

## C-Glycosylflavonoids in the Leaves of Gymnosperms

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The finding of vitexin derivatives as major flavonoids in needles of *Larix laricina*,<sup>1</sup> *L. sibirica*,<sup>2</sup> *L. leptolepis*,<sup>3</sup> *L. decidua*,<sup>4</sup> *L. gmelinii*<sup>4</sup> and, to a lesser extent, in those of *L. occidentalis*<sup>4</sup> instigated a literature survey on C-glycosides within the gymnosperms. Surprisingly, no reports had been made at the time of Alston's review in 1968,<sup>5</sup> although many species had been investigated for flavonoids.<sup>6</sup> Apart from the *Larix* compounds, C-glycosides had been reported in only two other species; a vitexin-like compound from *Ephedra americana*<sup>7</sup> and vitexin and orientin as minor constituents in *Dioon spinulosum* leaves.<sup>8</sup> The question, therefore, arose whether the larch

may be exceptional in this respect within the Pinaceae, or even among the Gymnospermae. Therefore, leaves of a 46 species of gymnosperms were subjected to a screening for C-glycoflavonoids occurring as major constituents.

Fresh material was collected from the Pinetum Blijdenstein, Hilversum and from the Cantonspark, Baarn, both in the Netherlands, in Sept. 1973. Extracts were compared directly and as acid hydrolysates; where C-glycosides were suspected they were isolated chromatographically and their nature was determined by *R<sub>f</sub>*, UV and spectral shifts.

C-Glycosides (vitexin and a related com-

TABLE 1. C-GLYCOSYL-FLAVONOIDS IN GYMNOSPERMS

Family	Species	C-glycosyl Flavonoid*
Cycadaceae	<i>Ceratozamia mexicana</i> Brongn.	—
	<i>Cycas taiwaniana</i> Carruth.	V + G
	<i>Dioon edule</i> Lindl.	—
	<i>D. spec.</i>	—
	<i>Encephalartos lehmannii</i> Lehm.	—
Ginkgoaceae	<i>Zamia media</i> Jacq.	—
	<i>Ginkgo biloba</i> L.	—
Araucariaceae	<i>Araucaria araucana</i> (Molina) C. Koch.	—
	<i>A. bidwillii</i> Hook.	—
Cephalotaxaceae	<i>Cephalotaxus fortunei</i> Hook.	—
	<i>C. harringtoniana</i> (Forbes) C. Koch	—
Cupressaceae	<i>Juniperus communis</i> L.	—
	<i>J. chinensis</i> L.	—
	<i>Chamaecyparis formonensis</i> Mats.	—
	<i>Thuja occidentalis</i> L.	—
	<i>Callitris robusta</i> R. Br. ex Mirb.	—
	<i>Tetraclinis articulata</i> (Vahl) Mast.	—
	<i>Cupressus sempervirens</i> L.	—
<i>Heyderia decurrens</i> C. Koch	—	

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TABLE 1. *Continued*

Taxodiaceae	<i>Cryptomeria japonica</i> (L.f.) D. Don.	—
	<i>Metasequoia glyptostroboides</i> Hu & Cheng	—
	<i>Sequoia sempervirens</i> (D. Don in Lamb.) Endl.	V
	<i>Sequoiadendron giganteum</i> (Lindl.) Buchh.	—
Taxaceae	<i>Taxodium ascendens</i> Brongn.	—
	<i>Taxus baccata</i> L.	—
	<i>T. cupidata</i> Sieb. & Zucc.	—
	<i>T. canadensis</i> Marsh.	—
	<i>Torreya californica</i> Torr.	—
	<i>T. grandis</i> Fort.	—
Pinaceae	<i>T. nucifera</i> (L.) Sieb. & Zucc.	—
	<i>Abies balsamea</i> (L.) Mill.	—
	<i>A. grandis</i> (Dougl.) Lindl.	—
	<i>Cedrus libani</i> A. Rich. in Bory	—
	<i>Keteleeria fortunei</i> (A. Murr.) Carr.	—
	<i>Larix sibirica</i> Ledeb.	V
	<i>L. laricina</i> (Du Roi) K. Koch†	V
	<i>L. leptolepis</i> (Sieb. & Zucc.) Gord.†	V
	<i>L. decidua</i> Mill.†	V
	<i>L. gmelinii</i> (Rupr.) Kuzenevat	V
	<i>L. occidentalis</i> Nutt.†	V
	<i>Picea breweriana</i> S. Watson	—
	<i>Pinus jeffreyi</i> Grev. & Balf.	—
	<i>P. sylvestris</i> L.	—
	<i>Pseudolarix amabilis</i> (J. Nels) Rehd.	—
<i>Pseudotsuga menziesii</i> (Mirbel) Franco	—	
Podocarpaceae	<i>Tsuga chinensis</i> (Franch.) Britzel	—
	<i>Podocarpus elatus</i> R. Br.	V
	<i>P. falcatus</i> (Thunb.) R. Br. ex Mirb.	—
Ephedraceae	<i>Saxegothea conspicua</i> Lindl.	V
	<i>Ephedra altissima</i> Desf.	G
	<i>E. distachya</i> L.	G

\* V—vitexin or vitexin glycoside; G—glycoflavone related to vitexin.

† Results from previous work on *Larix*.

pound) were found in 7 species (including one *Larix* spec.) distributed in 5 families (Table 1). It appears that C-glycoflavonoids are widely, but infrequently distributed in the

gymnosperms. The results suggest that there may be some genus-specific distribution but this can only be proved by investigation of many more species.