

Chapter 2

General Introduction

1. Introduction

Bone is a remarkable tissue with diverse functions. In close collaboration with ligaments, muscles, and tendons the skeleton literally forms the backbone of locomotion. In addition, the skeleton protects vital organs, including bone marrow, brain, spinal cord, heart, and lungs. And last but not least, bone has a metabolic function, acting as a reservoir of ions, especially calcium and phosphate, for the maintenance of serum homeostasis. Bone is a highly specialized connective tissue. The skeletal system consists of bone and cartilage. The rigidity of bone tissue relies on mineralization of the bone matrix with hydroxyapatite crystals, which are predominantly composed of $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$. There are two major families of bone cells, the osteoblast and the osteoclast lineages.^{179,180} Osteoblasts and osteoclasts are responsible for the dynamic turnover of bone both during growth and in adult life. The osteoblast is the bone-lining cell responsible for the production of bone matrix, whereas the osteoclast is a multinucleated bone-lining cell responsible for bone resorption. During bone formation osteoblasts are embedded within the bone matrix in small lacunae to form osteocytes (Fig 1).

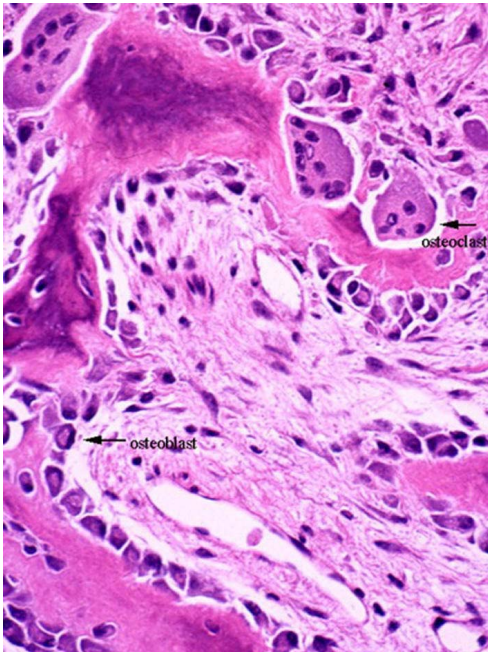


Fig 1. Osteoblasts are present on the lower left, lining a bone trabecula. Osteocytes can be seen embedded within the bone matrix. Osteoclasts are present at the right top corner.

Bone formation occurs through the coordinated production and mineralization of the osteoid matrix. Bone matrix components consist of three classes of macromolecules. The first group possesses repetitive structural motifs and includes collagen, hyaluronan, decorin and biglycan. The second group is characterized by a modular domain structure and includes versican, thrombospondin, fibronectin, osteonectin, and tenascin. The third group demonstrates no clear structural motifs and includes matrix γ -carboxylated protein, bone sialoprotein, osteopontin, and osteocalcin.¹³⁹ The complexity of the interactions between these various matrix components and bone cells conveys to bone its unique function of sustaining a stable yet dynamic structure. The major organic component of the extracellular bone matrix is type I collagen, accounting for up to 90% of the organic matter. Collagen I is composed of trimers of two $\alpha 1$ chains and one $\alpha 2$ chain, to form triple helical molecules. The collagen α chains are produced as procollagen which possesses amino- and carboxyl-terminal polypeptide extensions. Enzymatic removal of the noncollagenous N- and C-terminal extensions precedes collagen fibril formation. Collagen III is also found in bone matrix, but its role in bone metabolism is unclear.

2. Bone histology and histogenesis

On gross observation of bone, cancellous and compact bone can be distinguished. Cancellous bone consists of many trabecular walls separating numerous interconnecting cavities filled with bone marrow or fat tissue. Compact bone does not show these cavities. Nevertheless, on microscopic examination both cancellous and compact bone have the same basic histologic structure. In long bones, compact bone is mainly found in the diaphysis whereas cancellous bone is located in the metaphyseal areas and the epiphysis, surrounded by a thin layer of compact bone.¹⁹² Flat bones of the skull, scapula, and pelvis usually have a core of cancellous bone flanked by two plates of compact bone. On histology, there are two varieties of bone tissue: primary, immature or woven bone; and secondary, mature or lamellar bone. The difference between the two relies on the fact that collagen bundles are placed randomly in the first variety and organized into bone lamellae in the second.

Primary bone is the first bone tissue formed during bone formation. It is temporary and is readily replaced by secondary bone tissue in most places in the skeleton. In addition to the irregular deposition of collagen bundles, primary bone is characterized by a lower amount of mineralization and a larger amount of osteocytes compared to secondary bone.

Secondary bone is the variety mainly found in the mature skeleton. The bone lamellae, which characterize mature bone, are arranged parallel to each other or concentrically surrounding a central canal containing blood vessels and nerves. This complex of concentric lamellae is called the osteon or Haversian system and is the main building block of the skeleton (Fig 2). Between, and occasionally within the lamellae, lacunae containing osteocytes are encountered. These osteocytes have numerous long cell processes, which are in contact with other osteocytes and osteoblasts. Osteocytes are still capable of matrix production. The osteons communicate with each other, the endosteum, the marrow cavity, and the periosteum through transverse or oblique Volkmann's or perforating canals (Fig 2). These Volkmann's canals have no concentric lamellae and perforate the lamellae of the osteons. During growth and also in the adult skeleton continuous remodeling of osteons takes place. This explains the great variability in size and form of osteons.

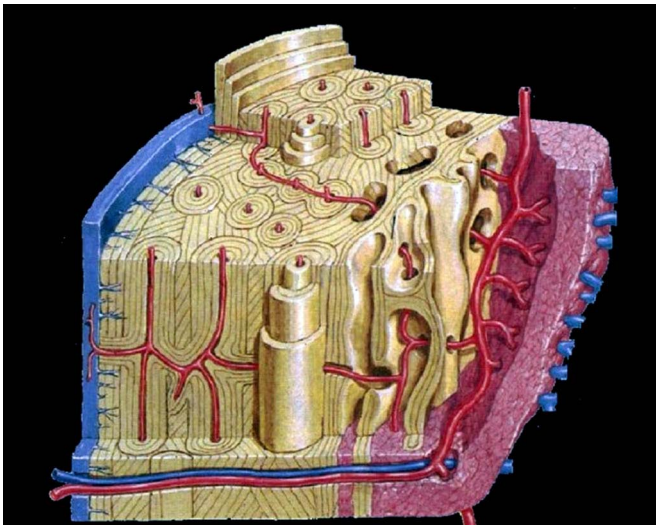


Fig 2. Representation of bone histology demonstrating the main building unit of bone i.e., the osteon.

Bone tissue is formed either by intramembranous ossification or by endochondral ossification.⁸⁷ Intramembranous ossification is characterized by direct bone formation within a layer or membrane of connective tissue i.e., periosteum. During endochondral ossification, a cartilaginous model precedes the actual accretion of bone tissue. In both intramembranous and endochondral ossification bone tissue is deposited first as primary or immature bone. Through

remodeling this primary bone is replaced by mature lamellar bone. Most of the bones of the skull, including parietal bones, frontal bones, mandible, and maxilla are formed by intramembranous ossification. Growth in width of short and long bones proceeds through intramembranous bone formation. Primary ossification starts within the connective tissue layers. Preosteoblasts differentiate into osteoblasts, which start producing osteoid.¹⁷⁹ Osteoid is mineralized in turn to form primary bone. The trabeculas of primary bone unite to create cancellous bone. Growing blood vessels and undifferentiated mesenchymal cells, which give rise to bone marrow cells, including preosteoclasts, penetrate the cancellous bone.

Endochondral ossification is responsible for the formation of most short and long bones. Endochondral ossification relies on the replacement of a hyaline cartilage and basically depends on two processes. The first process is hypertrophy and apoptosis of chondrocytes of the model of bone leaving expanded lacunas separated by septa of calcified cartilage matrix. In the second process, an osteogenic bud consisting of blood capillaries and osteogenic precursor cells penetrates into the lacunas left by the apoptotic chondrocytes. The undifferentiated cells give rise to osteoblasts, which lay down osteoid on the remnants of the calcified cartilage matrix. In this way, bone tissue appears at the site where there was cartilage.

Long bones are formed from cartilaginous models with a cylindrical shaft or diaphysis and enlarged extremities or epiphyses. The first bone to develop in the diaphysis is the bone collar, which surrounds the periphery of the cartilaginous matrix and thus forms the shaft of the bone. This bone collar is produced through intramembranous ossification. Within the forming bone collar, chondrocytes of the cartilage model start the process of hypertrophy, apoptosis, lacuna formation, and mineralization of the remaining cartilage matrix, also known as hypertrophication. Blood vessels of the osteogenic bud invade the lacuna and osteoblasts start to synthesize bone matrix. This ossification center, which appears in the diaphysis, is called the primary ossification center. At later stages of development a secondary ossification center arises at the end of the long bone to form an epiphysis or apophysis. Instead of the longitudinal growth of the primary ossification center, growth in the secondary center is radial. As bone formation in the primary and secondary ossification centers progresses, an epiphyseal plate or growth plate is formed between the diaphysis and epiphysis.⁸⁹ By analogy an apophyseal plate is formed between the diaphysis and apophysis. During adolescence, longitudinal bone growth continues in the growth plate through a highly coordinated type of endochondral ossification.²¹

In the epiphyseal plate five zones can be distinguished. Starting from the epiphyseal side these are: 1- the resting zone containing hyaline cartilage and small chondrocytes; 2- the proliferative zone with rapidly dividing chondrocytes, which

form columns of stacked cells parallel to the long axis on the bone; 3- the hypertrophic cartilage zone with enlarged chondrocytes and interspersed thin septa of resorbed cartilaginous matrix; 4- the calcified cartilage zone with chondrocyte apoptosis. The cartilaginous matrix septa are mineralized with hydroxyapatite; 5- the ossification zone in which primary bone is formed. Blood vessels and osteoblasts invade the calcified cartilage matrix and deposit osteoid on the septa. The osteoid is mineralized, thus forming primary bone tissue. Longitudinal bone growth relies on the continuous cell division of chondrocytes in the proliferative zone and bone accretion at the metaphyseal side (Fig 3).

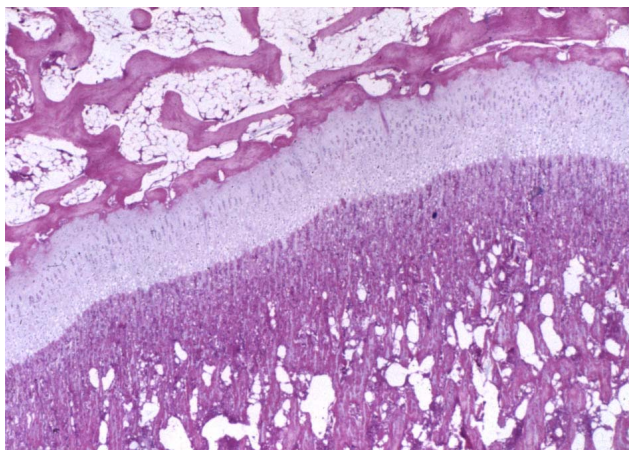


Fig 3. Microscopic section of the growth plate demonstrating endochondral bone formation. Courtesy of the Department of Pathobiology, Division Pathology, Faculty of Veterinary Medicine, Utrecht University.

3. Growth plate injuries.

Injuries of the growth plate can occur as long as the epiphyseal plate is not replaced by bone and usually occurs before the long bones have reached their full growth potential. Longitudinal bone growth in the dog ceases at approximately 10 months of age and is characterized by closure of the epiphyseal plates.³⁴ Growth plate fractures are classified according to the Salter-Harris system into 5 categories: type I, fracture through the growth plate; type II, fracture through the growth plate and metaphysis; type III, fracture through the growth plate and epiphysis; type IV, fracture through the growth plate, epiphysis and metaphysis, and type V, crush or compression injury of the growth plate.^{21,145} The Salter-Harris system was designed to predict the prognosis of growth plate injuries. Based on clinical and experimental physeal fractures they postulated that most type I and type II fractures were restricted to the zone of hypertrophic chondrocytes and thus should not

seriously affect longitudinal growth after careful reduction and stabilization. In dogs, the prognosis of these types of physal fractures was proven to be less favorable. Disruption of the cells of the proliferative zone was found in the majority of traumatic growth plate injuries, which accounts for the high incidence of growth retardation in these patients.^{59,89} In addition to decreased longitudinal growth, malformation of the limb is a common finding.

In dogs, growth deformity of the antebrachium is the most common limb malformation.⁸⁸ The antebrachium consists of the paired bones radius and ulna. Proximally these bones articulate with the humerus to form the elbow joint. The distal radius and ulna contribute to the antebrachio-carpal joint. During active growth the radius has a proximal and distal epiphyseal plate. The proximal growth plate contributes approximately 35%, whereas the distal plate contributes 75% to the total length of the radius.³⁴ The ulna has a proximal apophyseal growth plate, which accommodates for longitudinal growth of the olecranon. The ulna has only one distal epiphyseal growth plate, which is responsible for the entire longitudinal development of the ulnar diaphysis. This distal ulnar growth plate is shaped like an inverted cone to enlarge the proliferative zone area and thus the number of chondrocytes contributing to longitudinal growth. This adaptation in shape allows for synchronous longitudinal growth between radius and ulna. In short-legged dogs, this inverted cone shape is less obvious as growth rate in the radius and ulna is much slower.

By its high growth rate and configuration the distal ulnar growth plate is reported to be more vulnerable to trauma than the radial physes.^{60,135} The typical presentation of growth retardation or premature closure of the distal ulnar growth plate is the radius curvus syndrome. An ulnar length deficit, cranial bowing of the radius, exorotation of the antebrachium, and valgus deviation of the distal limb in the carpus characterize this syndrome.^{25,62,63} In the clinical situation, isolated disturbance of one physis is not common and usually both the distal radial and distal ulnar growth plates are involved.^{128,140,183} The severity and localization of the growth disturbance within the radial and ulnar physes will vary, resulting in a heterogeneous presentation of the growth deformities. In addition to the growth deformity, asynchronous development of the radius and ulna can lead to incongruity of the elbow joint.^{102,105,106,112} Asynchronous radial and ulnar growth can also result in carpal malalignment and subluxation.^{61,117,122,134}

In summary, antebrachial growth deformities are characterized by a combination of length deficits, angular and rotational malalignment, elbow incongruity, and carpal subluxation. Osteoarthritis of the elbow and carpal joint is a common sequel of radial and ulnar growth deformities.

4. Healing of bone fractures

Bone fractures are characterized by discontinuity of bone architecture and loss of function. Bone healing is the reparative process by which bone regains its function. Two different mechanisms of bone repair can be distinguished, i.e., direct and indirect bone healing. Direct bone union is characterized by direct osteonal reconstruction, whereas indirect bone healing depends on the formation of an intermediate fibrous and cartilaginous callus.

Direct bone healing is achieved by internal remodeling of the Haversian systems without resorption of the fracture surfaces and intramembranous ossification. This type of healing is also known as contact healing and occurs with stable fixation and compression of the fracture surfaces. Under these conditions blood vessels can cross the fracture line followed by osteoclasts and osteoblasts, thus forming new Haversian systems. In small stable gaps of up to 0.2 mm in width direct union can also occur by direct deposition of lamellar bone. In larger stable gaps of up to 0.8 mm in width direct healing can proceed by the formation of cancellous bone.⁴¹

Indirect bone healing is characterized by the formation of an intermediate callus. This callus consists of fibrous and cartilaginous tissue. After bone fracture the sequence of events leading to bone healing can be described as hemorrhage in the fracture area, clot formation, inflammatory response, angiogenesis, proliferation of pluripotential mesenchymal cells, fibrous and cartilaginous callus formation, bone formation, and bone remodeling.¹⁹² This sequence of events results in a gradual progressive stabilization of the fracture area with increasingly stronger and stiffer tissues. Callus formation can be subdivided on the basis of location into periosteal callus, intercortical callus, and medullary callus. The periphery of the callus consists of fibrous tissue, which encloses the more centrally located cartilaginous tissue of the callus. Bone formation in the callus proceeds from the periphery of the cartilaginous callus to the central area of the fracture zone. During this process of endochondral bone formation, cancellous bone is produced until the fracture gap is bridged. This cancellous bone is replaced by lamellar bone during remodeling, which can take up to several years. The extent of callus formation depends on several factors, including stability in the fracture area, age, and local blood supply. Increasing instability tends to result in a larger amount of callus formation. In contrast, excessive motion within the fracture zone will result in compromised angiogenesis and delayed union. Prolonged fracture union characterizes delayed union. Bone healing is progressive in delayed union and leads to full recovery of function. In nonunion, fracture healing stops altogether and results in either atrophic or hypertrophic nonunion. Atrophic or nonviable nonunion is characterized by resorption of the fracture ends without callus

formation. In hypertrophic or viable nonunion, callus formation is present at the ends of the fracture segments, but no bridging of the fracture gap occurs. Fractures can develop into nonunions by a variety of factors, including insufficient stability, inadequate reduction, interposition of soft tissues, compromised vascularity, and infection. In addition, systemic factors can contribute to the development of nonunion, including old age, hyperadrenocorticism, hypothyroidism, renal disease, osteoporosis, and GH deficiency.⁷⁵

Fracture healing will proceed by a combination of direct and indirect bone union after osteosynthesis in the clinical situation. The cascade of events during fracture healing culminates in the recruitment of bone forming cells. Various angiogenic and osteogenic growth factors interact with pluripotential mesenchymal cells and their respective differentiated cell lineages during progression of fracture healing. These bone growth factors will be discussed in more detail in the next section of this introduction.

5. Distraction osteogenesis

Distraction osteogenesis is the formation of new bone under gradual mechanical distraction of two bone surfaces. Dr. Gavriil Abramovich Ilizarov was the first to develop distraction osteogenesis into a clinical treatment option, using a circular external skeletal fixation system.^{84,85} The technique depends on a minimally invasive osteotomy of the bone while preserving soft tissues, periosteum, endosteum, bone marrow, and intramedullary blood vessels.⁹⁸ Stability of the external fixation is essential to allow new capillary blood vessels to bridge the osteotomy and allow for the lengthening procedure. After a latency period, the duration of which depends on the age of the patient and location of the osteotomy, gradual distraction is started with 1 mm per day, usually divided into 2 to 4 steps. New bone is formed in parallel columns extending from the osteotomy surfaces towards a central growth zone of the distraction gap. The growth zone that forms under the influence of tension-stress has features of both endochondral and intramembranous ossification.^{5,8,9} After lengthening is ceased, the newly formed bone regenerate is allowed to mature and consolidate until the stage that the bone can support its physiologic load.

In humans, distraction osteogenesis has been used in treating a variety of skeletal conditions, including bone length deficits, growth deformities, bone loss after trauma or radical resection, and craniofacial surgery.^{90,113,143,162} In dogs, distraction osteogenesis was introduced in the late 1980s mainly to treat growth deformities of the antebrachium and to a lesser extent of the crus.^{105,106,116,117} In

addition to the clinical use of distraction osteogenesis, the dog has been used extensively as an experimental model.^{4,8,56,57,95,100,118,129,155,186}

Distraction osteogenesis proved to be a successful model to study the role of growth factors during bone formation. Mechanical distraction stress induces the expression of transforming growth factor- β 1 (TGF- β 1), insulin-like growth factor-I (IGF-I), basic fibroblast growth factor (bFGF), and platelet-derived growth factor (PDGF) within the bone regenerate.^{31,49,52,100,110,155,186,190} During distraction osteogenesis bone morphogenetic proteins-2 (BMP-2) and BMP-4 are produced by osteoblasts.^{53,147} Recent interest has focused on angiogenesis of the distraction induced bone regenerate and the expression of vascular endothelial growth factor-A (VEGF-A), VEGF-D, angiopoietin-1 and angiopoietin-2.^{28,44,51,77,82,121,130,142} These studies demonstrate the interdependence of the mechanical environment, angiogenesis, and bone formation during distraction osteogenesis. Expression of angiogenic genes and a proper mechanical environment are essential in supporting new vasculature for bone regeneration. Although our knowledge in this field is expanding, the complex cascade of growth factors regulating bone formation during distraction osteogenesis is still unclear.

6. Circular external skeletal fixation

External skeletal fixation systems have been used already for fracture stabilization from the mid-19th century onwards.¹⁸⁵ The concept of external fixation relies on fixation of bone by percutaneous pins linked with external connectors. External skeletal fixation systems can be used in various configurations. The three basic frame designs are the type I or unilateral configuration, the type II or bilateral fixation with transosseus pins, and the type III external fixation which combines types I and II to create a three dimensional frame.

Circular external skeletal fixation stands apart from the traditional external fixation systems as it is characterized by the use of metal rings surrounding the limb. Although circular external skeletal fixation systems were described in the early 20th century, Dr. Gavriil Abramovich Ilizarov was the first to develop this method of fixation into a clinical treatment modality.^{84,85} Circular external skeletal fixation relies on the use of transosseus wires under tension rather than pins to connect the bone to the external fixation rings (Fig 4). The system is highly versatile and permits the use of partial rings and posts for transcutaneous pins thus encompassing the features of traditional external fixators. In addition, hybridization of the circular fixation with other external fixation configurations is possible. Initially, Dr. Ilizarov developed his circular fixation system to treat fractures and nonunion. In order to stabilize bone fractures under compression, threaded

connecting rods were supplemented to the system. Anecdotal, Dr. Ilizarov recognized new bone formation in a patient who accidentally distracted rather than compressed the fracture gap. This observation was the start of his pursuit culminating in the concept of distraction osteogenesis.

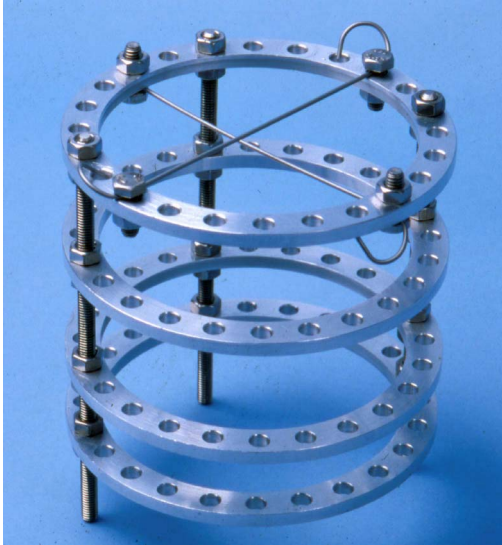


Fig 4. Circular external skeletal fixation system including rings, threaded connecting rods, and transosseus wires

Circular external skeletal fixation is used in veterinary medicine to treat a variety of fracture types.^{105,106,108,115} Its use in companion animals is restricted mainly to fractures distal of the elbow and stifle joint. Although hybrid frame designs can be used on the humerus and femur, the use of transosseus wires proximal of the elbow and knee has high morbidity associated with the large muscle volume in these areas. In addition to fracture treatment, circular fixation has been used effectively to perform panarthrodesis of carpal and tarsal joints. The stability of the frame depends on several factors. The most important parameter is ring diameter.¹⁰⁷ Larger ring diameters require longer transosseus wires, which result in larger moments on the wire-bone and wire-bolt interface. Ring diameter is determined depending on the diameter of the limb adding approximately 2 cm on either side to allow for soft tissue swelling. Larger diameters will result in increased instability at the fracture site. The second most important parameter is wire tension.^{7,107} Tensioning the transosseus wires will increase frame stability. Tension depends on the body weight of the patient and is applied by use of a dynamometric wire tensioner. An equivalent of 20 kg (± 200 N) is used for dogs up

to 10 kg in body weight, an equivalent of 40 kg (± 400 N) for dogs between 10 and 20 kg, an equivalent of 60 kg (± 600 N) for dogs between 20 and 30 kg, and an equivalent of 80 kg (± 800 N) for dogs 30 and 40 kg in body weight. Equivalents of up to 120 kg (± 1200 N) can be applied in giant breed dogs.^{84,85,105,106} In the basic configuration of the circular frame two transosseus wires are used per ring. The divergence angle between the transosseus wires has little impact on axial stability and is usually maintained between 60-90 degrees. Angles of divergence smaller than 60 degrees will decrease rotational and bending stability. Slippage of wires at the wire-bolt interface is a common cause for loss of initial stability. It is essential to tighten the nuts correctly after tensioning and to retighten them after cutting excess wire length and bending of the protruding wire ends to prevent auto-mutilation.

The basic design of circular fixators includes two full rings proximal of the fracture area each with two tensioned transosseus wires and two full rings distally. The rings are connected with 3 or 4 threaded rods to create a very stable frame. Transosseus wires with a diameter ranging from 1.2 to 1.6 mm are typically used in companion animals. Circular external skeletal fixation is very resistant to strain in bending and torsion. In contrast, it allows for micro-motion in the axial direction. This dynamic feature of circular fixation is considered to stimulate bone formation and thus bone healing.^{6,8}

By analogy with all other external skeletal fixation systems, infection at the transosseus wire-skin interface is the most common complication.^{105,106,113} This complication usually occurs at sites where there are large soft tissue masses covering the bone. Movement of soft tissues especially muscles will result in sliding movement at the wire-skin interface thus creating an easy access for bacteria. Avoiding large muscle masses can reduce this complication to a minimum. Proper care of the circular external skeletal fixation system especially of the transosseus wires remains essential as wire-tract infection is painful and restricts function. Bandages are effective in stabilizing the soft tissues adjacent to the transosseus wires and in protecting the skin-wire interface. Wire-tract infection should be treated aggressively with antibiotics and local wound care to avoid progression of the infection to the transosseus wire-bone interface. Established infection will necessitate transosseus wire removal and revision of the frame design.

Circular external skeletal fixation is well tolerated with proper care. Dogs are able to bear weight shortly after surgery, which is beneficial for both bone healing and musculoskeletal function. Dogs can be managed on an outpatient basis with weekly checkups. Exercise should be limited to leash walks until bone has consolidated and the frame can be removed. Although circular external skeletal

fixation is a valuable asset in treating fractures, its strength lies in the management of growth deformities and bone deficits.^{47,116,117}

7. Hormonal regulation of bone formation

The endocrine regulation of bone is mediated through several hormones, including growth hormone (GH), calcitonin, parathyroid hormone (PTH), calcitriol, androgen, and estrogen.^{126,167,174} Growth hormone is essential for longitudinal growth occurring in the epiphyseal plate through the process of endochondral ossification.^{12,33,54,125} Part of the actions of GH is mediated through the induction of IGF-I, but GH also directly promotes longitudinal bone growth.^{158,188} The expression of growth hormone receptor (GHR) within the growth plate is consistent with this finding.^{37,71,93} At least in humans, GH has a major role in the maintenance of bone mass in the adult skeleton by regulating bone remodeling through a complex interaction of circulating GH, insulin-like growth factors (IGFs), insulin-like growth factors binding proteins (IGFBPs), and locally produced IGFs and IGFBPs.¹⁷⁷ Relative or absolute GH deficiency thus results in osteoporosis.^{18,146,159} In dogs, GH deficiency or estrogen depletion does not lead to osteoporosis.¹⁷⁶ Osteoporosis does occur in renal or alimentary induced hyperparathyroidism in dogs.¹⁵⁷

Growth hormone has been used successfully in treating GH-deficiency related osteoporosis and postmenopausal osteoporosis in humans.^{101,146} In view of its role in bone metabolism, GH has been used experimentally to stimulate osteogenesis. Systemic GH application effectively stimulated bone formation in fracture models in different species, including dogs.^{10,149,191} In addition, GH was demonstrated to enhance consolidation of the bone regenerate after distraction osteogenesis.^{11,136}

Calcitonin, PTH and calcitriol are crucial in maintaining calcium homeostasis.^{123,166,174} Calcitonin is synthesized by the clear cells of the thyroid gland and stimulates calcium deposition while inhibiting bone resorption. Parathyroid hormone is synthesized by the parathyroid glands, which are located in or adjacent to the thyroid glands. The function of PTH is to mobilize calcium from bone and reabsorb calcium in the kidney. Continuously increased PTH levels increase osteoclast activity mediated through the shrinkage of osteoblasts thus exposing bone to osteoclasts, whereas intermittent treatment with PTH stimulates osteoblast activity and thus bone formation.⁶⁷

Skeletal growth and puberty are connected since sex hormones estrogen and androgen play a role in cartilage growth and endochondral ossification. Early castration results in delayed closure of the growth plate with subsequent increased

final bone length.¹⁴⁴ In adult dogs, estrogen and androgen do not seem to be essential in maintaining bone mass. Ovariectomy or orchidectomy, even at a young age, does not result in clinical osteoporosis. Nevertheless, estrogen depletion after ovariectomy does negatively affect cancellous bone growth and increases cortical porosity.^{156,193}

8. Skeletal growth factors

The family of insulin-like growth factors (IGF) consists of IGF-I, IGF-II and insulin. IGF-I and IGF-II are major constituents of both local and systemic growth factors.^{125,196} There are three receptors each with the highest affinity for their specific IGF. These receptors, particularly for IGF-I, are present in almost all tissues. IGF-I is mainly produced in the liver, but also in peripheral tissues including bone.^{46,158} Knowledge about the role of IGF-II in bone formation and bone healing remains limited.¹³ IGF-I and IGF-II are important as local growth factors for osteoblast survival and apoptosis.⁸¹ By modulating osteoblast-osteoclast interactions IGF-I and IGF-II are critical in bone remodeling and sustaining bone mass.^{78,80} In vitro, IGF-I stimulates existing bone resorption by existing osteoclasts and the formation of new osteoclasts from precursor cells.¹¹⁹ Upregulation of IGF-I and IGF-II expression occurs during bone healing.¹⁶⁴ IGF-I is also expressed in the bone regenerate during distraction osteogenesis.¹⁷⁰ Local treatment with IGF-I stimulates bone healing and bone consolidation after distraction osteogenesis.^{97,165}

Six high-affinity IGF binding proteins (IGFBP) regulate the actions of IGF-I and IGF-II.^{92,120} The IGFBPs are produced by osteoblast and IGFBP-4 and IGFBP-5 are the most abundant.^{74,120} In general, IGFBP-1, -2, -4, and -6 inhibit and IGFBP-3 and -5 stimulate osteoblast function.^{30,43,50} Overexpression of IGFBP-2 was demonstrated to impair long bone development in vivo by blocking the ability of IGF-I and IGF-II to promote cell proliferation and matrix synthesis.⁵⁸ Growth hormone treatment in postmenopausal women resulted in elevated serum levels of IGF-I, IGF-II, IGFBP-3, IGFBP-4, and decreased serum levels of IGFBP-1 and IGFBP-2. Serum levels of IGFBP-4 and IGFBP-5 correlated with bone mineral density in GH-deficient adults. Growth hormone replacement therapy increased both IGFBPs.¹⁷²

Although IGFs and IGFBPs are important for bone maintenance and formation their precise role is still to be elucidated.

The transforming growth factor- β superfamily consists of the transforming growth factor- β s (TGF- β s) and the bone morphogenetic proteins (BMPs). The TGF- β s are multifunctional peptides expressed in mammals as highly homologous isoforms, TGF- β 1, TGF- β 2, and TGF- β 3. The TGF- β s act via autocrine and

paracrine modes to control a variety of developmental processes.¹⁴ TGF- β s stimulate osteoblast proliferation and matrix production. In this way they are important regulators of bone formation and fracture repair^{17,19,163} An early decline in serum TGF- β 1 levels was demonstrated in patients with delayed fracture healing.¹⁹⁷ TGF- β s have been used successfully to stimulate fracture healing.¹⁶ In distraction osteogenesis, TGF- β s are typically expressed during the phase of active lengthening.^{110,170} Experimental stimulation of bone consolidation with TGF- β 1 during distraction osteogenesis was not successful indicating a role for TGF- β s especially in the early stages of osteogenesis.^{138,151}

BMPs were first described as factors capable of inducing new bone.¹⁷⁹ The group of BMPs comprises at least 15 growth factors, which are highly osteoinductive.²⁹ Besides their role in osteogenesis, BMPs exert essential functions during embryogenesis.¹⁸⁴ BMPs bind to two distinct types of transmembrane receptors with serine-threonine kinase activity. The activated receptors phosphorylate Smad proteins, which act as intracellular signal mediators.^{24,104} These Smads regulate the expression of target genes. The TGF- β s signal through the same Smad pathways. The BMPs can induce endochondral bone formation through the proliferation and differentiation of chondrocytes and osteoblasts.^{175,187} BMP-2, BMP-4, and BMP-7 expression was highest during the early phases of fracture healing.¹⁶¹ The expression of these BMPs diminished after the initial phases of osteogenesis. By analogy, BMP-2, BMP-4, and BMP-7 expression was most prominent during the phase of active lengthening during distraction osteogenesis, to decrease during the phase of bone consolidation.¹³⁷ BMP-2 and BMP-7 are the most potent osteoinductive growth factors and have been used both experimentally and clinically to stimulate bone formation.^{73,91,181} Part of the mitogenic action of BMP-7 is mediated through the modulation of IGF-II secretion and the balance between stimulatory and inhibitory IGF-BPs.⁹⁶ BMP-2 was most effective in stimulating bone healing in an experimental fracture model when administered immediately postoperatively.¹²⁴ The delivery systems by which BMPs are administered also play an important role in their effectiveness.¹⁵² In dogs, BMPs have been used successfully to treat nonunions and segmental bone defects.^{35,79}

The group of fibroblast growth factors (FGFs) consists of 18 growth factors. Fibroblast growth factor-1 (FGF-1 or acidic FGF) and fibroblast growth factor-2 (FGF-2 or basic FGF) are the prototypic members of the FGF family. Fibroblast growth factors and their receptors have an important role in the control of endochondral and intramembranous bone formation.^{14,72} The expression of FGFs is closely related to the expression of vascular endothelial growth factors (VEGFs). Both FGF-2 and VEGFs are expressed in the early stages of distraction osteogenesis and are considered essential for angiogenesis.^{82,130,195} Expression of

VEGFs occurs at the osteogenic front during distraction osteogenesis and precedes the expression of BMPs by osteoblasts. This demonstrates that angiogenesis is induced before osteogenesis.¹⁶⁰ VEGF was capable of stimulating bone healing in an atrophic nonunion model.⁴⁵ In contrast, VEGF application was not effective in increasing bone consolidation in a distraction osteogenesis model.⁴⁴ Combined delivery of angiogenic (VEGF) and osteogenic (BMP) factors was more effective in enhancing new bone formation than application of the single factors.⁸³

Platelet-derived growth factor (PDGF) was initially isolated from platelets, but was found to be expressed by skeletal and a variety of non-skeletal tissues.¹⁹ Nevertheless, platelets are the major source of this growth factor, which is released following platelet aggregation. Aggregation of platelets typically occurs after fracture and soft tissue trauma. This emphasizes the role of PDGF in fracture repair. Platelet-derived growth factor increases the replication of cells of osteoblastic and osteoclastic lineages.¹⁴

Hepatocyte growth factor (HGF) was discovered as a potent growth-promoting agent in liver cells. HGF is expressed in most tissues and plays an important role in tissue repair. This growth factor stimulates both osteoclasts and osteoblasts, thus regulating bone remodeling. In addition, HGF is expressed at the fracture site and induces upregulation of BMP receptors.⁸⁶ In this way, HGF facilitates BMP signaling and stimulates bone healing.

The identification of the receptor activator of nuclear factor kappaB ligand (RANKL), its cognate receptor RANK, and its decoy receptor osteoprotegerin (OPG) resulted in a new perspective on osteoclast function and bone homeostasis.¹⁶⁹ Osteoclast precursors express RANK and differentiate into osteoclasts after recognizing RANKL through cell-to-cell interaction with osteoblasts and stromal cells.^{166,171} Membrane bound colony-stimulating factor-1 (mCSF-1) plays an important role in osteoblast-mediated osteoclastogenesis.^{189,194} Osteoprotegerin acts as a soluble receptor antagonist for RANKL that prevents it from binding to and activating RANK. Hepatitis C-associated osteosclerosis is characterized by diffuse osteosclerosis, decreased osteoclast numbers, and increased OPG serum levels in adult humans. An imbalance in the OPG/RANKL system is supposed to be responsible for these findings.¹¹⁴ There is increasing evidence that the OPG/RANKL system links the skeletal with the vascular system.^{141,148} These findings may lead to new therapeutic regimes in preventing bone resorption. For instance, GH replacement therapy increased OPG levels, which may lead to a positive bone balance by inhibiting osteoclastogenesis.^{103,178}

Ghrelin, the endogenous ligand for the GH secretagogue receptor (GHS-R) is a recently discovered brain-gut peptide involved in GH secretion and energy homeostasis.⁶⁶ In addition, ghrelin is widely expressed in several tissues, where it might therefore act as a paracrine or autocrine factor. Ghrelin directly regulates

bone formation by stimulating osteoblast proliferation, differentiation, and function and by inhibiting apoptosis.^{66,94,111} Ghrelin is also synthesized and secreted by chondrocytes.²³ In future, ghrelin and synthetic GHS-R ligands may be effective in modulating bone homeostasis.

9. Bone markers

Bone markers are used to monitor bone metabolism non-invasively. These markers can be divided into markers of bone formation and markers of bone resorption, respectively. Although the commercially available bone marker assays were designed initially for use in humans, several of these assays are also validated to monitor bone metabolism in laboratory animals, dogs and horses.^{20,26,27,99}

The markers of bone metabolism can be subdivided into enzymatic markers and metabolic products of bone formation and resorption, respectively.^{22,153,154} Enzymatic markers include bone-specific alkaline phosphatase (BAP) and tartrate-resistant acid phosphatase (TRAP)^{90,127}. Bone-specific alkaline phosphatase is an osteoblast-related marker of bone formation.^{38,69} Tartrate-resistant acid phosphatase-5b (TRAP-5b) is an osteoclast-related marker of bone resorption.^{76,173} Serum or plasma markers of metabolic products of bone metabolism include osteocalcin (OC), carboxyterminal propeptide of type-I procollagen (PICP), aminoterminal propeptide of type-I procollagen (PINP), cross-linked carboxyterminal telopeptide of type-I collagen (ICTP), and C-terminal cross-linked telopeptide of type-I collagen (CTX). Osteocalcin is an osteoblast-related marker of bone formation, but its precise function is unknown.¹⁶⁸ Increased bone formation was present in OC-deficient mice, indicating a suppressive role of OC on bone formation, possibly through inhibition of osteopontin.⁴² Markers PICP and PINP are metabolic products of type-I collagen synthesis and hence bone formation.^{36,133,167} Markers ICTP and CTX are products of type-I collagen breakdown and hence bone resorption.^{2,20,132,150} Marker ICTP is released through the actions of matrix metalloproteinases (MMPs) and is therefore also known as CTX-MMP. Marker CTX is formed from collagen I by cysteine proteinases, including cathepsin K.⁷⁰ The collagen type I C-telopeptide is susceptible to molecular rearrangement. In newly synthesized collagen this site is in the native form, but during aging a spontaneous reaction occurs, resulting in three age-modified isomerized and racemized fragments. These modified forms of CTX may provide new diagnostic and monitoring tools in evaluating bone disease.³²

Bone markers have been used successfully in humans to monitor metabolic bone disease and the effect of treatment. In rheumatoid arthritis, characterized by increased bone resorption, ICTP levels were elevated while OC levels were

decreased.¹ Growth hormone treatment in GH-deficient patients resulted in transient changes in BAP, OC, and ICTP levels.^{18,146,159} Serum BAP and PINP were the most sensitive markers for monitoring treatment efficacy in Paget's disease.³ In Cushing's syndrome, the markers OC and CTX decreased consistent with increased bone resorption.³⁶ The markers BAP, OC, PINP, and CTX were most effective in monitoring osteoporosis.^{39,40} Although bone mass measurements at the present time still supply most information about fracture risk, markers of bone resorption may be useful in predicting fracture risk in osteoporosis.⁶⁸ In chronic renal failure, PICP and ICTP have been used as markers of bone formation and resorption, respectively.¹³³ Bone markers have also been used to monitor bone growth. In pubertal boys and girls, markers of bone metabolism related positively to growth velocity.¹⁸² This finding could be of great interest in monitoring the rapid growth phase in dogs.

In dogs, normal values for BAP, OC and ICTP have been established and are not affected by breed size.^{20,99} By analogy with humans, bone markers demonstrate a circadian rhythm in the dog.^{99,109,131} Therefore, sampling should be standardized both in an experimental and clinical setting. Bone markers have not been used extensively to monitor bone pathology in dogs. Experimentally induced osteomyelitis increased ICTP serum levels.¹³² Orchiectomy in male beagle dogs resulted in decreased BAP and OC levels indicating an imbalance in bone metabolism with diminished bone formation.⁶⁵ In canine appendicular osteosarcoma, BAP proved to be a prognostic factor in predicting survival time.⁴⁸ Experimentally, OC was used effectively to monitor new bone formation following distraction osteogenesis.^{15,55,100,155} Although bone markers can be an adjunct in monitoring bone metabolism, they cannot replace bone histomorphometry in orthopaedic research at the present time.

References

1. Al Awadhi A, Olusi S, Al Zaid N, Prabha K. Serum concentrations of interleukin 6, osteocalcin, intact parathyroid hormone, and markers of bone resorption in patients with rheumatoid arthritis. *J Rheumatol* 1999;26:1250-1256.
2. Allen MJ, Hoffmann WE, Richardson DC, Breur GJ. Serum markers of bone metabolism in dogs. *Am J Vet Res* 1998;59:250-254.
3. Alvarez L, Guanabens N, Peris P, Vidal S, Ros I, Monegal A, Bedini JL, Deulofeu R, Pons F, Munoz-Gomez J, Ballesta AM. Usefulness of biochemical markers of bone turnover in assessing response to the treatment of Paget's disease. *Bone* 2001;29:447-452.
4. Aronson J. Experimental and clinical experience with distraction osteogenesis. *Cleft Palate Craniofac J* 1994;31:473-481.
5. Aronson J, Good B, Stewart C, Harrison B, Harp J. Preliminary studies of mineralization during distraction osteogenesis. *Clin Orthop* 1990;250:43-49.
6. Aronson J, Harp JH. Mechanical forces as predictors of healing during tibial lengthening by distraction osteogenesis. *Clin Orthop* 1994;310:73-79.
7. Aronson J, Harrison B, Boyd CM, Cannon DJ, Lubansky HJ. Mechanical induction of osteogenesis: the importance of pin rigidity. *J Pediatr Orthop* 1988;8:396-401.
8. Aronson J, Harrison B, Boyd CM, Cannon DJ, Lubansky HJ, Stewart C. Mechanical induction of Osteogenesis. Preliminary studies. *Ann Clin Lab Sci* 1988;18:195-203.
9. Aronson J, Harrison BH, Stewart CL, Harp JH, Jr. The histology of distraction osteogenesis using different external fixators. *Clin Orthop* 1989;241:106-116.
10. Bail HJ, Kolbeck S, Krummrey G, Schmidmaier G, Haas NP, Raschke MJ. Systemic application of growth hormone for enhancement of secondary and intramembranous fracture healing. *Horm Res* 2002;58 Suppl 3:39-42.
11. Bail HJ, Kolbeck S, Lindner T, Dahne M, Weiler A, Windhagen HJ, Raun K, Skjaerbaek C, Flyvbjerg A, Orskov H, Haas NP, Raschke MJ. The effect of growth hormone on insulin-like growth factor I and bone metabolism in distraction osteogenesis. *Growth Horm IGF Res* 2001;11:314-323.
12. Baroncelli GI, Bertelloni S, Ceccarelli C, Cupelli D, Saggese G. Dynamics of bone turnover in children with GH deficiency treated with GH until final height. *Eur J Endocrinol* 2000;142:549-556.
13. Bautista CM, Mohan S, Baylink DJ. Insulin-like growth factors I and II are present in the skeletal tissues of ten vertebrates. *Metabolism* 1990;39:96-100.
14. Baylink DJ, Finkelman RD, Mohan S. Growth factors to stimulate bone formation. *J Bone Miner Res* 1993;8 Suppl 2:565-572.
15. Blair HC, Robinson LJ, Zaidi M. Osteoclast signalling pathways. *Biochem Biophys Res Commun* 2005;328:728-738.
16. Blumenfeld I, Srouji S, Lanir Y, Laufer D, Livne E. Enhancement of bone defect healing in old rats by TGF-beta and IGF-1. *Exp Gerontol* 2002;37:553-565.
17. Bolander ME. Regulation of fracture repair by growth factors. *Proc Soc Exp Biol Med* 1992;200:165-170.
18. Bollerslev J, Moller J, Thomas S, Djoseland O, Christiansen JS. Dose-dependent effects of recombinant human growth hormone on biochemical markers of bone and collagen metabolism in adult growth hormone deficiency. *Eur J Endocrinol* 1996;135:666-671.
19. Bourque WT, Gross M, Hall BK. Expression of four growth factors during fracture repair. *Int J Dev Biol* 1993;37:573-579.

20. Breur GJ, Allen MJ, Carlson SJ, Richardson DC. Markers of bone metabolism in dog breeds of different size. *Res Vet Sci* 2004;76:53-55.
21. Brown JH, DeLuca SA. Growth plate injuries: Salter-Harris classification. *Am Fam Physician* 1992;46:1180-1184.
22. Calvo MS, Eyre DR, Gundberg CM. Molecular basis and clinical application of biological markers of bone turnover. *Endocr Rev* 1996;17:333-368.
23. Caminos JE, Gualillo O, Lago F, Otero M, Blanco M, Gallego R, Garcia-Caballero T, Goldring MB, Casanueva FF, Gomez-Reino JJ, Dieguez C. The endogenous growth hormone secretagogue (ghrelin) is synthesized and secreted by chondrocytes. *Endocrinology* 2005;146:1285-1292.
24. Cao X, Chen D. The BMP signaling and in vivo bone formation. *Gene* 2005;357:1-8.
25. Carmichael S. Current concepts in the management of growth plate abnormalities. *Vet Ann* 1992;32:47-60.
26. Carstansen B, Hoyle NR, Gabriel A, Hars O, Sandersen C, Amory H, Remy B. Evaluation of plasma carboxy-terminal cross-linking telopeptide of type I collagen concentration in horses. *Am J Vet Res* 2004;65:104-109.
27. Carstansen B, Sulon J, Banga-Mboko H, Beckers JF, Remy B. Development and validation of a specific radioimmunoassay for equine osteocalcin. *Domest Anim Endocrinol* 2003;24:31-41.
28. Carvalho RS, Einhorn TA, Lehmann W, Edgar C, Al Yamani A, Apazidis A, Pacicca D, Clemens TL, Gerstenfeld LC. The role of angiogenesis in a murine tibial model of distraction osteogenesis. *Bone* 2004;34:849-861.
29. Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. *Growth Factors* 2004;22:233-241.
30. Chevalley T, Strong DD, Mohan S, Baylink D, Linkhart TA. Evidence for a role for insulin-like growth factor binding proteins in glucocorticoid inhibition of normal human osteoblast-like cell proliferation. *Eur J Endocrinol* 1996;134:591-601.
31. Cillo JE, Jr., Gassner R, Koepsel RR, Buckley MJ. Growth factor and cytokine gene expression in mechanically strained human osteoblast-like cells: implications for distraction osteogenesis. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2000;90:147-154.
32. Cloos PA, Fledelius C, Christgau S, Christiansen C, Engsig M, Delmas P, Body JJ, Garnero P. Investigation of bone disease using isomerized and racemized fragments of type I collagen. *Calcif Tissue Int* 2003;72:8-17.
33. Conzemi MG, Brown DC, Brabec M, Smith GK, Washabau R, LaFond E, Chakraborty PK. Correlation between longitudinal bone growth, growth hormone, and insulin-like growth factor-I in prepubertal dogs. *Am J Vet Res* 1998;59:1608-1612.
34. Conzemi MG, Smith GK, Brighton CT, Marion MJ, Gregor TP. Analysis of physal growths in dogs, using biplanar radiography. *Am J Vet Res* 1994;55:22-27.
35. Cook SD, Baffes GC, Wolfe MW, Sampath TK, Rueger DC. Recombinant human bone morphogenetic protein-7 induces healing in a canine long-bone segmental defect model. *Clin Orthop* 1994;301:302-312.
36. Cortet B, Cortet C, Blanckaert F, d'Herbomez M, Marchandise X, Wemeau JL, Decoux M, Dewailly D. Quantitative ultrasound of bone and markers of bone turnover in Cushing's syndrome. *Osteoporos Int* 2001;12:117-123.
37. Cruickshank J, Grossman DI, Peng RK, Famula TR, Oberbauer AM. Spatial distribution of growth hormone receptor, insulin-like growth factor-I receptor and apoptotic chondrocytes during growth plate development. *J Endocrinol* 2005;184:543-553.
38. de Vernejoul MC. Markers of bone remodelling in metabolic bone disease. *Drugs Aging* 1998;12 Suppl 1:9-14.

39. Delmas PD. Markers of bone turnover for monitoring treatment of osteoporosis with antiresorptive drugs. *Osteoporos Int* 2000;11 Suppl 6:66-76.
40. Delmas PD, Eastell R, Garnero P, Seibel MJ, Stepan J. The use of biochemical markers of bone turnover in osteoporosis. *Osteoporos In* 2000;11 Suppl 6:2-17.
41. Draenert Y, Draenert K. Gap healing of compact bone. *Scan Electron Microsc* 1980;103-111.
42. Ducy P, Desbois C, Boyce B, Pinero G, Story B, Dunstan C, Smith E, Bonadio J, Goldstein S, Gundberg C, Bradley A, Karsenty G. Increased bone formation in osteocalcin-deficient mice. *Nature* 1996;382:448-452.
43. Durham SK, Riggs BL, Conover CA. The insulin-like growth factor-binding protein-4 (IGFBP-4)-IGFBP-4 protease system in normal human osteoblast-like cells: regulation by transforming growth factor-beta. *J Clin Endocrinol Metab* 1994;79:1752-1758.
44. Eckardt H, Bundgaard KG, Christensen KS, Lind M, Hansen ES, Hvid I. Effects of locally applied vascular endothelial growth factor (VEGF) and VEGF-inhibitor to the rabbit tibia during distraction osteogenesis. *J Orthop Res* 2003;21:335-340.
45. Eckardt H, Ding M, Lind M, Hansen ES, Christensen KS, Hvid I. Recombinant human vascular endothelial growth factor enhances bone healing in an experimental nonunion model. *J Bone Joint Surg Br* 2005;87:1434-1438.
46. Edwall D, Schalling M, Jennische E, Norstedt G. Induction of insulin-like growth factor I messenger ribonucleic acid during regeneration of rat skeletal muscle. *Endocrinology* 1989;124:820-825.
47. Ehrhart N. Longitudinal bone transport for treatment of primary bone tumors in dogs: technique description and outcome in 9 dogs. *Vet Surg* 2005;34:24-34.
48. Ehrhart N, Dernel WS, Hoffmann WE, Weigel RM, Powers BE, Withrow SJ. Prognostic importance of alkaline phosphatase activity in serum from dogs with appendicular osteosarcoma: 75 cases (1990-1996). *J Am Vet Med Assoc* 1998;213:1002-1006.
49. Eingartner C, Coerper S, Fritz J, Gaissmaier C, Koveker G, Weise K. Growth factors in distraction osteogenesis. Immuno-histological pattern of TGF-beta1 and IGF-I in human callus induced by distraction osteogenesis. *Int Orthop* 1999;23:253-259.
50. Ernst M, Rodan GA. Increased activity of insulin-like growth factor (IGF) in osteoblastic cells in the presence of growth hormone (GH): positive correlation with the presence of the GH-induced IGF-binding protein BP-3. *Endocrinology* 1990;127:807-814.
51. Fang TD, Salim A, Xia W, Nacamuli RP, Guccione S, Song HM, Carano RA, Filvaroff EH, Bednarski MD, Giaccia AJ, Longaker MT. Angiogenesis is required for successful bone induction during distraction osteogenesis. *J Bone Miner Res* 2005;20:1114-1124.
52. Farhadieh RD, Dickinson R, Yu Y, Gianoutsos MP, Walsh WR. The role of transforming growth factor-beta, insulin-like growth factor I, and basic fibroblast growth factor in distraction osteogenesis of the mandible. *J Craniofac Surg* 1999;10:80-86.
53. noutsos MP, Yu Y, Walsh WR. The role of bone morphogenetic proteins BMP-2 and BMP-4 and their related postreceptor signaling system (Smads) in distraction osteogenesis of the mandible. *J Craniofac Surg* 2004;15:714-718.
54. Favier RP, Mol JA, Kooistra HS, Rijnberk A. Large body size in the dog is associated with transient GH excess at a young age. *J Endocrinol* 2001;170:479-484.
55. Fink B, Fox F, Singer J, Skripitz R, Feldkamp J. Monitoring of bone formation during distraction osteogenesis via osteocalcin: a time sequence study in dogs. *J Orthop Sci* 2002;7:557-561.
56. Fink B, Schwinger G, Singer J, Sager M, Wilke C, Braunstein S. The effect of tibial lengthening using the Ilizarov method on the cartilage and the menisci of the knee joint. *J Orthop Res* 2001;19:665-670.

57. Fink B, Schwinger G, Singer J, Schmielau G, Ruther W. Biomechanical properties of tendons during lower-leg lengthening in dogs using the Ilizarov method. *J Biomech* 1999;32:763-768.
58. Fisher MC, Meyer C, Garber G, Dealy CN. Role of IGFBP2, IGF-I and IGF-II in regulating long bone growth. *Bone* 2005;37:741-750.
59. Fjeld TO. Growth plate injury and retardation of growth in the canine radius and ulna. Traumatiske epifyseskiveskader og vekstretardasjon i radius og ulna hos hund. *Norsk Vet* 1986;98:513-519.
60. Fjeld TO. Patterns of traumatic premature closure of growth plates in the canine radius and ulna
De ulike typer av traumatisk betinget prematur epifyseskivelukning i radius og ulna hos hund. *Norsk Vet* 1986;98:633-641.
61. Forell EB, Schwarz PD. Use of external skeletal fixation for treatment of angular deformity secondary to premature distal ulnar physal closure. *J Am Anim Hosp Assoc* 1993;29:460-476.
62. Fox SM. Premature closure of distal radial and ulnar physes in the dog. I. Pathogenesis and diagnosis. *Compend Contin Educ Pract Vet* 1984;6:128-138.
63. Fox SM. Premature closure of distal radial and ulnar physes in the dog. II. Treatment. *Comp Cont Educ Pract Vet* 1984;6:212-221.
64. Frierson M, Ibrahim K, Boles M, Bote H, Ganey T. Distraction osteogenesis. A comparison of corticotomy techniques. *Clin Orthop* 1994;301:19-24.
65. Fukuda S, Iida H. Effects of orchidectomy on bone metabolism in beagle dogs. *J Vet Med Sci* 2000;62:69-73.
66. Fukushima N, Hanada R, Teranishi H, Fukue Y, Tachibana T, Ishikawa H, Takeda S, Takeuchi Y, Fukumoto S, Kangawa K, Nagata K, Kojima M. Ghrelin directly regulates bone formation. *J Bone Miner Res* 2005;20:790-798.
67. Gabet Y, Kohavi D, Muller R, Chorev M, Bab I. Intermittently administered parathyroid hormone 1-34 reverses bone loss and structural impairment in orchietomized adult rats. *Osteoporos Int* 2005;16:1436-1443.
68. Garnero P. Markers of bone turnover for the prediction of fracture risk. *Osteoporos Int* 2000;11 Suppl 6:55-65.
69. Garnero P, Delmas PD. Bone markers. *Baillieres Clin Rheumatol* 1997;11:517-537.
70. Garnero P, Ferreras M, Karsdal MA, Nicamhlaobh R, Risteli J, Borel O, Qvist P, Delmas PD, Foged NT, Delaïsse JM. The type I collagen fragments ICTP and CTX reveal distinct enzymatic pathways of bone collagen degradation. *J Bone Miner Res* 2003;18:859-867.
71. Gevers EF, van der Eerden BC, Karperien M, Raap AK, Robinson IC, Wit JM. Localization and regulation of the growth hormone receptor and growth hormone-binding protein in the rat growth plate. *J Bone Miner Res* 2002;17:1408-1419.
72. Goodman SB, Song Y, Yoo JY, Fox N, Trindade MC, Kajiyama G, Ma T, Regula D, Brown J, Smith RL. Local infusion of FGF-2 enhances bone ingrowth in rabbit chambers in the presence of polyethylene particles. *J Biomed Mater Res A* 2003;65:454-461.
73. Govender PV, Rampersaud YR, Rickards L, Fehlings MG. Use of osteogenic protein-1 in spinal fusion: literature review and preliminary results in a prospective series of high-risk cases. *Neurosurg Focus* 2002;13 e4:1-6.
74. Govoni KE, Baylink DJ, Mohan S. The multi-functional role of insulin-like growth factor binding proteins in bone. *Pediatr Nephrol* 2005;20:261-268.
75. Green E, Lubahn JD, Evans J. Risk factors, treatment, and outcomes associated with nonunion of the midshaft humerus fracture. *J Surg Orthop Adv* 2005;14:64-72.
76. Halleen JM, Alatalo SL, Suominen H, Cheng S, Janckila AJ, Vaananen HK. Tartrate-resistant acid phosphatase 5b: a novel serum marker of bone resorption. *J Bone Miner Res* 2000;15:1337-1345.

77. Hansen-Algenstaedt N, Algenstaedt P, Bottcher A, Joscheck C, Schwarzloh B, Schaefer C, Muller I, Koike C, Ruther W, Fink B. Bilaterally increased VEGF-levels in muscles during experimental unilateral callus distraction. *J Orthop Res* 2003;21:805-812.
78. Hayden JM, Mohan S, Baylink DJ. The insulin-like growth factor system and the coupling of formation to resorption. *Bone* 1995;17:93-98.
79. Heckman JD, Boyan BD, Aufdemorte TB, Abbott JT. The use of bone morphogenetic protein in the treatment of non-union in a canine model. *J Bone Joint Surg Am* 1991;73:750-764.
80. Hill PA, Reynolds JJ, Meikle MC. Osteoblasts mediate insulin-like growth factor-I and -II stimulation of osteoclast formation and function. *Endocrinology* 1995;136:124-131.
81. Hill PA, Tumber A, Meikle MC. Multiple extracellular signals promote osteoblast survival and apoptosis. *Endocrinology* 1997;138:3849-3858.
82. Hu J, Zou S, Li J, Chen Y, Wang D, Gao Z. Temporospatial expression of vascular endothelial growth factor and basic fibroblast growth factor during mandibular distraction osteogenesis. *J Craniomaxillofac Surg* 2003;31:238-243.
83. Huang YC, Kaigler D, Rice KG, Krebsbach PH, Mooney DJ. Combined angiogenic and osteogenic factor delivery enhances bone marrow stromal cell-driven bone regeneration. *J Bone Miner Res* 2005;20:848-857.
84. Ilizarov GA. The tension-stress effect on the genesis and growth of tissues. Part I. The influence of stability of fixation and soft-tissue preservation. *Clin Orthop* 1989;238:249-281.
85. Ilizarov GA. The tension-stress effect on the genesis and growth of tissues: Part II. The influence of the rate and frequency of distraction. *Clin Orthop* 1989;239:263-285.
86. Imai Y, Terai H, Nomura-Furuwatari C, Mizuno S, Matsumoto K, Nakamura T, Takaoka K. Hepatocyte Growth Factor Contributes to Fracture Repair by Upregulating the Expression of BMP Receptors. *J Bone Miner Res* 2005;20:1723-1730.
87. Jazrawi LM, Majeska RJ, Klein ML, Kagel E, Stromberg L, Einhorn TA. Bone and cartilage formation in an experimental model of distraction osteogenesis. *J Orthop Trauma* 1998;12:111-116.
88. Johnson JA, Austin C, Breur GJ. Incidence of canine appendicular musculoskeletal disorders in 16 veterinary teaching hospitals from 1980 through 1989. *Vet Comp Orthop Traumatol* 1994;7:56-69.
89. Johnson JM, Johnson AL, Eurell JA. Histological appearance of naturally occurring canine physeal fractures. *Vet Surg* 1994;23:81-86.
90. Kabata T, Tsuchiya H, Sakurakichi K, Yamashiro T, Watanabe K, Tomita K. Reconstruction with distraction osteogenesis for juxta-articular nonunions with bone loss. *J Trauma* 2005;58:1213-1222.
91. Kain MS, Einhorn TA. Recombinant human bone morphogenetic proteins in the treatment of fractures. *Foot Ankle Clin* 2005;10:639-650.
92. Kelley KM, Oh Y, Gargosky SE, Gucev Z, Matsumoto T, Hwa V, Ng L, Simpson DM, Rosenfeld RG. Insulin-like growth factor-binding proteins (IGFBPs) and their regulatory dynamics. *Int J Biochem Cell Biol* 1996;28:619-637.
93. Kelly PA, Finidori J, Moulin S, Kedzia C, Binart N. Growth hormone receptor signalling and actions in bone growth. *Horm Res* 2001;55 Suppl 2:14-17.
94. Kim SW, Her SJ, Park SJ, Kim D, Park KS, Lee HK, Han BH, Kim MS, Shin CS, Kim SY. Ghrelin stimulates proliferation and differentiation and inhibits apoptosis in osteoblastic MC3T3-E1 cells. *Bone* 2005;37:359-369.
95. Klotch DW, Ganey TM, Slater-Haase A, Sasse J. Assessment of bone formation during osteoneogenesis: a canine model. *Otolaryngol Head Neck Surg* 1995;112:291-302.

96. Knutsen R, Honda Y, Strong DD, Sampath TK, Baylink DJ, Mohan S. Regulation of insulin-like growth factor system components by osteogenic protein-1 in human bone cells. *Endocrinology* 1995;136:857-865.
97. Kobayashi K, Agrawal K, Jackson IT, Vega JB. The effect of insulin-like growth factor 1 on craniofacial bone healing. *Plast Reconstr Surg* 1996;97:1129-1135.
98. Kojimoto H, Yasui N, Goto T, Matsuda S, Shimomura Y. Bone lengthening in rabbits by callus distraction. The role of periosteum and endosteum. *J Bone Joint Surg Br* 1988;70:543-549.
99. Ladlow JF, Hoffmann WE, Breur GJ, Richardson DC, Allen MJ. Biological variability in serum and urinary indices of bone formation and resorption in dogs. *Calcif Tissue Int* 2002;70:186-193.
100. Lammens J, Liu Z, Aerssens J, Dequeker J, Fabry G. Distraction bone healing versus osteotomy healing: a comparative biochemical analysis. *J Bone Miner Res* 1998;13:279-286.
101. Landin-Wilhelmsen K, Nilsson A, Bosaeus I, Bengtsson BA. Growth hormone increases bone mineral content in postmenopausal osteoporosis: a randomized placebo-controlled trial. *J Bone Miner Res* 2003;18:393-405.
102. Langley-Hobbs SJ, Carmichael S, Pead MJ, Torrington AM. Management of antebrachial deformity and shortening secondary to a synostosis in a dog. *J Small Anim Pract* 1996;37:359-363.
103. Lanzi R, Losa M, Villa I, Gatti E, Sirtori M, Dal Fiume C, Rubinacci A. GH replacement therapy increases plasma osteoprotegerin levels in GH-deficient adults. *Eur J Endocrinol* 2003;148:185-191.
104. Larsson J, Karlsson S. The role of Smad signaling in hematopoiesis. *Oncogene* 2005;24:5676-5692.
105. Latte Y. Application of the Ilizarov method in veterinary orthopaedic surgery (part 1). *Eur J Comp Anim Pract* 1997;7:26-50.
106. Latte Y. 75 applications of the Ilizarov method (part 2). *Eur J Comp Anim Pract* 1998;8:64-81.
107. Lewis DD, Bronson DG, Cross AR, Welch RD, Kubilis PS. Axial characteristics of circular external skeletal fixator single ring constructs. *Vet Surg* 2001;30:386-394.
108. Lewis DD, Cross AR, Carmichael S, Anderson MA. Recent advances in external skeletal fixation. *J Small Anim Pract* 2001;42:103-112.
109. Liesegang A, Reutter R, Sassi ML, Risteli J, Kraenzlin M, Riond JL, Wanner M. Diurnal variation in concentrations of various markers of bone metabolism in dogs. *Am J Vet Res* 1999;60:949-953.
110. Liu Z, Luyten FP, Lammens J, Dequeker J. Molecular signaling in bone fracture healing and distraction osteogenesis. *Histol Histopathol* 1999;14:587-595.
111. Maccarinelli G, Sibilia V, Torsello A, Raimondo F, Pitto M, Giustina A, Netti C, Cocchi D. Ghrelin regulates proliferation and differentiation of osteoblastic cells. *J Endocrinol* 2005;184:249-256.
112. MacPherson GC, Lewis DD, Johnson KA, Allen GS, Yovich JC. Fragmented coronoid process associated with premature distal radial physeal closure in four dogs. *Vet Comp Orthop Traumatol* 1992;5:93-99.
113. Maffulli N, Lombardi C, Matarazzo L, Nele U, Pagnotta G, Fixsen JA. A review of 240 patients undergoing distraction osteogenesis for congenital post-traumatic or postinfective lower limb length discrepancy. *J Am Coll Surg* 1996;182:394-402.
114. Manganelli P, Giuliani N, Fietta P, Mancini C, Lazzaretti M, Pollini A, Quaini F, Pedrazzoni M. OPG/RANKL system imbalance in a case of hepatitis C-associated osteosclerosis: the pathogenetic key? *Clin Rheumatol* 2005;24:296-300.

115. Marcellin-Little DJ. Fracture treatment with circular external fixation. *Vet Clin North Am Small Anim Pract* 1999;29:1153-70.
116. Marcellin-Little DJ. Treating bone deformities with circular external skeletal fixation. *Compend Contin Educ Pract Vet* 1999;21:481-491.
117. Marcellin-Little DJ, Ferretti A, Roe SC, DeYoung DJ. Hinged Ilizarov external fixation for correction of antebrachial deformities. *Vet Surg* 1998;27:231-245.
118. Matsuyama J, Ohnishi I, Kageyama T, Oshida H, Suwabe T, Nakamura K. Osteogenesis and angiogenesis in regenerating bone during transverse distraction: quantitative evaluation using a canine model. *Clin Orthop Relat Res* 2005;433:243-250.
119. Mochizuki H, Hakeda Y, Wakatsuki N, Usui N, Akashi S, Sato T, Tanaka K, Kumegawa M. Insulin-like growth factor-I supports formation and activation of osteoclasts. *Endocrinology* 1992;131:1075-1080.
120. Mohan S. Insulin-like growth factor binding proteins in bone cell regulation. *Growth Regul* 1993;3:67-70.
121. Moore DC, Leblanc CW, Muller R, Crisco JJ, III, Ehrlich MG. Physiologic weight-bearing increases new vessel formation during distraction osteogenesis: a micro-tomographic imaging study. *J Orthop Res* 2003;21:489-496.
122. Morgan PW, Miller CW. Osteotomy for correction of premature growth plate closure in 24 dogs. *Vet Comp Orthop Traumatol* 1994;7:129-135.
123. Muhlbauer RC, Fleisch H. The diurnal rhythm of bone resorption in the rat. Effect of feeding habits and pharmacological inhibitors. *J Clin Invest* 1995;95:1933-1940.
124. Murnaghan M, McIlmurray L, Mushipe MT, Li G. Time for treating bone fracture using rhBMP-2: a randomised placebo controlled mouse fracture trial. *J Orthop Res* 2005;23:625-631.
125. Nap RC, Mol JA, Hazewinkel HA. Age-related plasma concentrations of growth hormone (GH) and insulin-like growth factor I (IGF-I) in Great Dane pups fed different dietary levels of protein. *Domest Anim Endocrinol* 1993;10:237-247.
126. Niu T, Rosen CJ. The insulin-like growth factor-I gene and osteoporosis: A critical appraisal. *Gene* 2005;361:38-56.
127. Ohashi S, Ohnishi I, Kageyama T, Fukuda S, Tsuchiya A, Imai K, Matsuyama J, Nakamura K. Effect of Vascularity on Canine Distracted Tibial Callus Consolidation. *Clin Orthop Relat Res* 2005;438:253-259.
128. Olson NC, Brinker WO, Carrig CB. Premature closure of the distal radial physis in two dogs. *J Am Vet Med Assoc* 1980;176:906-910.
129. Orbay JL, Frankel VH, Finkle JE, Kummer FJ. Canine leg lengthening by the Ilizarov technique. A biomechanical, radiologic, and morphologic study. *Clin Orthop* 1992;278:265-273.
130. Pacicca DM, Patel N, Lee C, Salisbury K, Lehmann W, Carvalho R, Gerstenfeld LC, Einhorn TA. Expression of angiogenic factors during distraction osteogenesis. *Bone* 2003;33:889-898.
131. Panteghini M, Pagani F. Biological variation in bone-derived biochemical markers in serum. *Scand J Clin Lab Invest* 1995;55:609-616.
132. Philipov JP, Pascalev MD, Aminkov BY, Grosev CD. Changes in serum carboxyterminal telopeptide of type I collagen in an experimental model of canine osteomyelitis. *Calcif Tissue Int* 1995;57:152-154.
133. Polak-Jonkisz D, Zwolinska D, Bednorz R, Owczarek H, Szymanska A, Nahaczewska W. Procollagen I carboxyterminal propeptide (PICP) as a bone formation marker and carboxyterminal telopeptide of type I collagen (ICTP) as a bone degradation marker in children with chronic renal failure under conservative therapy. *Med Sci Monit* 2003;9:19-23.

134. Quinn MK, Ehrhart N, Johnson AL, Schaeffer DJ. Realignment of the radius in canine antebrachial growth deformities treated with corrective osteotomy and bilateral (type II) external fixation. *Vet Surg* 2000;29:558-563.
135. Ramadan RO, Vaughan LC. Premature closure of the distal ulnar growth plate in dogs - a review of 58 cases. *J Small Anim Pract* 1978;19:647-667.
136. Raschke MJ, Bail H, Windhagen HJ, Kolbeck SF, Weiler A, Raun K, Kappelgard A, Skiaerbaek C, Haas NP. Recombinant growth hormone accelerates bone regenerate consolidation in distraction osteogenesis. *Bone* 1999;24:81-88.
137. Rauch F, Lauzier D, Croteau S, Travers R, Glorieux FH, Hamdy R. Temporal and spatial expression of bone morphogenetic protein-2, -4, and -7 during distraction osteogenesis in rabbits. *Bone* 2000;27:453-459.
138. Rauch F, Lauzier D, Travers R, Glorieux F, Hamdy R. Effects of locally applied transforming growth factor-beta1 on distraction osteogenesis in a rabbit limb-lengthening model. *Bone* 2000;26:619-624.
139. Robey PG. Vertebrate mineralized matrix proteins: structure and function. *Connect Tissue Res* 1996;35:131-136.
140. Robins G. The management of distal radial growth plate closure. *Austr Vet Pract* 1987;17:143-144.
141. Romas E, Gillespie MT, Martin TJ. Involvement of receptor activator of NFkappaB ligand and tumor necrosis factor-alpha in bone destruction in rheumatoid arthritis. *Bone* 2002;30:340-346.
142. Rowe NM, Mehrara BJ, Luchs JS, Dudziak ME, Steinbrech DS, Illei PB, Fernandez GJ, Gittes GK, Longaker MT. Angiogenesis during mandibular distraction osteogenesis. *Ann Plast Surg* 1999;42:470-475.
143. Sabharwal S, Paley D, Bhave A, Herzenberg JE. Growth patterns after lengthening of congenitally short lower limbs in young children. *J Pediatr Orthop* 2000;20:137-145.
144. Salmeri KR, Bloomberg MS, Scruggs SL, Shille V. Gonadectomy in immature dogs: effects on skeletal, physical, and behavioral development. *J Am Vet Med Assoc* 1991;198:1193-1203.
145. Salter RB, Harris WR. Injuries involving the epiphyseal plate. *J Bone Joint Surg Am* 1963;45:587-622.
146. Sartorio A, Ortolani S, Galbiati E, Conte G, Vangeli V, Arosio M, Porretti S, Faglia G. Effects of 12-month GH treatment on bone metabolism and bone mineral density in adults with adult-onset GH deficiency. *J Endocrinol Invest* 2001;24:224-230.
147. Sato M, Ochi T, Nakase T, Hirota S, Kitamura Y, Nomura S, Yasui N. Mechanical tension-stress induces expression of bone morphogenetic protein (BMP)-2 and BMP-4, but not BMP-6, BMP-7, and GDF-5 mRNA, during distraction osteogenesis. *J Bone Miner Res* 1999;14:1084-1095.
148. Sattler AM, Schoppet M, Schaefer JR, Hofbauer LC. Novel aspects on RANK ligand and osteoprotegerin in osteoporosis and vascular disease. *Calcif Tissue Int* 2004;74:103-106.
149. Schmidmaier G, Wildemann B, Heeger J, Gabelein T, Flyvbjerg A, Bail HJ, Raschke M. Improvement of fracture healing by systemic administration of growth hormone and local application of insulin-like growth factor-1 and transforming growth factor-beta1. *Bone* 2002;31:165-172.
150. Schoenmakers I, Hazewinkel HAW, Voorhout G, Carlson CS, Richardson D. Effect of diets with different calcium and phosphorus contents on the skeletal development and blood chemistry of growing Great Danes. *Vet Rec* 2000;147:652-660.
151. Sciadini MF, Dawson JM, Banit D, Juliao SF, Johnson KD, Lennington WJ, Schwartz HS. Growth factor modulation of distraction osteogenesis in a segmental defect model. *Clin Orthop* 2000;381:266-277.

152. Seeherman H, Wozney JM. Delivery of bone morphogenetic proteins for orthopedic tissue regeneration. *Cytokine Growth Factor Rev* 2005;16:329-345.
153. Seibel MJ. Molecular markers of bone turnover: biochemical, technical and analytical aspects. *Osteoporos Int* 2000;11 Suppl 6:18-29.
154. Seibel MJ, Woitge HW. Basic principles and clinical applications of biochemical markers of bone metabolism: biochemical and technical aspects. *J Clin Densitom* 1999;2:299-321.
155. Shevtsov VI, Asonova SN. Ultrastructural changes of articular cartilage following joint immobilization with the Ilizarov apparatus. *Bull Hosp Jt Dis* 1995;54:69-75.
156. Shih LY, Shih HN, Chen TH. The effects of sex and estrogen therapy on bone ingrowth into porous coated implant. *J Orthop Res* 2003;21:1033-1040.
157. Sietsema WK. Animal models of cortical porosity. *Bone* 1995;17:297-305.
158. Sjogren K, Jansson JO, Isaksson OG, Ohlsson C. A model for tissue-specific inducible insulin-like growth factor-I (IGF-I) inactivation to determine the physiological role of liver-derived IGF-I. *Endocrine* 2002;19:249-256.
159. Sneppen SB, Hoeck HC, Kollerup G, Sorensen OH, Laurberg P, Feldt-Rasmussen U. Bone mineral content and bone metabolism during physiological GH treatment in GH-deficient adults--an 18-month randomised, placebo-controlled, double blinded trial. *Eur J Endocrinol* 2002;146:187-195.
160. Sojo K, Sawaki Y, Hattori H, Mizutani H, Ueda M. Immunohistochemical study of vascular endothelial growth factor (VEGF) and bone morphogenetic protein-2, -4 (BMP-2, -4) on lengthened rat femurs. *J Craniomaxillofac Surg* 2005;33:238-245.
161. Spector JA, Luchs JS, Mehrara BJ, Greenwald JA, Smith LP, Longaker MT. Expression of bone morphogenetic proteins during membranous bone healing. *Plast Reconstr Surg* 2001;107:124-134.
162. Stanitski DF, Shahcheraghi H, Nicker DA, Armstrong PF. Results of tibial lengthening with the Ilizarov technique. *J Pediatr Orthop* 1996;16:168-172.
163. Steinbrech DS, Mehrara BJ, Rowe NM, Dudziak ME, Luchs JS, Saadeh PB, Gittes GK, Longaker MT. Gene expression of TGF-beta, TGF-beta receptor, and extracellular matrix proteins during membranous bone healing in rats. *Plast Reconstr Surg* 2000;105:2028-2038.
164. Steinbrech DS, Mehrara BJ, Rowe NM, Dudziak ME, Saadeh PB, Gittes GK, Longaker MT. Gene expression of insulin-like growth factors I and II in rat membranous osteotomy healing. *Ann Plast Surg* 1999;42:481-487.
165. Stewart KJ, Weyand B, van't Hof RJ, White SA, Lvoff GO, Maffulli N, Poole MD. A quantitative analysis of the effect of insulin-like growth factor-1 infusion during mandibular distraction osteogenesis in rabbits. *Br J Plast Surg* 1999;52:343-350.
166. Suda T, Ueno Y, Fujii K, Shinki T. Vitamin D and bone. *J Cell Biochem* 2003;88:259-266.
167. Syed F, Khosla S. Mechanisms of sex steroid effects on bone. *Biochem Biophys Res Commun* 2005;328:688-696.
168. Szulc P, Seeman E, Delmas PD. Biochemical measurements of bone turnover in children and adolescents. *Osteoporos Int* 2000;11:281-294.
169. Takahashi N, Udagawa N, Suda T. A new member of tumor necrosis factor ligand family, ODF/OPGL/TRANCE/RANKL, regulates osteoclast differentiation and function. *Biochem Biophys Res Commun* 1999;256:449-455.
170. Tavakoli K, Yu Y, Shahidi S, Bonar F, Walsh WR, Poole MD. Expression of growth factors in the mandibular distraction zone: a sheep study. *Br J Plast Surg* 1999;52:434-439.
171. Theoleyre S, Wittrant Y, Tat SK, Fortun Y, Redini F, Heymann D. The molecular triad OPG/RANK/RANKL: involvement in the orchestration of pathophysiological bone remodeling. *Cytokine Growth Factor Rev* 2004;15:457-475.

172. Thoren M, Hilding A, Brismar T, Magnusson P, Degerblad M, Larsson L, Saaf M, Baylink DJ, Mohan S. Serum levels of insulin-like growth factor binding proteins (IGFBP)-4 and -5 correlate with bone mineral density in growth hormone (GH)- deficient adults and increase with GH replacement therapy. *J Bone Miner Res* 1998;13:891-899.
173. Torres R, de la PC, Rapado A. Clinical usefulness of serum tartrate-resistant acid phosphatase in Paget's disease of bone: correlation with other biochemical markers of bone remodelling. *Calcif Tissue Int* 1991;49:14-16.
174. Tryfonidou MA, Holl MS, Vastenburg M, Oosterlaken-Dijksterhuis MA, Birkenhager-Frenkel DH, van den Brom WE, Hazewinkel HA. Hormonal regulation of calcium homeostasis in two breeds of dogs during growth at different rates. *J Anim Sci* 2003;81:1568-1580.
175. Tsumaki N, Yoshikawa H. The role of bone morphogenetic proteins in endochondral bone formation. *Cytokine Growth Factor Rev* 2005;16:279-285.
176. Turner RT, Maran A, Lotinun S, Hefferan T, Evans GL, Zhang M, Sibonga JD. Animal models for osteoporosis. *Rev Endocr Metab Disord* 2001;2:117-127.
177. Ueland T. Bone metabolism in relation to alterations in systemic growth hormone. *Growth Horm IGF Res* 2004;14:404-417.
178. Ueland T. GH/IGF-I and bone resorption in vivo and in vitro. *Eur J Endocrinol* 2005;152:327-332.
179. Urist MR, DeLange RJ, Finerman GA. Bone cell differentiation and growth factors. *Science* 1983;220:680-686.
180. Vaananen HK, Zhao H, Mulari M, Halleen JM. The cell biology of osteoclast function. *J Cell Sci* 2000;113:377-381.
181. Valentin-Opran A, Wozney J, Csimma C, Lilly L, Riedel GE. Clinical evaluation of recombinant human bone morphogenetic protein-2. *Clin Orthop* 2002;395:110-120.
182. van Coeverden SC, Netelenbos JC, de Ridder CM, Roos JC, Popp-Snijders C, Delemarre-van-de-Waal HA. Bone metabolism markers and bone mass in healthy pubertal boys and girls. *Clin Endocrinol (Oxf)* 2002;57:107-116.
183. Vandewater A, Olmstead ML. Premature closure of the distal radial physis in the dog. A review of eleven cases. *Vet Surg* 1983;12:7-12.
184. Varga AC, Wrana JL. The disparate role of BMP in stem cell biology. *Oncogene* 2005;24:5713-5721.
185. Vidal J. External fixation. Yesterday, today, and tomorrow. *Clin Orthop Relat Res* 1983;7-14.
186. Walsh WR, Hamdy RC, Ehrlich MG. Biomechanical and physical properties of lengthened bone in a canine model. *Clin Orthop* 1994;306:230-238.
187. Wan M, Cao X. BMP signaling in skeletal development. *Biochem Biophys Res Commun* 2005;328:651-657.
188. Wang J, Zhou J, Cheng CM, Kopchick JJ, Bondy CA. Evidence supporting dual, IGF-I-independent and IGF-I-dependent, roles for GH in promoting longitudinal bone growth. *J Endocrinol* 2004;180:247-255.
189. Wei S, Lightwood D, Ladyman H, Cross S, Neale H, Griffiths M, Adams R, Marshall D, Lawson A, McKnight AJ, Stanley ER. Modulation of CSF-1-regulated post-natal development with anti-CSF-1 antibody. *Immunobiology* 2005;210:109-119.
190. Weiss S, Baumgart R, Jochum M, Strasburger CJ, Bidlingmaier M. Systemic regulation of distraction osteogenesis: a cascade of biochemical factors. *J Bone Miner Res* 2002;17:1280-1289.
191. Wilkens BE, Millis DL, Daniel GB, Munson L, Patel KR, Buonomo FC. Metabolic and histologic effects of recombinant canine somatotropin on bone healing in dogs, using an unstable osteotomy gap model. *Am J Vet Res* 1996;57:1395-1401.

192. Willenegger H, Perren SM, Schenk R. [Primary and secondary healing of bone fractures]. *Chirurg* 1971;42:241-252.
193. Wilson AK, Bhattacharyya MH, Miller S, Mani A, Sacco-Gibson N. Ovariectomy-induced changes in aged beagles: histomorphometry of rib cortical bone. *Calcif Tissue Int* 1998;62:237-243.
194. Yao GQ, Sun BH, Weir EC, Insogna KL. A role for cell-surface CSF-1 in osteoblast-mediated osteoclastogenesis. *Calcif Tissue Int* 2002;70:339-346.
195. Yeung HY, Lee SK, Fung KP, Leung KS. Expression of basic fibroblast growth factor during distraction osteogenesis. *Clin Orthop Relat Res* 2001;385:219-229.
196. Zangger I, Zapf J, Froesch ER. Insulin-like growth factor I and II in 14 animal species and man as determined by three radioligand assays and two bioassays. *Acta Endocrinol (Copenh)* 1987;114:107-112.
197. Zimmermann G, Henle P, Kusswetter M, Moghaddam A, Wentzensen A, Richter W, Weiss S. TGF-beta1 as a marker of delayed fracture healing. *Bone* 2005;36:779-785.