

ISOTOPICALLY EXCHANGEABLE PHOSPHATE IN FRESHWATER SEDIMENTS: EFFECTS OF U.V.-IRRADIATION, FORMALDEHYDE, SOLID/SOLUTION RATIO, AND pH ON ITS EXPERIMENTAL DETERMINATION

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Abstract—The influence of bioactivities, solid/solution ratio and the pH on the isotopic exchangeability of phosphate in a freshwater sediment was investigated. From the comparison of the results obtained for the same sample in the presence or absence of formaldehyde, it is concluded that microorganisms can effect the analysis for isotopically exchangeable phosphate. Irradiation with u.v.-light caused a sharp rise in isotopic exchangeability. In the pH-range 6.6–8.4 isotopic exchangeability of phosphate increases with decreasing pH-value which is attributed to an easier exchange of $H_2PO_4^-$ than of HPO_4^{2-} . The influence of the solid/solution ratio on the isotopic exchangeability of phosphate in the solid phase is small or nil. However, because of the relatively large amount of phosphate that goes into solution, the total isotopic exchangeability E , of phosphate in the solid and liquid phase together is strongly increased at a low solid/solution ratio. From these results it is concluded that in order to make a meaningful comparison of isotopically exchangeable phosphate in different soils or sediments it is essential to work at a nearly constant pH and solid/solution ratio. Such a comparison was made for 26 freshwater sediments from the Rhine/Meuse delta, in the presence and absence of 0.17 mol l^{-1} formaldehyde as a biological inhibitor and it is concluded that addition of the latter is essential. The lowest total isotopic exchangeabilities of phosphate, (15–25) were measured in the sediments collected from the Haringvliet, whereas higher values (40–80) were found in the sediments from the Brielse Meer and the Grote Rug. This could well be indicative of a similar variance in the biological availability of the phosphate in the investigated sediments.

Key words—freshwater sediment, isotopically exchangeable phosphate, pH, phosphorus, radio-phosphorus, sediment–water exchange, formaldehyde, u.v.-destruction, eutrophication

INTRODUCTION

The amount of biologically available phosphate in sediments or suspended matter is an important parameter for the quality of fresh water (Syers *et al.*, 1973). Similarly, in soils the amount of biologically available phosphate determines whether or not phosphate is a limiting factor for growth. The best way to determine the availability of phosphate to biota is to use bioassays or pot experiments. However, these tests are often time-consuming and labour-intensive. Consequently much effort has been put into finding reliable alternative methods.

In the agricultural sciences isotopic dilution analysis is frequently used to determine the isotopic exchangeability (E -value) of phosphate in soils (Amer, 1962; Becket and White, 1964; Mekhael *et al.*, 1965; Amer *et al.*, 1969; Ryden and Syers, 1977).

Li *et al.* (1972, 1973) used this method for freshwater sediments. Although there is no consensus

about the precise meaning of isotopically exchangeable phosphate in soils or sediments it is generally agreed upon that such phosphate has a strong positive correlation with biologically available phosphate (Dalal and Hallsworth, 1977; Bowman *et al.*, 1978; Bowman and Olsen, 1979; Le Mare, 1981; Venkat Reddy *et al.*, 1982; Kucey and Bole, 1984). The main advantage of the isotopic dilution method is that the analysis can be done under natural conditions.

Three techniques of isotopic dilution analysis are used: carrier-free, with carrier, and inverse dilution. For most soils the choice of the method seems to be free. Only in the case of a high phosphate fixing capacity of the soil does the inverse dilution technique give better correlations with the results of greenhouse experiments (Dalal and Hallsworth, 1977).

When we compare the various experimental methods used to measure the E -value, striking differences are found in the pH-control, solid/solution ratio and

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equilibration time. Evidently there is a lack of standardization. Therefore it is almost impossible to make a direct comparison between exchangeabilities found by different investigators. The purpose of this investigation was to determine the influence of biological processes, the solid/solution ratio and the pH on the isotopic exchangeability of phosphate in Dutch freshwater sediments. We studied the effect of bioactivities by determining the E -value of a sample in an untreated state, of the same sample irradiated with u.v.-light, and again of the same sample after addition of formaldehyde as a general biological inhibitor. Based upon the findings in these experiments a comparison was made of exchangeable phosphate in a number of sediments from the Rhine and the Meuse delta.

THEORY

The experimental procedure for determining the isotopically exchangeable phosphate can be expressed in the following manner. In the experiments G (kg) sediment loaded with c_{SO} (mg P kg⁻¹) orthophosphate is suspended in V litres of ionic medium yielding at equilibrium a phosphate concentration c_L (mg P l⁻¹) in the liquid phase and c_S (mg P kg⁻¹) in the solid phase.

The mass balance for the phosphate in this model is:

$$c_{SO}G = c_L V + c_S G. \quad (1)$$

Then ³²PO₄ is added, giving an initial ³²PO₄-concentration c_{LO}^* (Bq l⁻¹) and, at equilibrium, a final concentration c_L^* (Bq l⁻¹) whereby a fraction $f = c_L^*/c_{LO}^*$ of the total quantity of ³²PO₄ remains in solution.

This fraction f is easily determined, because it is equal to the counting rate of an aliquot of the liquid phase divided by that of an aliquot of a blank solution to which an equal quantity of radioactivity had been added.

Defining E as the fraction of the phosphate mass in the sediment which actually participates in the isotopic exchange, then according to the principles of isotopic dilution f is also equal to the quotient of the mass of isotopically exchangeable phosphate in the solution ($= c_L V$) and in the total system ($= c_L V + E c_S G$).

After rearrangement and substitution of (1) for E we find:

$$E = \frac{(1-f)c_L V}{f(c_{SO}G - c_L V)}. \quad (2)$$

The total exchangeability E_t in the system can be defined as the quotient of the mass of isotopically exchangeable phosphate and the total mass of phosphate:

$$E_t = \frac{c_L V + E c_S G}{c_{SO}G} = \frac{c_L}{f c_{SO}G/V}. \quad (3)$$

The values of E and E_t give directly the amount of isotopically exchangeable phosphate on the sediment ($E c_S$ mg P kg⁻¹) and in the total system ($E_t c_{SO}$ mg P kg⁻¹). It is obvious that these two quantities have the following relationship:

$$E_t c_{SO} = (c_{SO} - c_S) + E c_S. \quad (4)$$

Here $(c_{SO} - c_S)$ represents the orthophosphate which has gone into solution. E_t is the best measure for the isotopic exchangeability of phosphate in a given sediment, because it includes both the contribution of the phosphate that has remained on the sediment and the contribution of the dissolved phosphate which, by definition, is 100% isotopically exchangeable.

MATERIALS AND METHODS

Samples

Two samples were used to investigate the influence of biological processes, the solid/solution ratio, and the pH on the exchangeability of phosphate. The first (M1) was a mixture of suspended particulate matter from freshwater in the Rhine/Meuse delta.

This particulate matter was collected at various locations (Nieuwe Merwede, Hollands Diep, Haringvliet) with a continuous-flow centrifuge. The second sample (L2) was a sediment collected at one site in the Brielse Meer. Three extraction techniques were used to characterize these samples (Hieltjes and Lijklema, 1980):

- (1) 1.0 M (mol l⁻¹) NH₄Cl-extractable PO₄ (loosely bound)
- (2) 0.1 M NaOH-extractable PO₄ (Al- and Fe-bound)
- (3) 0.5 M HCl-extractable PO₄ (Ca-bound).

The total phosphorus content c_{SO} was determined after destruction of the samples with H₂SO₄/K₂S₂O₈ in an autoclave at 2 bar (Eisenreich *et al.*, 1975). Table 1 shows the results of these chemical analyses.

Stock suspensions of samples M1 and L2, each with a particle size < 2 μm were prepared by suspending the samples in 10⁻² M NaCl. The solid/solution-ratio was then determined by evaporating a known volume to dryness followed by a drying period of 12 h at 383 K. The solid/solution ratio was found by weighing and correcting for the weight of the dissolved NaCl. All sampling from the stock suspensions and from the suspensions in the experiments was done with a pipette while the suspensions were being stirred thoroughly in order to maintain a constant solid/solution-ratio. This technique was checked by weighing and proved to be reliable.

In some cases part of the stock suspension of sample M1 was irradiated for 24 h with u.v.-light. The irradiation took

Table 1. Discrete inorganic P fractions in samples M1 and L2

Sample	c_{SO} (mg P kg ⁻¹)	NH ₄ Cl-P (mg P kg ⁻¹)	NaOH-P (mg P kg ⁻¹)	HCl-P (mg P kg ⁻¹)
M1	5400	90	4100	30
L2	3300	50	2300	50

place in a photochemical reactor with quartz immersion well (Ace Glass Inc.) equipped with a 450 W medium pressure Hg-lamp (Hanovia); during the irradiation the temperature of the suspension stayed below 308 K. At the start of the irradiation 0.02 M H_2O_2 was added (Armstrong *et al.*, 1966, 1968).

Twenty-six sediment samples from the Rhine/Meuse delta were used for the comparison of phosphate exchangeability and were taken from undisturbed cores collected at locations in the Brielse Meer, Haringvliet, and Grote Rug. These samples were taken from the oxidized and/or reduced zones in the sediment cores. These zones were distinguished either visually on the basis of colour difference or electrochemically by measurement of the redox potential. Most of the water in the pores of the sediments was removed using a filter-press at 6 bar nitrogen pressure. Then the samples were freeze-dried and homogenized. Preparation, sampling and determination of the solid/solution-ratio of the stock suspensions in 10^{-2} M NaCl were the same as for the two test samples (M1 and L2).

The first four columns of Table 6 give the code of the sediment core, the origin of the core, the state of oxidation of the sub-sample and the total phosphate content c_{SO} respectively. When the codes in the table are identical this means that sub-samples were taken from both the oxidized and the reduced zone of the same sediment core. Only in the case of cores A9 and A10 were two samples taken from the reduced zone, the second being taken at a greater depth in the core.

Experimental procedures

The sample containers used were of polyethylene and were cleaned with 6 M HCl. The possible interference by adsorption of phosphate on the container wall was investigated over a period of 24 days. Adsorption at pH 7 and 8 (ionic medium 2×10^{-3} M KCl) proved to be <4% for aqueous concentrations exceeding 0.1 mg P l^{-1} .

Preliminary experiments with sediments from the Rhine/Meuse delta frequently yielded irrational and irreproducible exchangeabilities. This was presumed to be the result of biological activities. Samples from a sediment core were used to investigate the influence of formaldehyde (CH_2O) concentration between 0.003 and 0.17 M. Based on these results (cf. Table 5) all experiments were repeated in the presence of 0.17 M CH_2O as a general biological inhibitor, as was earlier used by Pomeroy *et al.* (1965).

Consequently the following experimental procedure was chosen. Aliquots of 75 ml suspension of the samples with known solid/solution-ratio in 10^{-2} M NaCl were prepared. Optionally 1 ml 35% CH_2O was added to give a concentration of 0.17 M (Pomeroy *et al.*, 1965).

In all experiments samples were continuously agitated by placing the containers with sample suspensions on a reciprocating shaker mounted in a waterbath kept at 298 ± 1 K.

After 20 days of contact time 20 ml of suspension was centrifuged (20 min at $2 \times 10^4 \text{ m s}^{-2}$) and used to determine c_L (Murphy and Riley, 1962). Then 1 ml (75 kBq) $^{32}\text{PO}_4$ -solution, with a negligible amount of $^{31}\text{PO}_4$ -carrier, was added. After 27, 29 and 31 days the pH of the suspension was measured and 2 ml of suspension was centrifuged and used to determine c_L^* by means of Cerenkov-counting (Packard 2425 liquid scintillation counter). Some samples were slightly coloured; colour-quenching was corrected for by the channels-ratio method. After 34 days the pH of the suspensions was measured again and c_L and c_L^* were determined. In the experiments on the pH-influence the pH was adjusted three times a week by the addition of dilute solutions of NaOH or HCl.

RESULTS AND DISCUSSION

Preliminary experiments showed that phosphate was released slowly into the liquid phase of the

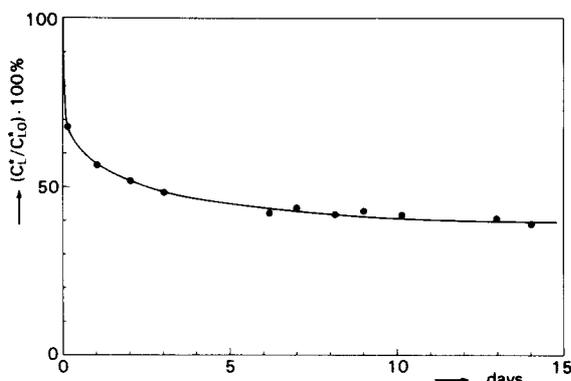


Fig. 1. Typical example of the kinetics of the isotopic exchange of $^{32}\text{PO}_4$ with stable phosphate in sediment samples (sample L2, Table 2 column 4). The ordinate represents the activity of ^{32}P in solution, expressed as a percentage of the amount of radiotracer originally added to the solution.

sample suspensions. Only after 2–3 weeks of continuous agitation did the suspensions reach a state of constant phosphate concentration in the solution (c_L).

As illustrated in Fig. 1, the subsequent addition of $^{32}\text{PO}_4$ (without disturbing the volume of the system or the mass of $^{31}\text{PO}_4$) resulted in an initially rapid uptake of the radiotracer after about 7 days, followed by a slow decrease in the tracer concentration in the solution (c_L^*). In all experiments isotopic equilibrium had not been reached within 14 days.

In order to estimate the final c_L^* we used the approach of Ryden *et al.* (1977) by plotting c_L^* vs the reciprocal value of the exchange-time and extrapolated to $t^{-1} = 0$. This was done by means of linear regression using values obtained after 7, 9, 10 and 14 days (Fig. 2). These t^{-1} plots do not imply more than being a method to obtain extrapolated values that can be compared. The time scale is still one in weeks, which is appropriate for the rate of isotopic exchange that is of interest for the purpose of this study. The extrapolated values never deviated by more than 15% from the last measured values of c_L^* . It was further observed that the pH variations were small

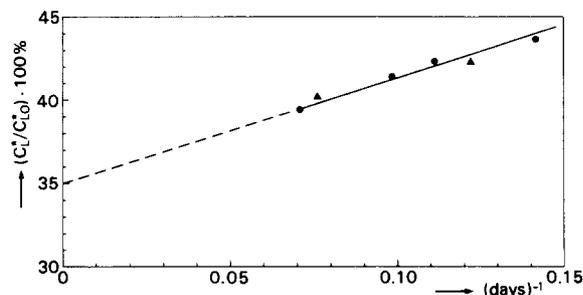


Fig. 2. Example of the extrapolation method proposed by Ryden *et al.* (1977) applied on data points shown in Fig. 1. The dots indicate the data for the time intervals used in the standard procedure as described in the text (7, 9, 10 and 14 days), the triangles indicate data obtained for two intermediate measurements (8 and 13 days).

Table 2. Isotopically exchangeable phosphate in sample M1

	No treatment (n = 5)	Addition of CH ₂ O (n = 5)	u.v.-irradiated (n = 8)
pH	7.51 ± 0.06	7.66 ± 0.03	7.33 ± 0.05
G/V (g l ⁻¹)	0.899	0.887	0.418
c _L (mg P l ⁻¹)	0.80 ± 0.01	0.82 ± 0.01	0.60 ± 0.01
c _S (mg P kg ⁻¹)	4510 ± 10	4470 ± 10	3960 ± 30
(c _{SO} - c _S) (mg P kg ⁻¹)	890 ± 10	930 ± 10	1440 ± 30
f	0.41 ± 0.02	0.54 ± 0.01	0.36 ± 0.01
E	0.28 ± 0.02	0.18 ± 0.01	0.66 ± 0.03
E c _S (mg P kg ⁻¹)	1300 ± 100	790 ± 40	2600 ± 100
E _i	0.40 ± 0.02	0.32 ± 0.01	0.75 ± 0.02
E _i c _{SO} (mg P kg ⁻¹)	2200 ± 100	1720 ± 40	4100 ± 100

(SD < 0.25). Therefore no attempt was made to control the pH of the sample suspensions.

Effect of irradiation with u.v.-light and of the addition of formaldehyde

Table 2 shows the isotopic exchangeability of phosphate in sample M1, in the same sample after irradiation with u.v.-light and in the presence 0.17 M CH₂O.

In order to get an impression of the reproducibility, the experiments were repeated 5–8 times; the table gives the mean values and the standard deviations. Since the results of the analysis for isotopically exchangeable phosphate are reproducible, it is concluded that the method is satisfactory. It is reasonable to assume that in the experiments with CH₂O or in the experiments using u.v.-irradiated material, no biological activities took place (Pomeroy *et al.*, 1965; Jackson and Schindler, 1975). The effect of u.v.-irradiation is less general but most micro-organisms are inactivated (Harm, 1980). Furthermore quite a number of organic phosphates are known to be mineralized to inorganic phosphate by irradiation with u.v.-light (Armstrong *et al.*, 1966, 1968).

The differences between the results for sample M1 in the presence and absence of CH₂O is at least partly due to bioactivities. The absence of CH₂O leads to a slightly higher isotopic exchangeability. One can argue that coating of inorganic particles by CH₂O might have a similar effect, but it will be shown for experiments with 26 sediment samples that this, if at all, is not the main reason for the observed differences. Because in both cases the values of c_S are

about equal, the increased total isotopic exchangeability E_i in the absence of CH₂O is, according to equation (4), merely the result of the increased E value.

Comparison of the results obtained in the presence of CH₂O with those for the u.v.-irradiated sample M1 shows a spectacular rise of 2400 mg P kg⁻¹ in E_i c_{SO} [equation (4)] upon irradiation. About 500 mg P kg⁻¹ of this rise can be explained by the influence of the lower G/V-ratio in the u.v.-irradiated experiment (cf. pertinent paragraph), which results in an increased value for (c_{SO} - c_S). After irradiation with u.v.-light a reduction of 42 g C kg⁻¹ in the organic carbon content was found in sample M1, using the method of Sims and Haby (1971). On the basis of a C/P-weight ratio of 40:1 (using an overall formula of C₁₀₆H₂₆₃O₁₁₀N₁₆P for algal protoplasm) the estimated u.v.-mineralization of biologically/organically bound PO₄ is then 1000 mg P kg⁻¹ at most. So the presence of biologically or organically bound phosphate can only explain part of the rise in E_ic_{SO} [Equation (4)]. Possibly one of the extra effects of u.v.-irradiation is the activation of inorganic phosphates (sample M1: 900 mg P kg⁻¹ or more) that do not normally participate in isotopic exchange reactions.

Two explanations for this behaviour are proposed. Firstly: an attenuation of the bonds of (chemi)sorbed phosphate; secondly: removal of adsorbed organic compounds that form a protective coating on phosphate groups at the surface of solid phases.

When the results of Tables 1 and 2 are compared, it is clear that not all phosphate that is supposedly bound in or adsorbed on aluminium or iron oxides or

Table 3. Effect of the pH on the E- or E_i-value of phosphate in sediment L2

	6.57 ± 0.10	7.25 ± 0.12	7.64 ± 0.09	7.96 ± 0.19	8.41 ± 0.18
pH	6.57 ± 0.10	7.25 ± 0.12	7.64 ± 0.09	7.96 ± 0.19	8.41 ± 0.18
I (mol l ⁻¹)	0.0123	0.0114	0.0105	0.0101	0.0100
G/V (g l ⁻¹)	1.105	1.108	1.114	1.119	1.115
c _L (mg P l ⁻¹)	0.363	0.412	0.472	0.577	0.760
c _S (mg P kg ⁻¹)	2970	2930	2880	2780	2620
(c _{SO} - c _S) (mg P kg ⁻¹)	330	370	420	520	680
f	0.14 ± 0.04	0.21 ± 0.01	0.29 ± 0.01	0.35 ± 0.01	0.49 ± 0.01
E	0.69 ± 0.21	0.47 ± 0.01	0.36 ± 0.01	0.31 ± 0.01	0.27 ± 0.01
E c _S (mg P kg ⁻¹)	2040 ± 640	1390 ± 20	1030 ± 30	860 ± 20	710 ± 40
E _i	0.72 ± 0.19	0.53 ± 0.01	0.44 ± 0.01	0.42 ± 0.01	0.42 ± 0.01
E _i c _{SO} (mg P kg ⁻¹)	2370 ± 630	1760 ± 20	1450 ± 20	1380 ± 20	1400 ± 30
F(H ₂ PO ₄ ⁻)	0.69 ± 0.04	0.32 ± 0.06	0.16 ± 0.03	0.08 ± 0.03	0.03 ± 0.01
F(HPO ₄ ²⁻)	0.31 ± 0.05	0.68 ± 0.06	0.84 ± 0.03	0.92 ± 0.03	0.97 ± 0.01

oxihydroxides present in sample M1 is isotopically exchangeable (~40%).

Effect of the pH on isotopically exchangeable phosphate

Table 3 shows the effect of the pH on the isotopic exchangeability of phosphate in sediment L2 in the presence of 0.17 M CH₂O. These experiments were carried out in duplicate at 5 pH-values ranging from 6.6 to 8.4. The mean values and standard deviations are given in the table. The agreement between the duplicates was good except at pH 6.6 where large differences were observed. An unavoidable result of the pH-corrections, especially in the experiments at low pH-value, was the rise in ionic strength. The change in ionic strength was calculated from the quantities of added HCl or NaOH. It was assumed that only Na⁺ and Cl⁻ contributed to this increase. In interpreting the results we assumed that the influence of the rise in ionic strength on the isotopic exchangeability could be neglected.

The results show clearly a decrease of $E c_s$ (equation (4)) with increasing pH-value. At higher pH-values this pH-effect is masked in the total exchangeability E_t , because the drop in $E c_s$ [equation (4)] is compensated for by the rise in $(c_{s0} - c_s)$, which is due to partial desorption/dissolution of phosphate at the higher pH-values. The fact that, in the investigated pH-range, E and t are high at low pH-values suggests that H₂PO₄⁻ is more easily exchanged than HPO₄²⁻.

To investigate this suggestion we calculated the species distribution of H₃PO₄ at the pH-values of Table 3. In this calculation we used $pK_1 = 2.15$, $pK_2 = 7.06$ and $pK_3 = 12.07$ as equilibrium constants for the dissociation steps of H₃PO₄ at ionic strength 10⁻² M (Stumm and Morgan, 1981) and as activity constants we took values which had been estimated with the Davies equation (Stumm and Morgan, 1981). From the dissociation constants of H₃PO₄, given by Stumm and Morgan (1981) it turns out (cf. Table 3) that in the pH-range of the experiments only H₂PO₄⁻ and HPO₄²⁻ are important. Although not measured, the Ca and Mg concentration eventually generated by partial dissolution of the suspended solids, are assumed to be low enough to justify the neglect of complexing of the monohydrogen phosphate anion in Ca and Mg complexes (log K respectively 2.7 and 2.5; Stumm and Morgan, 1981).

Without the use of the less precise measurements at pH 6.6 we found the following relationships between the exchangeabilities and the fraction of the dihydrogenphosphate ion

$$\alpha_{H_2PO_4^-} = (H_2PO_4^-)/(PO_4)_T$$

by means of linear regression:

$$E = 0.25 + 0.68 \alpha_{H_2PO_4^-} \quad (r = 1.00) \quad (5)$$

$$E = 0.39 + 0.41 \alpha_{H_2PO_4^-} \quad (r = 0.96). \quad (6)$$

The high, positive correlation coefficients sustain the suggestion that H₂PO₄⁻ is the species which is easily exchanged. Atkinson *et al.* (1972) discussed the influence of pH on the rate of isotopic exchange of phosphate at the goethite-aqueous solution interface. They explained the increase of this rate with decreasing pH (pH < 9) by catalysis involving protonation of one oxygen in the Fe_s-O-P-O-Fe_s bonds (s stands for surface sites) leading to a single bonded Fe_s-OPO(OH)₂ surface complex. *Mutatis mutandis* this is essentially quite similar to the explanation as given by us.

In the pH-region 7.6–8.4 the influence of the pH on E or E_t is not large. Therefore, as long as one is dealing with small differences in this pH-range, exchangeabilities of different sediments can be compared. Next to the results of Atkinson *et al.* (1972), little is known from literature about the pH-effect on isotopic exchangeability. Murmann and Peech (1969) reported the opposite effect for 53 different soils in 10⁻² M CaCl₂ as ionic medium. The precipitation of Ca-PO₄-compounds may have been responsible for this. White and Taylor (1977) found practically no pH-effect between pH 5.2 and 6.9 with a clay loam suspended in 2 × 10⁻³ M CaCl₂.

Comparison of Tables 1 and 3 shows that at the same pH as used for sample M1 (Table 2), the isotopically exchangeable phosphate is about 60% of the amount of phosphate, that is mainly present in/at aluminium or iron oxide phases.

Effect of the solid/solution-ratio on isotopically exchangeable phosphate

Table 4 shows the influence of the solid/solution ratio on the E - and E_t -values of sediment L2 in the presence of 0.17 M CH₂O. It is obvious that the relative errors in E and $E c_s$ must be large at low G/V -values due to the high f -values and the resulting uncertainty in the $(1 - f)$ -term [see equation (2)]. The

Table 4. Effect of G/V ratio on the E - or E_t -value of phosphate in sediment L2

G/V (g l ⁻¹)	0.0936	0.187	0.375	0.749	1.50	3.00
pH	7.66 ± 0.05	8.01 ± 0.18	8.16 ± 0.09	8.03 ± 0.05	7.93 ± 0.02	7.86 ± 0.06
c_L (mg P l ⁻¹)	0.227	0.426	0.642	0.662	0.668	0.705
c_s (mg P kg ⁻¹)	880	1030	1590	2420	2850	3070
$c_{s0} - c_s$ (mg P kg ⁻¹)	2420	2270	1710	880	450	230
f	0.91	0.86	0.75	0.54	0.35	0.23
E	0.27 ± 0.09	0.36 ± 0.05	0.36 ± 0.05	0.31 ± 0.02	0.29 ± 0.02	0.26 ± 0.01
$E c_s$ (mg P kg ⁻¹)	240 ± 80	370 ± 80	580 ± 70	760 ± 60	830 ± 50	800 ± 40
E_t	0.81 ± 0.04	0.80 ± 0.04	0.70 ± 0.05	0.50 ± 0.02	0.39 ± 0.02	0.31 ± 0.02
$E_t c_{s0}$ (mg P kg ⁻¹)	2670 ± 130	2640 ± 130	2290 ± 120	1640 ± 80	1270 ± 60	1030 ± 50

Table 5. Influence of the concentration of formaldehyde on isotopically exchangeable phosphate in the oxidized zone of sediment core A4

CH ₂ O (mol l ⁻¹)	0	0	0.003	0.005	0.011	0.021	0.043	0.085	0.170	0.170
pH	8.27	8.34	7.58	7.22	8.63	8.60	8.62	8.42	8.40	8.39
G/V (g l ⁻¹)	1.34	0.994	0.625	0.625	0.625	0.625	0.625	0.625	1.16	1.33
c _L (mg P kg ⁻¹)	0.206	0.225	0.158	0.143	0.251	0.245	0.238	0.216	0.239	0.214
f	0.065	0.029	0.12	0.064	0.24	0.23	0.23	0.21	0.11	0.12
E _i	1.12	3.68	1.00	1.71	0.79	0.82	0.78	0.77	0.88	0.66

standard deviations in the calculated isotopic exchangeabilities were estimated from the relative errors of 3, 2½ and 3% in the determination of c_L, G/V and f respectively. The influence of the pH on the results was neglected since the differences in the pH-value were small.

The G/V-ratio has practically no effect on the E_i-value. However, the total exchangeability E_i is strongly increased at low G/V-ratio. This is caused by the influence of G/V on (c_{SO} - c_S). At the lowest value of G/V, about 70% of the total phosphate content, c_{SO}G is dissolved/desorbed but this percentage continuously drops to about 7% at the highest value of G/V. Theoretically E_i approaches unity as G/V approaches zero because at extremely low G/V-values c_LV = c_{SO}G and f = 1, so E_i = 1 [equation (3)]. The experiments of this investigation seem to agree with this boundary condition.

De Bussetti *et al.* (1977), using 0.1–1.4 g l⁻¹ aqueous suspensions of model compounds like hydroxyapatite and amorphous AlPO₄, found the same reduction in E_i with increasing solid/solution-ratio. These results deviate from the results of Bowman and Olsen (1979) obtained with a silty clay loam suspended in water at pH-values between 7.9 and 8.4. They found that the G/V-ratio had no effect on E_i at high G/V-values (50–400 g l⁻¹) probably because the relative contribution of (c_{SO} - c_S) in E_ic_{SO} [equation (4)] is much smaller there. The isotopic exchangeability seems to be complete (cf. Tables 1 and 4) at the lowest values of the solid/solution-ratio but decreases to about 45% at the highest value.

It is concluded that it is important to work at nearly constant solid/solution-ratio if one's objective is to compare the isotopic exchangeabilities of different soils or sediments.

Application to sediments from the Rhine/Meuse delta

As was already mentioned under "experimental procedures", irreproducible and irrational results were sometimes obtained that allegedly were caused by biological activity. Therefore, one of the samples collected in the area was used to study the effect of the concentration of CH₂O between 0.0 and 0.17 M. The results are shown in Table 5. All but two pH-values fall within a small range. The reason for these two lower pH-values is not understood. However, their influence and the effect of the differences in solid/solution-ratio cannot invalidate the conclusion that, with this particular sediment, at CH₂O concentrations below 10⁻² M irrational and irre-

producibile results are obtained. Therefore, in accordance with the experimental conditions used by Pomeroy *et al.* (1965), a concentration 20 times as strong, namely 0.17 M CH₂O, was used in the routine experiments.

Table 6 shows the total isotopic exchangeability E_i of phosphate in the 26 sediment samples (18 sediment cores) investigated. The table gives the results of the duplicate experiments in the presence and absence of 0.17 M CH₂O. In no case was equilibrium in the isotopic exchange established within 14 days of tracer addition and therefore the value of f was estimated using the extrapolation method (Fig. 2).

In order to indicate the differences in the solid/solution-ratio and in the pH, we give in Table 6 the mean values and standard deviations of these parameters. These values are based on the four experiments with each separate sediment. For all 104 experiments together the mean solid/solution-ratio is 1.09 ± 0.09 g l⁻¹ and the mean pH is 8.10 ± 0.25. In a foregoing paragraph it was concluded that the influence of G/V and of the pH on the E_i-value can be neglected if the differences in G/V and pH are small. It appears that these conditions are fulfilled and therefore the E_i-values of the various sediments can be compared directly. In the group of sediment cores A1–A6 many irreproducible and irrational (E_i < 1) exchangeabilities were found in the absence of CH₂O. The corresponding experiments are characterized by a relatively strong reduction in the fraction of the radiotracer remaining in solution in the period 7–14 days after tracer addition. Apparently one is dealing with a situation that is well out of equilibrium. Only a small fraction of the total phosphate, c_{SO}, is yielded to the solution, giving rise to a phosphate concentration, c_L. To this solution radiotracer is added. When growth of the biological phase occurs at the expense of the dissolved and most labile phosphate, containing most of the labeled phosphate, it can be understood that this excess uptake of the radiotracer leads to an artificially high value of exchangeable phosphate. As a result the f-value found by extrapolation is unreliable. These problems are eliminated by the addition of CH₂O, which suggests that the deviations are caused by phosphate consumption by microorganisms attached to the sediment. The phosphate that is consumed must be supplied by the sediment, because the phosphate concentration in the solution remains constant. In this situation of phosphate transport the steady state condition is not fulfilled, which implies that

Table 6. Total isotopic exchangeability of phosphate in 26 freshwater sediments from the Rhine/Meuse-estuary

Core	Origin*	Redox state	c_{50}^{SO} (mg P kg ⁻¹)	pH	G/V (g l ⁻¹)	E_i			
						No CH ₂ O		+ CH ₂ O	
						1	2	3	4
A1	G.R.	Ox.	2400	8.26 ± 0.07	1.08 ± 0.26	0.73	1.10	0.60	0.55
A1	G.R.	Red.	1450	8.42 ± 0.11	1.13 ± 0.15	0.77	0.75	0.58	0.53
A2	G.R.	Ox.	2200	8.16 ± 0.18	1.14 ± 0.14	1.48	0.82	0.63	0.55
A2	G.R.	Red.	1500	8.23 ± 0.12	1.16 ± 0.15	1.94	0.65	0.52	0.51
A3	G.R.	Ox.	2050	8.17 ± 0.22	1.09 ± 0.17	1.50	0.67	0.65	0.58
A3	G.R.	Red.	1500	8.30 ± 0.13	1.11 ± 0.13	3.64	0.97	0.60	0.56
A4	G.R.	Ox.	2100	8.35 ± 0.06	1.21 ± 0.17	1.12	3.68	0.88	0.66
A4	G.R.	Red.	1100	8.48 ± 0.07	1.16 ± 0.15	0.74	0.94	0.62	0.54
A5	G.R.	Ox.	1600	8.26 ± 0.16	1.20 ± 0.11	0.96	1.97	0.78	0.70
A5	G.R.	Red.	1450	8.33 ± 0.16	1.11 ± 0.13	0.74	2.20	0.67	0.51
A6	G.R.	Ox.	2100	8.19 ± 0.19	1.01 ± 0.21	0.95	1.08	0.73	0.65
A6	G.R.	Red.	1500	8.27 ± 0.12	1.18 ± 0.14	0.44	0.66	0.51	0.50
A7	H.	Red.	3800	7.78 ± 0.11	1.16 ± 0.02	0.28	0.16	0.23	0.21
A8	H.	Red.	3000	7.90 ± 0.07	0.89 ± 0.02	0.19	0.22	0.23	0.17
A9	H.	Red.	2700	7.84 ± 0.13	1.13 ± 0.02	0.24	0.20	0.24	0.18
A9	H.	Red.2	2900	7.88 ± 0.15	1.14 ± 0.02	0.27	0.26	0.27	0.14
A10	H.	Red.	4000	7.71 ± 0.14	0.93 ± 0.02	0.38	0.28	0.28	0.17
A10	H.	Red.2	4200	7.64 ± 0.15	1.03 ± 0.02	0.29	0.25	0.27	0.20
L1	B.M.	Ox.	2300	7.94 ± 0.12	0.91 ± 0.02	0.74	0.59	0.72	0.56
L2	B.M.	Ox.	3300	7.91 ± 0.15	1.08 ± 0.02	0.72	0.55	0.56	0.50
L3	B.M.	Red.	1950	8.13 ± 0.08	1.08 ± 0.02	0.70	0.52	0.56	0.33
L4	B.M.	Ox.	2800	7.85 ± 0.13	1.09 ± 0.02	0.77	0.64	0.59	0.55
L5	B.M.	Ox.	2700	7.88 ± 0.11	1.05 ± 0.02	0.81	0.56	0.61	0.54
L6	G.R.	—	2400	8.36 ± 0.11	1.07 ± 0.02	0.62	0.53	0.53	0.56
L7	G.R.	—	2600	8.48 ± 0.06	0.93 ± 0.02	0.62	0.60	0.60	0.56
L8	G.R.	—	2000	7.93 ± 0.25	1.18 ± 0.02	0.86	0.55	0.49	0.39

*G.R. = Grote Rug, H. = Haringvliet, B.M. = Brielse Meer.

equation (1) does not hold and that the E_i -value has been overestimated.

Inspection of the results from the last four columns in Table 6 gives little support to an alternative explanation of the effect of CH₂O on E_i , that is that CH₂O decreases the exchangeability of phosphate in a purely inorganic system. For instance, no suppression of exchangeability is noticed in cores A7–A10. We are therefore confident that CH₂O inhibits bioactivity without complicating side-effects.

It is confirmed once more that in analysing for isotopically exchangeable phosphate one must add a biological inhibitor in order to prevent interference from bioactivity.

Columns 3 and 4 in Table 6 show that the results for the duplicate experiments in the presence of 0.17 M CH₂O are not quite the same. The values for E_i in column 4 are on average systematically 0.08 lower than those in column 3. The reason is that the experiments were not carried out under the same state of agitation of the suspensions. Much better agreement between duplicates is obtained when the experimental conditions are identical (cf. foregoing paragraphs).

Comparison of the total exchangeabilities of phosphate in the presence of CH₂O (Table 6) shows that the sediments collected from the Haringvliet have the lowest E_i values (about 15–25%) whereas high E_i values (about 40–80%) are found in the sediments from the Brielse Meer and the Grote Rug. Consequently the bioavailability of phosphate in the sediments collected from the Brielse Meer and the

Grote Rug can be assumed to be higher than in the sediments from the Haringvliet.

In the group of sediment cores A1–A6 and E_i -value of the reduced zone is, in the presence of CH₂O, systematically lower than the E_i -value of the oxidized zone in the same core. This indicates that phosphate in the reduced zone is fixed in a different manner. Apparently phosphate adsorbed on iron oxy-hydroxide phases in the oxidized zone takes part in the isotopic exchange.

The high total isotopic exchangeabilities of phosphate found in most of the sediments of this study agree well with the results of Li *et al.* (1972, 1973) which were also obtained with freshwater sediments.

CONCLUSION

In order to get useful results on the isotopic exchangeability of phosphate present in different phases in sediments or soils, it is necessary to carry out the experiments under well defined conditions of pH and solid/solution-ratio. A biological inhibitor like formaldehyde must be added to prevent interference from bioactivity. Only then one can compare the results for different samples and correlate them with results obtained in various experiments on bioavailability of phosphate in these samples.

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