend 2005).

### Ribose and Deoxyribose

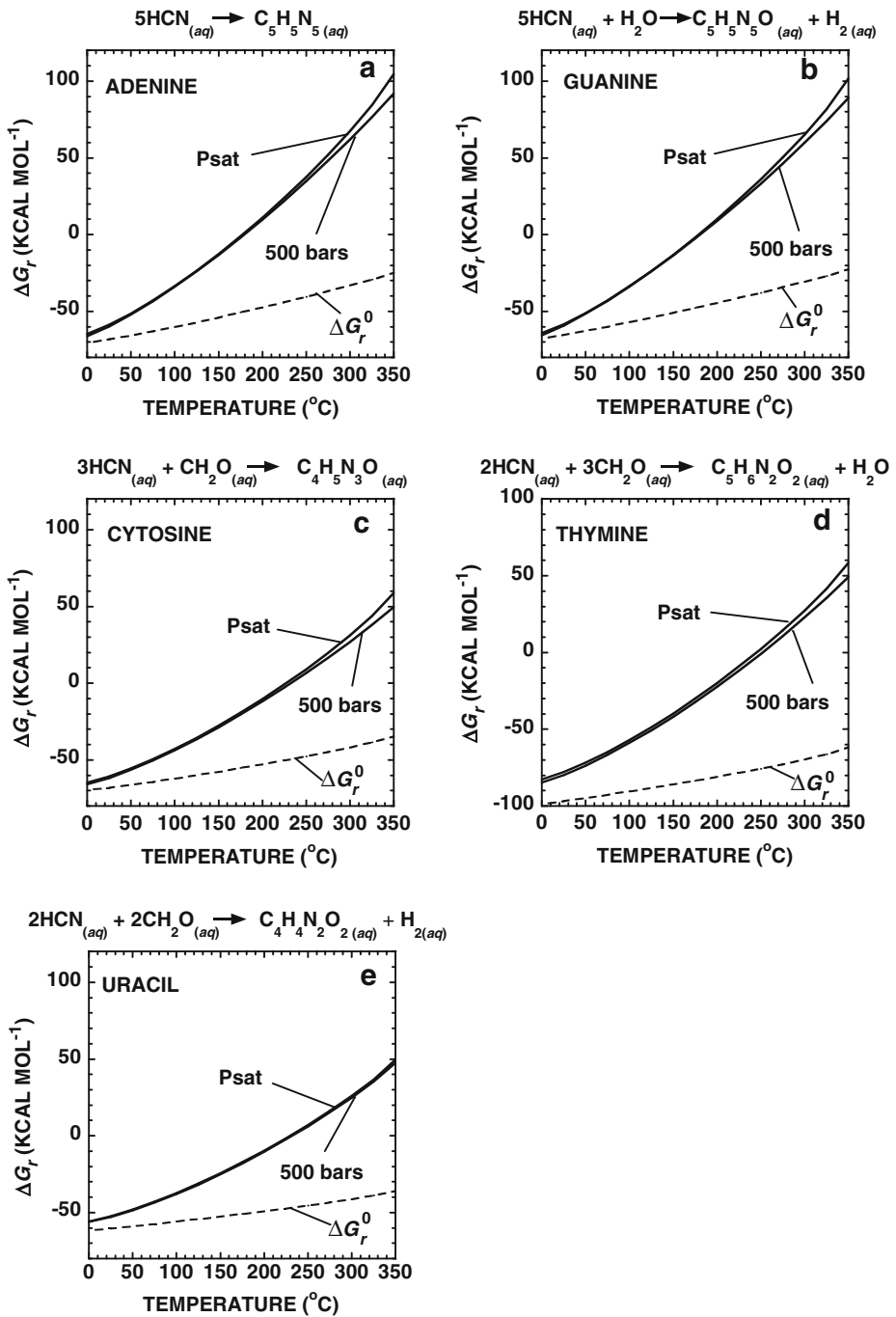
Values of  $\Delta G_r$  for the synthesis of aqueous ribose, C<sub>5</sub>H<sub>10</sub>O<sub>5(aq)</sub>, and deoxyribose, C<sub>5</sub>H<sub>10</sub>O<sub>4(aq)</sub>, from CO<sub>2</sub>-derived formaldehyde,



and

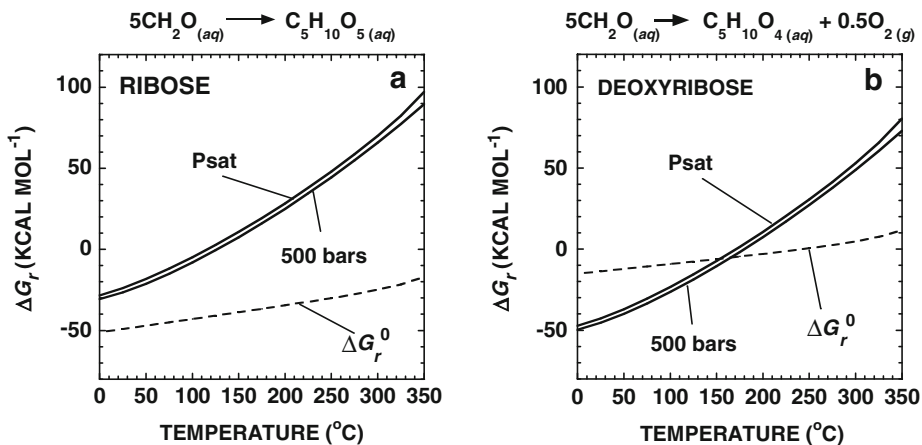


are shown as a function of temperature at Psat and 500 bars in Fig. 4a,b, respectively. For reference, the dashed lines in these figures stand for values of  $\Delta G_r^0$  for the respective reactions. The overall Gibbs energy of reaction for Reactions 16 and 17 are negative from 0°C to ~140°C and 0°C to ~175°C, respectively. The temperatures at which these reactions become endergonic at Psat and 500 bars differ by 5°C or less. These calculations were carried out using Eqs. 14 and 15 and the thermodynamic properties of ribose and deoxyribose (Amend and Plyasunov 2001; LaRowe and Helgeson 2006). The values of  $a_i$  for CH<sub>2</sub>O required to evaluate Eq. 15 were taken from the equilibrium activities of CO<sub>2</sub>-derived formaldehyde shown in Fig. 1d. The activities of ribose and deoxyribose were set equal to 10<sup>-12</sup>. The values of oxygen fugacity,  $f_{\text{O}_2}$ , used to compute  $Q$  for Reaction 17 were determined by calculating the oxygen fugacity that is in equilibrium with the fayalite, magnetite, quartz (FMQ) buffer for the temperatures and pressures considered here, a



**Fig. 3** Standard (dashed lines) and overall Gibbs energies of reaction ( $\Delta G_r^0$  and  $\Delta G_r$ , respectively) for the synthesis of nucleobases from  $\text{CH}_2\text{O}$  and  $\text{HCN}$  in accordance with the reactions shown above each panel. The activities of  $\text{CH}_2\text{O}$  and  $\text{HCN}$  used to construct these diagrams are shown in Fig. 1, while those for the nucleobases were all set equal to  $10^{-12}$





**Fig. 4** Standard (*dashed lines*) and overall Gibbs energies of reaction ( $\Delta G_r^0$  and  $\Delta G_r$ , respectively) for the synthesis of ribose and deoxyribose from  $\text{CH}_2\text{O}$  in accordance with the reactions shown above each panel. The activities of  $\text{CH}_2\text{O}$  used to construct these diagrams are shown in Fig. 1, while those for the sugars were all set equal to  $10^{-12}$ . The values of oxygen fugacity used in the calculations for deoxyribose are taken to be equal to those in equilibrium with the FMQ buffer, which is characteristic of hydrothermal systems (Schulte and Shock 1995)

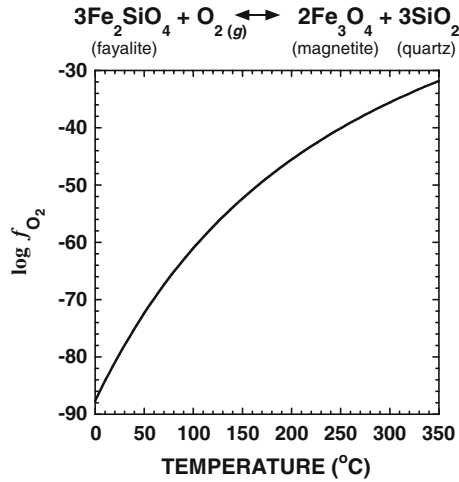
common proxy for the oxidation state of submarine hydrothermal systems (Schulte and Shock 1995). These values are shown in Fig. 5.

The feasibility of Reaction 16 representing a likely pathway for the abiotic synthesis of ribose has been doubted (Shapiro 1988, 1995). However, due to recent progress in determining the chemical environments that promote the abiotic synthesis of ribose (Orgel 2004; Ferris 2005; Holm et al. 2006), the likelihood that Reaction 16 represents a plausible origin of this fundamental sugar has been substantiated. Initially, a number of studies showed that, under indiscriminating laboratory conditions, the formose reaction produces dozens of aldoses, ketoses, and sugar alcohols from  $\text{CH}_2\text{O}$ , i.e., very little ribose (Shapiro 1988). However, it has recently been found that in the presence of  $\text{Pb}^{2+}$ , aldopentoses are preferentially formed during the formose reaction and some of these reaction products are isomerized to ribose (Zubay 1998; Zubay and Mui 2001). Also, Ricardo et al. (2004) showed that borate minerals preferentially bind pentoses, ribose most so, thus stabilizing and potentially concentrating them over other pentoses. Furthermore, when phosphate is present, ribose-phosphate compounds, the repetitive-unit backbone of RNA, are formed and do not readily convert into other molecules (Müller et al. 1990).

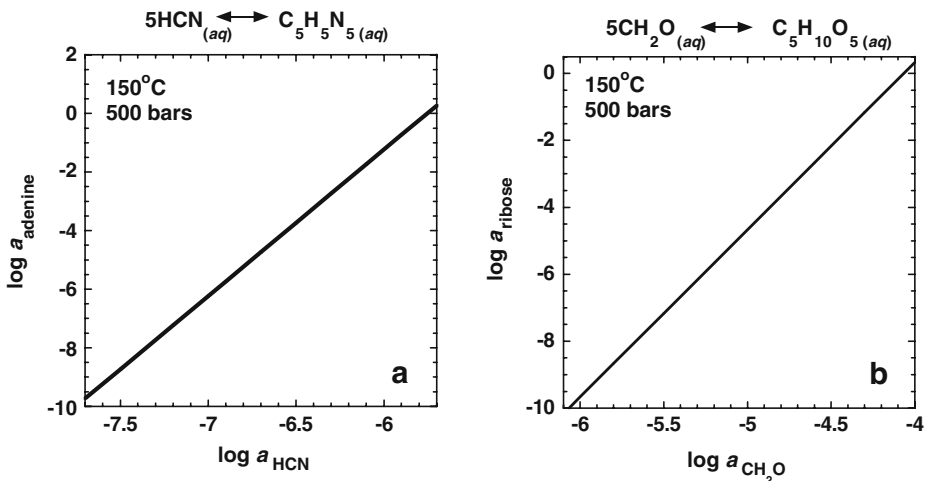
### Variable $\text{CH}_2\text{O}$ and HCN Activities

Because the concentrations of  $\text{N}_2$ ,  $\text{H}_2$ ,  $\text{CO}$ , and  $\text{CO}_2$  used to calculate the activities of  $\text{CH}_2\text{O}$  and HCN in this study are highly variable in modern systems (Von Damm 1995) and, likely, ancient hydrothermal systems, the activities of the precursor molecules produced in them may have also been quite variable. In order to quantitatively assess the impact that variable precursor molecule concentrations would have on the production of nucleobases, ribose and deoxyribose, the activities of these species in equilibrium with varying activities of  $\text{CH}_2\text{O}$  and HCN and, where relevant,  $\text{H}_2$  and  $\text{O}_2$ , were calculated under temperature and pressure conditions that favor their formation.

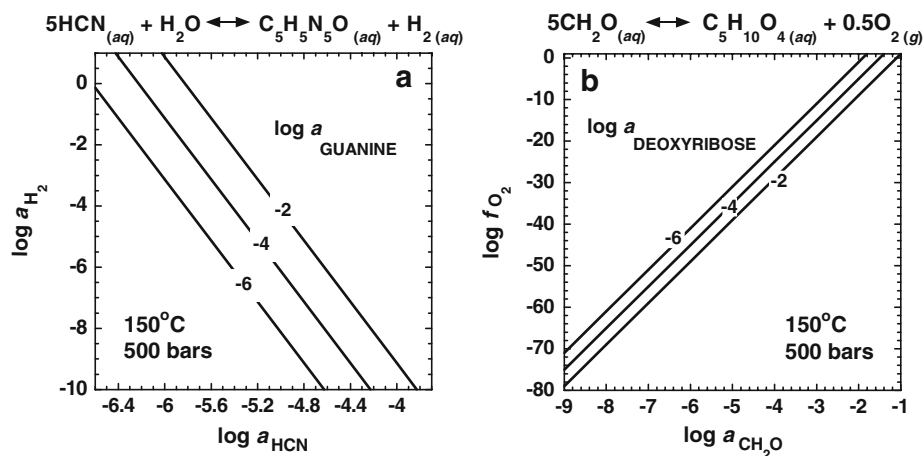
**Fig. 5** Logarithm of the fugacity of  $O_2$  ( $g$ ) that is in equilibrium with the minerals fayalite, magnetite and quartz (FMQ redox buffer) in accordance with the reaction written at the top of the figure



The activities of adenine and ribose in equilibrium with varying activities of HCN and  $\text{CH}_2\text{O}$ , respectively, at  $150^\circ\text{C}$  and 500 bars are shown in Fig. 6a,b. Activities of adenine between  $10^{-2}$ – $10^{-6}$  are in equilibrium with those of HCN equal to from  $10^{-6}$  to  $10^{-7}$ . Similarly, ribose can achieve equilibrium activities between  $10^{-2}$ – $10^{-6}$  with  $\text{CH}_2\text{O}$  activities ranging from  $10^{-4.5}$  to  $10^{-5.2}$ . The activities of the other biomolecules considered in this study depend on more than one chemical variable and are thus represented on the plane intersecting this parameter space. For example, Fig. 7a,b show contoured activities of guanine and deoxyribose as a function of the activities of HCN and  $\text{CH}_2\text{O}$ , respectively, and the oxidation state. In the case of guanine, the oxidation state is described by the activity of  $\text{H}_2$ ,  $a_{\text{H}_2}$ , while that for deoxyribose is the fugacity of oxygen. It can be seen in both Fig. 7a,b that large activities of guanine and deoxyribose ( $10^{-2}$ – $10^{-6}$ ) are in equilibrium with relatively small activities of their respective precursor molecules ( $\log a$



**Fig. 6** Activities of adenine (a) and ribose (b) in equilibrium with variable activities of HCN and  $\text{CH}_2\text{O}$ , respectively, at  $150^\circ\text{C}$  and 500 bars. The reaction written above each panel represents the equilibrium relationship between the species of interest and precursor molecule



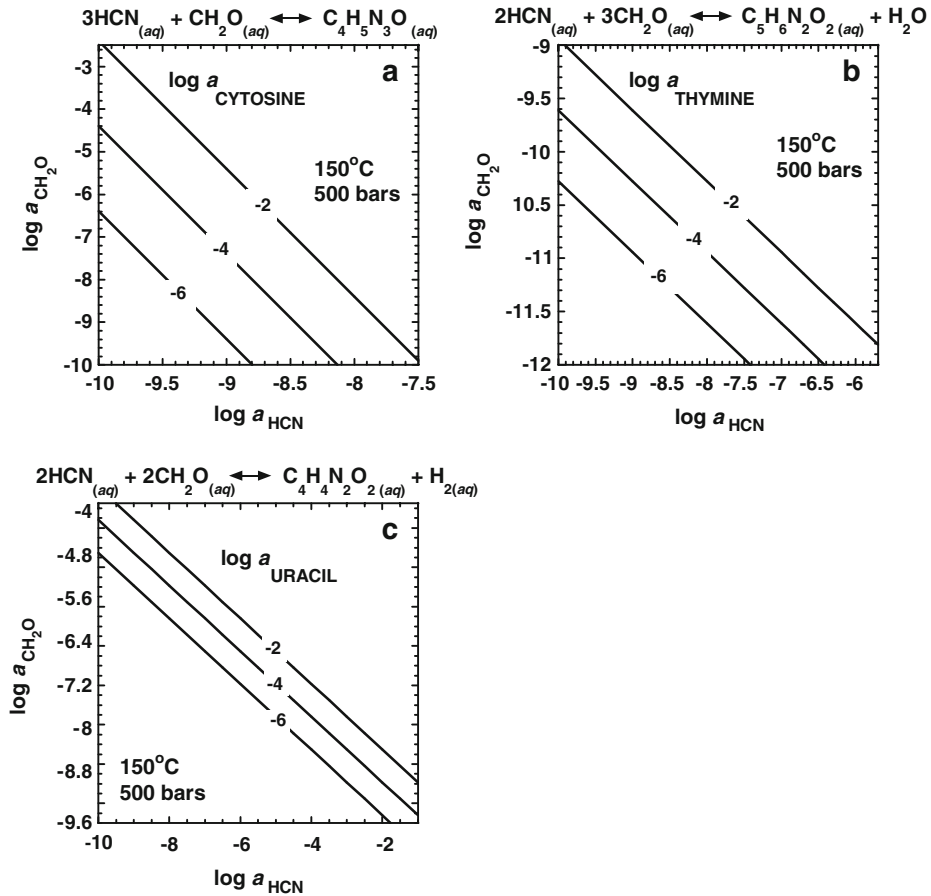
**Fig. 7** Contours of logarithms of the activities of guanine (a) and deoxyribose (b) in equilibrium with variable activities of HCN and  $\text{H}_2$  and  $\text{CH}_2\text{O}$  and  $\text{O}_2$ , respectively, at  $150^\circ\text{C}$  and 500 bars. The reaction written above each panel represents the equilibrium relationship between the species of interest and precursor molecules

$\text{HCN} = -3.8$  to  $-6.6$  in the case of guanine and  $\log a \text{CH}_2\text{O} = -1$  to  $-9$  for deoxyribose). Also, in both cases, for a given activity of precursor molecule, more reducing conditions favor higher activities of guanine and deoxyribose.

Contoured equilibrium activities of cytosine, thymine and uracil are shown as a function of the activities of  $\text{CH}_2\text{O}$  and HCN in Fig. 8a–c. All three of these pyrimidines can coexist at high activities ( $10^{-2}$ – $10^{-6}$ ) with relatively low activities of the precursor molecules ( $10^{-10}$  or lower) at  $150^\circ\text{C}$  and 500 bars.

## Conclusions

Thermodynamic calculations have revealed that adenine, guanine, cytosine, thymine, uracil, ribose, and deoxyribose can be synthesized from the precursor molecules  $\text{CH}_2\text{O}$  and HCN under temperatures, pressures, and fluid compositions that are characteristic of hydrothermal systems. However, nucleobase and sugar synthesis is thermodynamically favored only at the lower end of the temperature range considered here. This finding corroborates previous studies which have suggested that organic synthesis in submarine hydrothermal systems most likely did not occur in black smoker vent sites where the temperatures can exceed  $400^\circ\text{C}$ , but more plausibly on the distal, off-axis, portions of hydrothermal systems where the temperature is lower (Shock 1990, 1992a, b). Greater activities of the precursor molecules,  $\text{CH}_2\text{O}$  and HCN, can co-exist in equilibrium with  $\text{CO}$ ,  $\text{CO}_2$ ,  $\text{H}_2$ , and  $\text{N}_2$  as measured in modern hydrothermal systems at cooler temperatures. Activities of HCN corresponding to concentrations in the millimolar range have been calculated to exist at the lower end of the temperature spectrum considered in this study regardless of whether it was generated from  $\text{CO}$  or  $\text{CO}_2$ . However, the concentrations of  $\text{CO}$ ,  $\text{CO}_2$ ,  $\text{H}_2$ , and  $\text{N}_2$  from a modern hydrothermal system may not be representative of past geologic conditions. If the concentrations of these building block molecules would be higher, as hypothesized for the early-Earth atmosphere (Kasting 1993), then higher activities of  $\text{CH}_2\text{O}$  and HCN could then coexist in equilibrium with  $\text{CO}$ ,  $\text{CO}_2$ ,  $\text{H}_2$ , and  $\text{N}_2$ . It follows that the synthetic potential for



**Fig. 8** Contours of logarithms of the activities of cytosine (a), thymine (b) and uracil (c) in equilibrium with variable activities of HCN and CH<sub>2</sub>O at 150°C and 500 bars. The reaction written above each panel represents the equilibrium relationship between the species of interest and precursor molecules

the nucleobases, ribose and deoxyribose would also be greater than what is reported in the present study. Therefore, we have explored the range of CH<sub>2</sub>O and HCN activities on the equilibrium activities of nucleobase, ribose, and deoxyribose at what can be considered one particular hydrothermal flank condition (150°C and 500 bars). Under these conditions, activities of all of the biomolecules considered here are in equilibrium with the precursor molecules at concentrations comparable to those in modern living organisms (Voet et al. 1999).

Complementing other studies that have shown that various biological and organic molecules such as amino acids (Amend and Shock 2000), carboxylic acids, alcohols, and ketones (Shock and Schulte 1998) can be synthesized in hydrothermal systems, the results of this study support the hypothesis that hydrothermal systems could have served as efficient anabolic reactors for building the molecules that are essential to living organisms. The thermodynamic calculations shown here also provide constraints on the temperature, pressure and bulk composition necessary for the synthesis of these fundamental biomolecules in hydrothermal systems. Our results provide a useful theoretical framework

for the design of experiments aimed at synthesizing nucleobases and sugars from inorganic precursors.

**Acknowledgments** This work is supported by the Netherlands Organization for Scientific Research (NWO) grant number 815.01.008.

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