Synthesis of three tetrasaccharides containing 3-O-methyl-D-mannose, as model compounds for xylose-containing carbohydrate chains from N-glycoproteins

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ABSTRACT

The synthesis is reported of methyl 3,6-di-O-(3-O-methyl- α -D-mannopyranosyl)-2- $O-\beta$ -Dxylopyranosyl- β -D-mannopyranoside (2), methyl 6-O- α -D-mannopyranosyl-3-O-(3-O-methyl- α -D-mannopyranosyl)-2-O- β -D-xylopyranosyl- β -D-mannopyranoside (3), and methyl 3-O- α -D-mannopyranosyl-6- $O-(3-O-\text{methyl}-\alpha-D-\text{mannopyranosyl})-2-O-\beta-D-xylopyranosyl-\beta-D-mannopyranoside (4).$ The various methyl β -D-Man p acceptor derivatives were prepared from the corresponding methyl β -D-Glc p derivatives via oxidation-reduction. All glycosyl donors were coupled using the trichloroacetimidate method at -40° C in dichloromethane with trimethylsilyl triflate as a catalyst. Methyl-3-O-benzyl-4,6-O-benzylidene- β -D-mannopyranoside (7) was condensed with 2,3,4-tri-O-acetyl- α -D-xylopyranosyl trichloroacetimidate (8). Regioselective reductive 4,6-O-benzylidene ring-opening on the resulting disaccharide derivative, followed by acetylation, and hydrogenation gave methyl 4-O-acetyl-2-O-(2,3,4tri-O-acetyl-B-D-xylopyranosyl)-B-D-mannopyranoside (12). Coupling of 12 with 2,4,6-tri-O-acetyl-3-Omethyl- α -D-mannopyranosyl trichloroacetimidate (18) afforded tetrasaccharide derivative 19, and subsequent O-deacetylation gave 2. Methyl 3-O-benzyl-4.6-O-prop-2-enylidene-B-D-mannopyranoside (22) was condensed with 2,3,4-tri-O-acetyl-a-D-xylopyranosyl trichloroacetimidate (8). Regioselective reductive 4,6-O-prop-2-enylidene ring-opening on the resulting disaccharide derivative, followed by acetylation, and deallylation at O-6 gave methyl 4-O-acetyl-3-O-benzyl-2-O-(2,3,4-tri-O-acetyl- β -Dxylopyranosyl)- β -D-mannopyranoside (26-a), which was either condensed with 2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl trichloroacetimidate (27) or 18, to give trisaccharide derivatives 28 or 31, respectively. Debenzylation of 28 followed by condensation with 18 gave, after O-deacetylation, 3, whereas debenzylation of 31 followed by condensation with 27 gave, after O-deacetylation, 4.

INTRODUCTION

Methylated monosaccharide residues only rarely occur in N-linked carbohydrate chains of glycoproteins. So far, 3-O-methyl-D-mannose has been found in hemocyanin of Lymnaea stagnalis^{1,2} and in N-glycoproteins of the dimorphic fungus

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*Mucor rouxii*³. In the hemocyanin, 3-O-methyl- α -D-mannose forms an integral part of the core element which also contains a β -D-xylose residue $(1 \rightarrow 2)$ -linked to β -D-mannose^{1,2,4}, whereas novel extensions, including the occurrence of 3-O-methyl- β -D-galactose, were established² (1).

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(*i*)
$$R^1 = R^2 = 3 - O - Me$$

- (*ii*) $R^1 = 3-O-Me-\beta-D-Gal p-(1 \rightarrow 3)-\beta-D-Gal pNAc-(1 \rightarrow 4)-\beta-D-Glc pNAc-(1 \rightarrow 2)$ and $R^2 = 3-O-Me$
- (*iii*) $R^1 = H$ and $R^2 = 3$ -O-Me- β -D-Galp-(1 \rightarrow 3)- β -D-GalpNAc-(1 \rightarrow 4)- β -D-GlcpNAc-(1 \rightarrow 2)
- (iv) $R^1 = R^2 = 3 \cdot O \cdot Me \cdot \beta \cdot D \cdot Gal p \cdot (1 \rightarrow 3) \cdot \beta \cdot D \cdot Gal p \cdot NAc \cdot (1 \rightarrow 4) \cdot \beta \cdot D \cdot Glc p \cdot NAc \cdot (1 \rightarrow 2)$
- (v) $R^1 = \alpha$ -L-Fuc p-(1 \rightarrow 2)- β -D-Gal p-(1 \rightarrow 3)- β -D-Gal pNAc-(1 \rightarrow 4)- β -D-Glc pNAc-(1 \rightarrow 2) and $R^2 = 3$ -O-Me
- (vi) $R^1 = H$ and $R^2 = \alpha - L - Fuc p - (1 \rightarrow 2) - \beta - D - Gal p - (1 \rightarrow 3) - \beta - D - Gal p NAc - (1 \rightarrow 4) - \beta - D - Glc p NAc - (1 \rightarrow 2)$
- $(v\ddot{u}) \quad R^{1} = R^{2} = \alpha \text{-L-Fuc} p \text{-} (1 \rightarrow 2) \text{-}\beta \text{-D-Gal} p \text{-} (1 \rightarrow 3) \text{-}\beta \text{-} \text{D-Gal} p \text{NAc-} (1 \rightarrow 4) \text{-}\beta \text{-} \text{D-Glc} p \text{NAc-} (1 \rightarrow 2)$
- (viii) $R^1 = 3$ -O-Me- β -D-Galp- $(1 \rightarrow 3)$ - β -D-GalpNAc- $(1 \rightarrow 4)$ - β -D-GlcpNAc- $(1 \rightarrow 2)$ and $R^2 = \alpha$ -L-Fucp- $(1 \rightarrow 2)$ - β -D-Galp- $(1 \rightarrow 3)$ - β -D-GalpNAc- $(1 \rightarrow 4)$ - β -D-GlcpNAc- $(1 \rightarrow 2)$

As a part of our program concerning the biosynthesis and conformation analysis of these types of glycans⁵⁻⁷, the synthesis of some related oligosaccharide elements has been reported^{8,9}. Here, we describe the synthesis of the tetrasaccharides 2, 3, and 4, containing either one or two 3-O-methyl- α -D-mannose residues, representing the branching part of the core structure in which the β -D-mannose configuration is mimicked as a methyl glycoside. It should be noted that 4 is not a structural element of *L. stagnalis* hemocyanin.

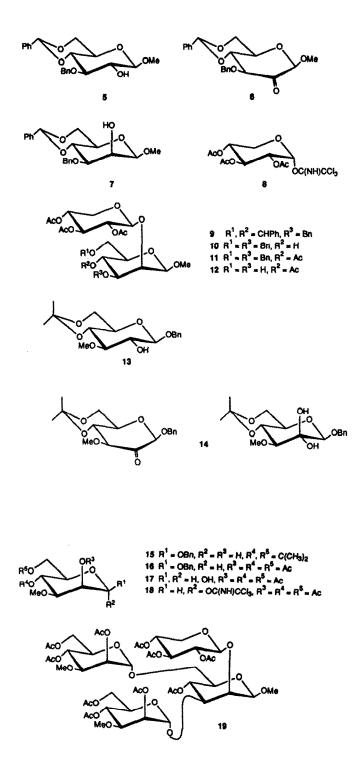
RESULTS AND DISCUSSION

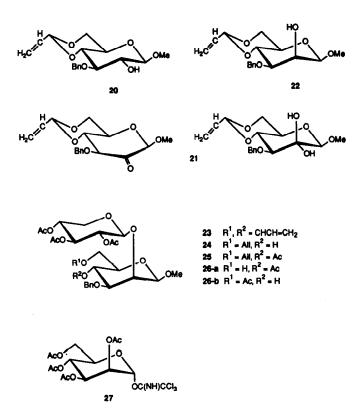
For the synthesis of tetrasaccharide 2, methyl 4-O-acetyl-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranoside (12) and 2,4,6-tri-O-acetyl-3-O-methyl- α -D-mannopyranosyl trichloroacetimidate (18) where chosen as building blocks. The

preparation of methyl β -D-Man p, or β -D-mannopyranosides in general, can be accomplished via 'direct' or 'indirect' methods⁸⁻¹⁰. Because the synthesis of xylose-containing carbohydrate chains using the indirect method via oxidation-reduction has proved to be very convenient in our hands^{8,9}, this strategy was also followed in the present report. Thus, methyl 3-O-benzyl-B-D-glucopyranoside¹¹ was benzylidenated using α, α -dimethoxytoluene and p-toluenesulfonic acid to give crystalline 5 (75%). Oxidation of 5 using 2:1 dimethyl sulfoxide-acetic anhydride gave 6 and a small amount of a by-product. Evaporation of the reagents followed by treatment of the residue with sodium borohydride afforded 7 (66% from 5) and some 5 (10% from 5). The by-product in the oxidation of 5, probably methyl 3-O-benzyl-4,6-O-benzylidene-2-O-methylthiomethyl- β -D-glucopyranoside¹², could easily be separated from the reaction products after the reduction. Its amount can be minimized by using acetic anhydride free from acetic acid¹³. Stereoselective xvlosvlation of 7 proceeded satisfactorily using 2.3.4-tri-O-acetyl- α -D-xvlopyranosvl trichloroacetimidate (8)^{9,14} in dichloromethane at -40° C, with trimethylsilyl triflate as a catalyst, to give exclusively the β -linked disaccharide derivative 9 in 75% yield. Regioselective reductive ring-opening of the 4,6-benzylidene acetal, using sodium cyanoborohydride and hydrochloric acid in tetrahydrofuran¹⁵ (\rightarrow 10, 67%). followed by treatment with pyridine-acetic anhydride (\rightarrow 11, 99%) and subsequent hydrogenation using 10% Pd-C gave 12 in 90% yield.

The synthesis of the 3-O-methyl-D-mannose donor (18) involved also the application of the oxidation-reduction method, whereby benzyl 4,6-O-isopropylidene-3-O-methyl- β -D-glucopyranoside (13) was converted into the corresponding manno derivative (15). Isopropylidenation of benzyl 3-O-methyl- β -D-glucopyranoside¹⁶, using 2,2-dimethoxypropane and p-toluenesulfonic acid, yielded crystalline 13 (88%), which was oxidised using 2:1 dimethyl sulfoxide-acetic anhydride to give 14 in its keto and gem-diol form (79%), as shown by ¹³C NMR spectroscopy. Reduction of this mixture with sodium borohydride afforded 15 (79%) and some 13 (7%). Then compound 15 was de-isopropylidenated and acetylated (\rightarrow 16, 93%), hydrogenolysed (\rightarrow 17, 97%), and treated with trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene¹⁷ to afford 18 (73%). Coupling of 18 with 12 in dichloromethane (1 h, -40°C), using trimethylsilyl triflate as a catalyst, gave tetrasaccharide derivative 19 in 71% yield. Finally, Zemplén deacetylation of 19 afforded 2 (90%). For ¹H NMR structural-reported-group data, see Table I.

For the synthesis of the tetrasaccharides 3 and 4, having a 3-O-methyl- α -D-mannose residue linked either $1 \rightarrow 3$ or $1 \rightarrow 6$, the 4,6-benzylidene acetal in compound 9 was changed for a 4,6-(prop-2-enylidene) acetal¹⁸. Regioselective reductive ring-opening and acetylation gives a synthon with an allyl group at O-6 and a benzyl group at O-3, making the regiospecific introduction of 3-O-methyl- α -D-mannose and α -D-mannose residues possible. To this end, methyl 3-O-benzyl- β -D-glucopyranoside¹¹ was treated with acrolein dimethyl acetal and *p*-toluenesulfonic acid in *N*,*N*-dimethylformamide¹⁸ to give crystalline 20 (75%). Oxidation of 20 using 2:1 dimethyl sulfoxide-acetic anhydride gave 21, in its keto and gem-diol





form. After evaporation of the reagents, the residue, still containing a small amount of by-product (see above), was directly treated with sodium borohydride to afford 22 (56% from 20) and 20 (11% from 20). Xylosylation of 22 with 8 in dichloromethane at -40° C, using trimethylsilyl triflate as a catalyst, gave 23 (89%). Regioselective reductive ring-opening of the 4,6-(prop-2-enylidene) acetal, using sodium cyanoborohydride and hydrochloric acid in tetrahydrofuran (\rightarrow 24, 69%), followed by treatment with pyridine-acetic anhydride gave 25 (98%). De-allylation of 25 using the Wilkinson catalyst in the presence of 1,4-diazabicyclo[2.2.2]octane¹⁹ followed by hydrolysis yielded one spot in TLC (26-a). However, during column chromatography, acetyl migration in the β -D-mannose residue occurred from O-4 \rightarrow O-6, and the 4-O-acetyl derivative 26-a could be isolated in only 53% yield, together with the undesired 6-O-acetyl derivative 26-b (40%). This problem could be overcome by direct glycosylation after work-up, as is shown for the synthesis of 31.

Condensation of compound 26-a with 27 in dichloromethane (1 h, -40° C), using trimethylsilyl triflate as a catalyst ($\rightarrow 28$, 77%), followed by hydrogenolysis of the benzyl ether of the resulting trisaccharide derivative gave acceptor 29 (98%), which was mannosylated with 18 in dichloromethane (trimethylsilyl triflate, 1.5 h,

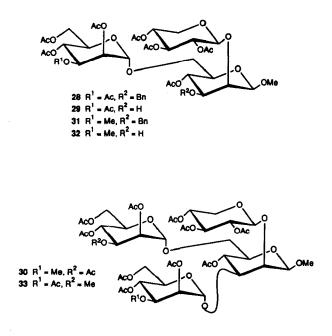
Residue	Reporter group (J)	δ (ppm)/J (Hz)				
		2	3	4	R	
β-Man	H-1 ª	4.689	4.682	4.687	4.681	
	H-2 $(J_{2,3})$	4.203 (2.3)	4.197 (2.1)	4.200 (2.3)	4.197 (2.8)	
	CH ₃ O	3.532	3.526	3.531	3.523	
Xyl	H-1 (J _{1,2})	4.488 (7.4)	4.482 (7.4)	4.483 (7.4)	4.474 (7.5)	
α -Man-(1 \rightarrow 6)	H-1 $(J_{1,2})$	4.948 (1.8)	4.903 (1.7)	4.945 (1.8)	4.900 (1.8)	
	$H-2(J_{2,3}^{1,2})$	4.263 (3.2)	n.d. ^b	4.258 (3.0)	4.010 (3.5)	
	CH ₃ O ^{2,3}	3.459		3.457		
α -Man-(1 \rightarrow 3)	H-1 (J _{1,2})	5.183 (1.5)	5.180 (1.5)	5.141 (1.4)	5.138 (1.8)	
	H-2 $(J_{2,3})$	4.303 (2.9)	4.294 (n.d.)	4.043 (3.3)	4.035 (3.4)	
	CH ₃ O	3.445	3.445			

TABLE I

300-MHz ¹H NMR data of the tetrasaccharides 2-4, together with those of reference compound methyl 3,6-di-O- α -D-mannopyranosyl-2-O- β -D-xylopyranosyl- β -D-mannopyranoside⁸ (**R**)

^a Because of the very small $J_{1,2}$ value, H-1 is observed as a singlet. ^b Not determined.

 -40° C) to afford tetrasaccharide derivative **30** (61%). Zemplén deacetylation then gave compound **3** (96%). Trisaccharide derivative **31** was prepared in 51% yield by condensation of non-purified **26-a** (see above) with **18** in dichloromethane, using trimethylsilyl triflate as a catalyst (1 h, -40° C). Hydrogenolysis of **31** (\rightarrow **32**, 98%) and subsequent glycosylation of **32** with **27** gave tetrasaccharide derivative **33** (70%), which after Zemplén deacetylation afforded **4** (96%). For ¹H NMR structural-reporter-group data of **3** and **4**, see Table I.



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Compound	J _{1,2}	J _{2,3}	J _{3,4}	J _{4,5eq}	J _{4,5ax}	J _{5,5}
9	6.1	8.1	7.9	4.7	7.5	- 12.0
10	6.3	8.0	8.0	4.8	8.0	- 11.9
12	6.9	n.d. <i>a</i>	n.d.	5.1	n.d.	- 11.8
19	7.1	9.1	n.d.	5.6	9.0	- 12.3
23	5.6	7.7	7.7	4.6	7.4	-12.0
24	6.3	8.2	8.0	4.8	8.0	- 12.9
25	n.d.	n.d.	n.d.	3.4	n.d.	-12.0
26-a	n.d.	n.d.	n.d.	3.5	4.7	-12.4
26-b	7.0	9.0	8.8	5.1	8.8	-11.8
28	n.d.	n.d.	n.d.	3.5	4.9	- 12.3
29	6.7	8.6	n.d.	n.d.	8.8	-11.9
30	n.d.	n.d.	n.d.	5.2	8.2	- 11.6
31	4.7	6.5	n.d.	5.5	n.d.	-12.3
33	5.8	n.d.	7.1	4.4	7.1	n.d.

TABLE II

Coupling constants (J, Hz) between the skeleton protons of the xylose residue in protected oligosaccharides synthesised in this study

^a Not determined.

Evaluation of the ¹H NMR spectra of the compounds containing a protected β -D-xylopyranose residue showed that the J values between the skeleton protons vary to some extent (Table II). Taking into account especially the $J_{1,2}$, $J_{4,5ax}$, and $J_{4,5eq}$ values, it can be concluded that these variations have to reflect changes in conformation of the xylose residue. This behaviour is not uncommon for protected β -D-Xylp derivatives, and quite often ${}^{1}C_{4}(D) \Leftrightarrow {}^{4}C_{1}(D)$ conformational equilibria are suggested in case of small $J_{1,2}/J_{4,5ax}/J_{4,5eq}$ values, explained by the anomeric effect which balances the destabilising effect of the three axially oriented O-acetyl groups on the Xylp ring^{9,20-26}. In this context it has to be mentioned that distortions of the ${}^{4}C_{1}(D)$ chair can also lead to deviating J values.

EXPERIMENTAL

General methods.—The ¹H (300 and 360 MHz) and ¹³C (APT, 50 MHz) NMR spectra were recorded at 25°C with a Bruker AC 300, WP 200, or HX 360 spectrometer. Chemical shifts (δ) are given in ppm relative to the signal for internal Me₄Si (CDCl₃) or sodium 4,4-dimethyl-4-silapentane-1-sulfonate (D₂O; indirectly to internal acetone, δ 2.225) for ¹H, and to the signal for internal Me₄Si (CDCl₃; indirectly to CDCl₃, δ 76.9) or external Me₄Si (D₂O; indirectly to internal acetone, δ 31.55) for ¹³C. Column chromatography was performed on Kieselgel 60 (Merck, < 230 mesh) and fractions were monitored by TLC on Kieselgel 60 F₂₅₄ (Merck) by detection with UV light and then charring with H₂SO₄. Optical rotations were measured for solutions in CHCl₃, unless otherwise stated, at 20°C with a Perkin–Elmer 241 polarimeter, using a 10-cm 1-mL cell. Melting points were determined with a Mettler FP 51 instrument. In the work-up procedures, washings were carried out three times with appropriate quantities of water or aq 5% NaHCO₃ unless indicated otherwise. Solvents were evaporated under reduced pressure at 40°C (bath) unless indicated otherwise. All solvents were distilled from the appropriate drying agents.

Methyl 3-O-benzyl-4,6-O-benzylidene- β -D-glucopyranoside (5).—To a solution of methyl 3-O-benzyl- β -D-glucopyranoside¹¹ (1.8 g, 6.3 mmol) in N,N-dimethylformamide (17 mL) and α, α -dimethoxytoluene (1.8 mL, 12 mmol) was added ptoluenesulfonic acid monohydrate (150 mg). After stirring for 1 h under reduced pressure (20 mmHg), TLC (95:5 CH₂Cl₂-acetone) indicated the reaction to be complete, and solid NaHCO₃ was added. The mixture was diluted with CH₂Cl₂ (300 mL), washed with water, dried (MgSO₄), filtered, and concentrated. The residue was crystallised from EtOH to give 5 (1.8 g, 75%); mp 181°C; $[\alpha]_D - 38^\circ$ (c 1); R_f 0.60; lit²⁷ mp 182–183°C; $[\alpha]_D = 51^\circ$ (c 0.86, CHCl₃). NMR data (CDCl₃): ¹H, δ 7.48–7.25 (m, 10 H, 2 Ph), 5.576 (s, 1 H, PhCH), 4.974 and 4.792 (2 d, each 1 H, PhC H_2 O), 4.364 (dd, 1 H, $J_{6a,5}$ 5.0, $J_{6a,6b}$ – 10.5 Hz, H-6a), 4.328 (d, 1 H, $J_{1,2}$ 7.6 Hz, H-1), 3.804 (t, 1 H, $J_{4,3} = J_{4,5} = 10.2$ Hz, H-4), 3.572 (s, 3 H, CH₃O), 2.442 (d, 1 H, $J_{HO-2.2}$ 2.2 Hz, HO-2); ¹³C, δ 137.1, 128.8–127.7, and 125.9 ($C_6H_5CH_2O$ and C₆H₅CHO), 104.1 (PhCH), 101.1 (C-1), 81.3, 80.1, 74.1, and 66.2 (C-2,3,4,5), 74.5 and 68.6 (PhCH₂O and C-6), 57.3 (CH₃O). Anal. Calcd for C₂₁H₂₄O₆: C, 67.73; H, 6.50. Found: C, 67.67; H, 6.65.

Methyl 3-O-benzyl-4,6-O-benzylidene-β-D-mannopyranoside (7).—A solution of 5 (1.2 g, 3.2 mmol) in 1:2 acetic anhydride–Me₂SO (22 mL) was stirred for 20 h. TLC (95:5 CH₂Cl₂-acetone) then showed the ulose **6** (R_f 0.50–0.72) and a faster moving product (R_f 0.93). The mixture was concentrated, and toluene (3 × 15 mL) was evaporated from the residue, which was then further concentrated in vacuo (bath < 60°C) to yield **6**, isolated as a yellow solid (1.2 g). A purified sample gave: $[\alpha]_D - 80^\circ$ (c 1); lit²⁷ $[\alpha]_D - 81^\circ$ (c 1, CHCl₃). NMR data (CDCl₃): ¹H, δ 7.51–7.30 (m, 10 H, 2 Ph), 5.587 (s, 1 H, PhCH), 4.970 and 4.764 (2 d, each 1 H, PhCH₂O), 4.780 (s, 1 H, H-1), 4.460 (dd, 1 H, $J_{6a,5}$ 4.7, $J_{6a,6b} - 10.3$ Hz, H-6a), 4.240 (dd, 1 H, $J_{3,4}$ 10.3 Hz, H-3), 3.761 (m, 1 H, H-5), 3.601 (s, 3 H, CH₃O); ¹³C, δ 188.1 (C-2), 137.1, 136.7, 129.0–127.8, and 125.9 ($C_6H_5CH_2O$ and C_6H_5CHO), 101.4 and 100.9 (C-1 and PhCH), 81.9, 81.8, and 66.3 (C-3,4,5), 73.1 and 68.4 (PhCH₂O and C-6), 56.9 (CH₃O).

To a solution of 6 (1.0 g, 2.7 mmol) in 1:1 CH₂Cl₂-MeOH (20 mL) containing Kieselgel 60 (70-230 mesh, 0.5 g) was added NaBH₄ (0.4 g, 11 mmol) in portions during 30 min at 0°C. After 30 min, TLC (95:5 CH₂Cl₂-acetone) showed the formation of 5 (R_f 0.60) and 7 (R_f 0.41) in the ratio ~1:7. The mixture was diluted with CH₂Cl₂ (250 mL), filtered, washed with water, dried (MgSO₄), filtered, and concentrated. Column chromatography (95:5 CH₂Cl₂-acetone) of the residue gave 5 (100 mg, 10%), and then 7 (660 mg, 66%), both isolated as white solids (yields calculated from 5). Compound 7: mp 119°C (from EtOH); [α]_D - 28° (c 1); lit²⁷ mp 119-120°C (EtOH); [α]_D - 33° (c 0.86, CHCl₃). NMR data

(CDCl₃): ¹H, δ 7.52–7.20 (m, 10 H, 2 Ph), 5.614 (s, 1 H, PhC*H*), 4.871 and 4.777 (2 d, each 1 H, PhC*H*₂O), 4.431 (bs, 1 H, H-1), 4.344 (dd, 1 H, *J*_{6a,5} 4.9, *J*_{6a,6b} –10.5 Hz, H-6a), 3.657 (dd, 1 H, *J*_{3,2} 3.3, *J*_{3,4} 9.6 Hz, H-3), 3.565 (s, 3 H, CH₃O), 3.355 (m, 1 H, H-5), 2.529 (d, 1 H, *J*_{HO-2,2} 1.2 Hz, HO-2); ¹³C, δ 137.8, 137.3, 128.8–127.7, and 125.9 (*C*₆H₅CH₂O and *C*₆H₅CH), 101.3 (C-1 and Ph*C*H), 78.3, 76.5, 69.7, and 66.7 (C-2,3,4,5), 72.4 and 68.4 (C-6 and Ph*C*H₂O), 57.1 (CH₃O). Anal. Calcd for C₂₁H₂₄O₆: C, 67.73; H, 6.50. Found: C, 67.48; H, 6.61.

Methyl 3-O-benzyl-4,6-O-benzylidene-2-O- $(2,3,4-tri-O-acetyl-\beta-D-xylopyranosyl)$ β-D-mannopyranoside (9).-A solution of 7 (568 mg, 1.5 mmol) and 2,3,4-tri-Oacetyl-a-D-xylopyranosyl trichloroacetimidate¹⁴ (8; 977 mg, 2.3 mmol) in CH₂Cl₂ (10 mL) containing powdered 4A molecular sieves (5.5 g) was stirred for 30 min under N₂. Then a solution of trimethylsilyl triflate (65 μ L) in CH₂Cl₂ (12 mL) was added dropwise at -40° C, and after 2 h, when TLC showed the disappearance of 7 and the formation of 9 (R_f 0.76, 95:5 CH₂Cl₂-acetone), pyridine (1 mL) was added. The mixture was filtered through Celite, then concentrated, and toluene $(3 \times 10 \text{ mL})$ was evaporated from the residue. Two-fold column chromatography [first 95:5 CH₂Cl₂-acetone, then 1:1 light petroleum (bp 40-60°C)-EtOAc] of the residue gave 9, isolated as a white foam (740 mg, 75%); $[\alpha]_D - 96^\circ$ (c 1); R_f 0.46 [1:1 light petroleum (bp 40-60°C)-EtOAc]. NMR data (CDCl₃): ¹H, δ 7.51-7.26 (m, 10 H, 2 Ph), 5.591 (s, 1 H, PhCH), 5.189 (t, 1 H, J_{3',4'} 7.9 Hz, H-3'), 5.058 (dd, 1 H, J_{2',3'} 8.1 Hz, H-2'), 4.933 (m, 1 H, J_{4',5'eq} 4.7 Hz, H-4'), 4.930 (d, 1 H, J_{1',2'} 6.1 Hz, H-1'), 4.763 (s, 2 H, PhCH₂O), 4.315 (s, 1 H, H-1), 4.155 (d, 1 H, J_{2.3} 3.0 Hz, H-2), 3.564 (dd, 1 H, J_{3.4} 9.9 Hz, H-3), 3.462 (s, 3 H, CH₃O), 3.378 (dd, 1 H, $J_{5'ax,4'}$ 7.5, $J_{5'ax,5'eq}$ – 12.0 Hz, H-5'ax), 3.279 (m, 1 H, $J_{5,6a}$ 4.8, $J_{5,6b}$ 9.9 Hz, H-5), 2.074, 2.056, and 2.047 (3 s, each 3 H, 3 Ac); 13 C, δ 169.7 (2 C) and 169.2 (3 $COCH_3$, 138.1, 137.4, 128.7–127.4, and 125.9 ($C_6H_5CH_2O$ and C_6H_5CH), 102.3, 101.3, and 100.6 (C-1,1' and PhCH), 78.0, 76.0, 74.9, 70.3, 70.0, 68.8, and 67.4 (C-2,3,4,5,2',3',4'), 71.5, 68.4, and 61.4 (C-6,5' and PhCH₂O), 56.8 (CH₃O), 20.7 (COCH₃). Anal. Calcd for C₃₂H₃₈O₁₃: C, 60.94; H, 6.07. Found: C, 60.43; H, 6.00.

Methyl 3,6-di-O-benzyl-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranoside (10).—A solution of 9 (627 mg, 1.0 mmol) and sodium cyanoborohydride (750 mg, 12 mmol) in tetrahydrofuran (18 mL) containing 3A molecular sieves (2.0 g) was stirred for 30 min under N₂. Then a saturated solution of HCl in diethyl ether was added until the evolution of gas ceased, and the mixture was stirred for 30 min. TLC (95:5 CH₂Cl₂-acetone) then showed the conversion of 9 (R_f 0.76) into 10 (R_f 0.22). The mixture was diluted with CH₂Cl₂ (200 mL), filtered through Celite, washed with water, aq 5% NaHCO₃, and water, dried (MgSO₄), filtered, and concentrated. Column chromatography (95:5 CH₂Cl₂acetone) of the residue gave 10, isolated as a white solid (421 mg, 67%); [α]_D - 101° (c 1). NMR data (CDCl₃): ¹H, δ 7.38-7.27 (m, 10 H, 2 Ph), 5.175 (t, 1 H, $J_{3',4'}$ 8.0 Hz, H-3'), 4.996 (dd, 1 H, $J_{2',3'}$ 8.0 Hz, H-2'), 4.917 (m, 1 H, $J_{4',5'eq}$ 4.8, $J_{4',5'ax}$ 8.0 Hz, H-4'), 4.902 (d, 1 H, $J_{1',2'}$ 6.3 Hz, H-1'), 4.799 and 4.495 (2 d, each 1 H, PhCH₂O), 4.602 and 4.557 (2 d, each 1 H, PhCH₂O), 4.267 (s, 1 H, H-1), 4.228 (dd, 1 H, $J_{5'eq,5'ax}$ – 11.9 Hz, H-5'eq), 4.150 (d, 1 H, $J_{2,3}$ 2.9 Hz, H-2), 3.821 (dd, 1 H, $J_{6a,5}$ 3.3, $J_{6a,6b}$ – 10.4 Hz, H-6a), 3.743 (m, 1 H, $J_{4,5}$ 9.4 Hz, H-4), 3.658 (dd, 1 H, $J_{6b,5}$ 6.4 Hz, H-6b), 3.472 (s, 3 H, CH₃O), 3.401 (m, 1 H, H-5), 3.356 (dd, 1 H, H-5'ax), 3.289 (dd, 1 H, $J_{3,4}$ 9.4 Hz, H-3), 2.527 (d, 1 H, $J_{HO-4,4}$ 1.9 Hz, HO-4), 2.047, 2.011, and 1.989 (3 s, each 3 H, 3 Ac); ¹³C, δ 169.9, 169.7, and 169.3 (3 COCH₃), 138.0, 137.5, and 128.3–127.5 ($C_6H_5CH_2O$), 101.5 and 100.6 (C-1,1'), 79.5, 75.1, 72.4, 70.0, 68.9, and 67.5 (2 C) (C-2,3,4,5,2',3',4'), 73.3, 70.6, 70.4, and 61.4 (C-6,5' and 2 PhCH₂O), 56.6 (CH₃O), 20.6 (COCH₃). Anal. Calcd for $C_{32}H_{40}O_{13}$: C, 60.75; H, 6.37. Found: C, 60.52; H, 6.42.

Methyl 4-O-acetyl-3,6-di-O-benzyl-2-O-(2,3,4-tri-O-acetyl-β-D-xylopyranosyl)-β-D-mannopyranoside (11).—A solution of 10 (336 mg, 0.52 mmol) in 1:1 pyridine-acetic anhydride (30 mL) was stirred for 16 h at room temperature, and toluene (3 × 15 mL), EtOH (3 × 15 mL), and CH₂Cl₂ (3 × 15 mL) were evaporated from the residue, to give 11, isolated as a yellow solid (350 mg, 99%); R_f 0.60 (95:5 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 7.35–7.16 (m, 10 H, 2 Ph), 5.145 (t, 1 H, $J_{4,3} = J_{4,5} = 9.7$ Hz, H-4), 4.711 and 4.492 (2 d, each 1 H, PhCH₂O), 4.528 (s, 2 H, PhCH₂O), 4.290 (s, 1 H, H-1), 4.152 (d, 1 H, $J_{2,3}$ 3.0 Hz, H-2), 3.480 (s, 3 H, CH₃O), 2.101, 2.071, 2.050, and 1.946 (4 s, each 3 H, 4 Ac); ¹³C, δ 137.6 and 128.2–127.4 ($C_6H_5CH_2O$), 101.4 and 99.0 (C-1,1'), 73.5, 70.7, 70.1, and 59.6 (C-6,5' and 2 PhCH₂O), 56.7 (CH₃O), 20.6 (COCH₃). Anal. Calcd for C₃₄H₄₂O₁₄: C, 60.53; H, 6.27. Found: C, 60.29; H, 6.45.

Methyl 4-O-acetyl-2-O-(2,3,4-tri-O-acetyl-β-D-xylopyranosyl)-β-D-mannopyranoside (12).—To a solution of 11 (375 mg, 0.58 mmol) in 1:1 EtOH–EtOAc (12 mL) were added acetic acid (0.3 mL) and 10% Pd–C (250 mg). Hydrogenolysis was performed at atmospheric pressure for 1 h, the mixture was filtered and concentrated, and toluene (3 × 10 mL) was evaporated from the residue, to yield 12, isolated as a white solid (249 mg, 90%); R_f 0.76 (9:1 CH₂Cl₂–MeOH). NMR data (CDCl₃): ¹H, δ 4.745 (d, 1 H, $J_{1',2'}$ 6.9 Hz, H-1'), 4.380 (s, 1 H, H-1), 4.136 (dd, 1 H, $J_{5'eq,4'}$ 5.1, $J_{5'eq,5'ax}$ – 11.8 Hz, H-5'eq), 4.000 (d, 1 H, $J_{2,3}$ 3.1 Hz, H-2), 3.493 (s, 3 H, CH₃O), 2.846 and 2.810 (2 bs, each 1 H, HO-3,6), 2.104, 2.060, 2.058, and 2.052 (4 s, each 3 H, 4 Ac); ¹³C, δ 170.5 and 169.8–169.6 (COCH₃), 101.1 (C-1,1'), 77.8, 74.2, 71.1, 70.5 (2 C), 69.0, and 68.5 (C-2,3,4,5,2',3',4'), 61.8 and 61.3 (C-6,5'), 56.9 (CH₃O), 20.8, 20.6, and 20.5 (2 C) (4 COCH₃). Anal. Calcd for C₂₀H₃₀O₁₄: C, 48.58; H, 6.12. Found: C, 48.42; H, 6.33.

Benzyl 4,6-O-isopropylidene-3-O-methyl- β -D-glucopyranoside (13).—To a solution of benzyl 3-O-methyl- β -D-glucopyranoside¹⁶ (6.0 g, 21.1 mmol) in 2,2-dimethoxypropane (50 mL) was added *p*-toluenesulfonic acid monohydrate (100 mg), and the mixture was stirred for 45 min. TLC (95:5 CH₂Cl₂-acetone) then showed the reaction to be complete, and solid NaHCO₃ was added. The mixture was diluted with CH₂Cl₂ (300 mL), washed with water, dried (MgSO₄), filtered, and concentrated. The residue was crystallised from EtOH to give 13 (6.0 g, 88%); [α]_D - 69° (*c* 1); mp 108°C; R_f 0.43 (95:5 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 7.37–7.31 (m, 5 H, Ph), 4.911 and 4.632 (2 d, each 1 H, PhCH₂O), 4.448 (d, 1 H,

 $J_{1,2}$ 7.7 Hz, H-1), 3.944 (dd, 1 H, $J_{6a,5}$ 5.4, $J_{6a,6b}$ – 10.5 Hz, H-6a), 3.811 (t, 1 H, $J_{6b,5}$ 10.5 Hz, H-6b), 3.661 (t, 1 H, $J_{4,3} = J_{4,5} = 9.4$ Hz, H-4), 3.598 (s, 3 H, CH₃O), 3.505 (m, 1 H, $J_{2,3}$ 8.9 Hz, H-2), 2.399 (d, 1 H, $J_{HO-2,2}$ 2.2 Hz, HO-2), 1.512 and 1.425 (2 s, each 3 H, CMe₂); ¹³C, δ 137.0 and 128.3–127.8 ($C_6H_5CH_2O$), 102.0 and 99.1 (C-1 and Me₂C), 82.3, 73.9, 73.7, and 67.0 (C-2,3,4,5), 71.0 and 62.0 (C-6 and PhCH₂O), 28.9 and 18.9 [C(CH₃)₂]. Anal. Calcd for C₁₇H₂₄O₆: C, 62.95; H, 7.46. Found: C, 62.61; H, 7.45.

Benzyl 4,6-O-isopropylidene-3-O-methyl-β-D-mannopyranoside (15).—A solution of 13 (5.8 g, 17.9 mmol) in 1:2 acetic anhydride-Me₂SO (100 mL) was stirred for 20 h. TLC (95:5 CH₂Cl₂-acetone) then showed the disappearance of 13 (R_f 0.43), and the formation of ulose 14 (R_f 0.28-0.43) and a faster moving product (R_f 0.66). After partial evaporation of the solvent using toluene (3 × 30 mL), the residue was further concentrated in vacuo (bath < 60°C). Column chromatography (95:5 CH₂Cl₂-acetone) of the residue gave 14, isolated as a syrup (4.6 g, 79%); [α]_D + 20° (c 1) for C-2_{CO}:C-2_{C(OH)2} = 0.2:1. ¹³C NMR data (CDCl₃): δ 187.7 (C-2 as carbonyl) and 93.6 (C-2 as gem-diol).

To a solution of 14 (3.0 g, 9.2 mmol) in 1:1 CH₂Cl₂-MeOH (50 mL) containing Kieselgel 60 (70–230 mesh, 1.0 g) was added NaBH₄ (1.4 g, 37 mmol), as described for 7. After 30 min, TLC (95:5 CH₂Cl₂-acetone) showed the formation of 13 (R_f 0.43) and 15 (R_f 0.17) in the ratio ~ 1:10. The mixture was diluted with CH₂Cl₂ (300 mL), filtered, washed with water, dried (MgSO₄), filtered, and concentrated. Column chromatography [3:2 light petroleum (bp 40–60°C)–EtOAc] of the residue first gave 13 (200 mg, 7%), and then 15 (2.4 g, 79%), both isolated as a white solid. Compound 15: [α]_D -95° (c 1); R_f 0.17. NMR data (CDCl₃): ¹H, δ 7.41–7.29 (m, 5 H, Ph), 4.934 and 4.664 (2 d, each 1 H, PhC H_2 O), 4.523 (s, 1 H, H-1), 4.176 (bd, 1 H, $J_{2,3}$ 3.2 Hz, H-2), 4.084 (t, 1 H, H-4), 3.492 (s, 3 H, CH₃O), 3.278 (dd, 1 H, $J_{3,4}$ 9.3 Hz, H-3), 3.180 (m, 1 H, $J_{5,6a}$ 5.8, $J_{5,6b} = J_{5,4} = 9.8$ Hz, H-5), 2.482 (bs, 1 H, HO-2), 1.527 and 1.431 (2 s, each 3 H, CMe₂); ¹³C, δ 136.3 and 128.2–127.8 (C_6 H₅CH₂O), 99.5 and 98.4 (C-1 and Me₂C), 79.4, 70.1, 68.2, and 67.6 (C-2,3,4,5), 70.4 and 61.8 (C-6 and PhCH₂O), 57.3 (CH₃O), 28.9 and 18.9 [C(CH₃)₂]. Anal. Calcd for C₁₇H₂₄O₆: C, 62.95; H, 7.46. Found: C, 63.33; H, 7.31.

Benzyl 2,4,6-tri-O-acetyl-3-O-methyl-β-D-mannopyranoside (16).—A solution of 15 (2.2 g, 6.8 mmol) in 1:1 acetic acid-water (20 mL) was boiled under reflux for 45 min, then concentrated, and toluene (3 × 20 mL) was evaporated from the residue. The residue was dissolved in 1:1 pyridine-acetic anhydride (20 mL), stirred for 16 h at room temperature, and concentrated, and toluene (3 × 10 mL), EtOH (3 × 10 mL), and CH₂Cl₂ (3 × 10 mL) were evaporated from the residue to yield 16, isolated as a yellow syrup (2.6 g, 93%); R_f 0.43 (95:5 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 7.39–7.32 (m, 5 H, Ph), 5.567 (bd, 1 H, $J_{2,3}$ 3.3 Hz, H-2), 5.132 (t, 1 H, $J_{4,5}$ 9.8 Hz, H-4), 4.893 and 4.665 (2 d, each 1 H, PhCH₂O), 4.518 (d, 1 H, $J_{1,2}$ 0.9 Hz, H-1), 4.293 (dd, 1 H, $J_{6b,5}$ 5.9 Hz, H-6b), 4.183 (dd, 1 H, $J_{6a,5}$ 2.7, $J_{6a,6b}$ – 12.2 Hz, H-6a), 3.541 (m, 1 H, H-5), 3.335 (s, 3 H, CH₃O), 3.321 (dd, 1 H, $J_{3,4}$ 9.8 Hz, H-3), 2.190, 2.114, and 2.072 (3 s, each 3 H, 3 Ac); ¹³C, δ 170.5, 170.2, and 169.5 (3 $COCH_3$), 136.3, 128.2, and 127.8 ($C_6H_5CH_2O$), 96.7 (C-1), 79.3, 72.2, 67.4, and 66.8 (C-2,3,4,5), 70.1 and 62.6 (C-6 and Ph CH_2O), 57.4 (CH₃O), 20.7 and 20.6 (2 C) (3 COCH₃). Anal. Calcd for $C_{20}H_{26}O_9$: C, 58.52; H, 6.39. Found: C, 58.24; H, 6.21.

2,4,6-Tri-O-acetyl-3-O-methyl- α -D-mannopyranosyl trichloroacetimidate (18).— To a solution of 16 (2.5 g, 6.1 mmol) in 1:1 EtOH-EtOAc (30 mL) were added acetic acid (0.5 mL) and 10% Pd-C (250 mg). Hydrogenolysis, and work-up as described for 6, gave 17, isolated as a syrup (1.9 g, 97%); R_f 0.16 (95:5 CH₂Cl₂acetone). Anal. Calcd for C₁₃H₂₀O₉: C, 48.75; H, 6.29. Found: C, 48.12; H, 6.23.

To a solution of 17 (1.9 g, 5.9 mmol) in CH₂Cl₂ (25 mL) and trichloroacetonitrile (7.3 mL, 73 mmol) was added 1,8-diazabicyclo[5.4.0]undec-7-ene (1.0 mL, 6.7 mmol), and the mixture was stirred for 1 h. Then the reaction was complete (TLC 95:5 CH₂Cl₂-acetone, R_f 0.52), and the mixture was concentrated. Column chromatography (95:5 CH₂Cl₂-acetone) of the residue gave **18**, isolated as a yellow syrup (2.0 g, 73%); $[\alpha]_D + 23^\circ$ (c 1). NMR data (CDCl₃): ¹H, δ 8.799 (s, 1 H, NH), 6.303 (d, 1 H, $J_{1,2}$ 2.0 Hz, H-1), 5.536 (dd, 1 H, $J_{2,3}$ 3.4 Hz, H-2), 5.293 (t, 1 H, $J_{4,5}$ 10.0 Hz, H-4), 4.247 (dd, 1 H, $J_{6b,5}$ 5.4, $J_{6b,6a}$ – 12.5 Hz, H-6b), 3.732 (dd, 1 H, $J_{3,4}$ 10.0 Hz, H-3), 3.395 (s, 3 H, CH₃O), 2.204, 2.119, and 2.083 (3 s, each 3 H, 3 Ac); ¹³C, δ 170.0, 169.3, and 169.2 (3 COCH₃), 159.0 (OCNHCCl₃), 94.3 (C-1), 90.0 (OCNHCCl₃), 76.4, 70.8, 66.4, and 65.7 (C-2,3,4,5), 61.8 (C-6), 57.5 (CH₃O), 20.3 (COCH₃). Anal. Calcd for C₁₅H₂₀O₉NCl₃: C, 38.77; H, 4.34. Found: C, 38.40; H, 4.45.

Methyl 4-O-acetyl-3,6-di-O-(2,4,6-tri-O-acetyl-3-O-methyl- α -D-mannopyranosyl)-2-O-(2,3,4-tri-O-acetyl-\beta-b-xylopyranosyl)-\beta-b-mannopyranoside (19).—A solution of 12 (50 mg, 0.1 mmol) and 18 (170 mg, 0.37 mmol) in CH₂Cl₂ (10 mL) containing powdered 4A molecular sieves (1.0 g) was stirred for 30 min under N₂. A solution of trimethylsilyl triflate (40 μ L) in CH₂Cl₂ (2 mL) was added dropwise at -40°C, and after 1 h, when TLC showed the disappearance of 12 and a new compound 19 $(R_f 0.18, 9:1 \text{ CH}_2\text{Cl}_2\text{-acetone})$, pyridine (1 mL) was added, the mixture was filtered through Celite and concentrated, and toluene $(3 \times 10 \text{ mL})$ was evaporated from the residue. Column chromatography (9:1 CH₂Cl₂-acetone) of the residue gave 19, isolated as a white solid (72 mg, 71%); $[\alpha]_D - 34^\circ$ (c 1). NMR data (CDCl₃): ¹H, δ 5.438 and 5.102 (2 dd, each 1 H, $J_{2'',1''} = J_{2'',1'''} = 1.7$, $J_{2'',3''} = J_{2''',3'''}$ = 3.4, H-2",2""), 5.028 (dd, 1 H, $J_{2',3'}$ 9.1 Hz, H-2'), 4.908 and 4.814 (bs and d, each 1 H, H-1",1"'), 4.900 (m, 1 H, H-4'), 4.765 (d, 1 H, J_{1',2'} 7.1 Hz, H-1'), 4.353 (s, 1 H, H-1), 4.275 (dd, 1 H, $J_{5'eq,4'}$ 5.6, $J_{5'eq,5'ax}$ –12.3 Hz, H-5'eq), 3.497 and 3.398 (2 s, 3 and 6 H, 3 CH₃O), 3.306 (dd, 1 H, J_{5'ax,4'} 9.0 Hz, H-5'ax), 2.148, 2.144, 2.128, 2.098, 2.089, 2.077, 2.068, and 2.043 (8 s, 3,3,3,3,6,6,3,3 H, 10 Ac); 13 C, δ 170.5-169.0 (COCH₃), 101.3 (2 C), 99.9, and 97.6 (C-1,1',1",1"), 78.5, 76.4, 76.2, 75.9, 72.6, 70.8, 70.3, 69.4, 68.7, 68.5, 68.3, 68.0, 67.8, 67.5, and 67.4 (C-2,3,4,5,2',3',4',2",3",4",5",2",3",4"',5"'), 66.4, 63.0, 62.5, and 61.6 (C-6,5',6",6"'), 58.0, 57.6, and 56.7 (3 CH₃O), 20.8–20.5 (COCH₃). Anal. Calcd for C₄₆H₆₆O₃₀: C, 50.27; H, 6.05. Found: C, 50.15; H, 6.12.

Methyl 3,6-di-O-(3-O-methyl- α -D-mannopyranosyl)-2-O- β -D-xylopyranosyl- β -D-mannopyranoside (2).—To a solution of **19** (25 mg, 24 μ mol) in MeOH (5 mL) was added NaOMe (pH 10). After 16 h, the solution was neutralised with Dowex-50 (H⁺) resin, filtered, and concentrated, and CH₂Cl₂ (3 × 5 mL) was evaporated from the residue, to give **2**, isolated as a white powder (13 mg, 90%); [α]_D + 0.4° (c 0.5, H₂O); R_f 0.50 (2:1:1 1-butanol–MeOH–water). ¹³C NMR data (D₂O): δ 106.4, 103.3, 102.6, and 100.9 (C-1,1',1",1"'), 58.2, 57.6, and 57.5 (3 CH₃O). For ¹H NMR data, see Table I.

Methyl 3-O-benzyl-4,6-O-prop-2-enylidene-β-D-glucopyranoside (20).—To a solution of methyl 3-O-benzyl-β-D-glucopyranoside¹¹ (11.1 g, 39 mmol) in N,N-dimethylformamide (33 mL) and acrolein dimethyl acetal (15.9 mL, 134 mmol) was added p-toluenesulfonic acid monohydrate (250 mg), and the solution was stirred for 16 h at 60°C. TLC (95:5 CH₂Cl₂-acetone) then showed the reaction to be complete, and triethylamine (2 mL) was added. The mixture was concentrated in vacuo (bath $< 60^{\circ}$ C), diluted with CH₂Cl₂ (250 mL), washed with water, dried (MgSO₄), filtered, and concentrated. Crystallisation of the residue from EtOH gave 20 (9.4 g, 75%); mp 146°C; $[\alpha]_D = -6^\circ$ (c 1); $R_f = 0.51$ (95:5 CH₂Cl₂-acetone). NMR data $(CDCl_{3})$: ¹H, δ 7.40–7.28 (m, 5 H, Ph), 5.879 (m, 1 H, H₂C=CHCH), 5.509 and 5.345 (2 m, each 1 H, H₂C=CHCH), 5.012 (m, 1 H, H₂C=CHCH), 4.948 and 4.771 (2 d, each 1 H, PhCH₂O), 4.287 (d, 1 H, J_{1,2} 7.6 Hz, H-1), 4.248 (dd, 1 H, J_{6a,5} 5.0, $J_{6a,6b}$ -10.5 Hz, H-6a), 3.550 (s, 3 H, CH₃O), 3.351 (m, 1 H, $J_{5,4} = J_{5,6b} = 9.5$ Hz, H-5), 2.386 (d, 1 H, $J_{HO-2,2}$ 2.1 Hz, HO-2); ¹³C, δ 138.2 and 128.3–127.7 (C₆H₅CH₂O), 133.4 (H₂C=CHCH), 118.9 (H₂C=CHCH), 104.0 (H₂C=CHCH), 100.3 (C-1), 80.9, 80.0, 74.0, and 66.1 (C-2,3,4,5), 74.4 and 68.2 (C-6 and PhCH₂O), 57.2 (CH₃O). Anal. Calcd for C₁₇H₂₂O₆: C, 63.34; H, 6.88. Found: C, 63.31; H, 6.76.

Methyl 3-O-benzyl-4,6-O-prop-2-enylidene- β -D-mannopyranoside (22).—A solution of 20 (7.2 g, 22 mmol) in 1:2 acetic anhydride–Me₂SO (10 mL) was stirred for 20 h. TLC (95:5 CH₂Cl₂-acetone) then showed the formation of the ulose 21 (R_f 0.23–0.35) and a faster moving product (R_f 0.87). Work-up as described for 6 gave 21, isolated as a yellow solid (8.3 g). A purified sample gave [α]_D – 50° (c 1) for C-2_{CO}:C-2_{C(OH)2} = 1:1. ¹³C NMR data (CDCl₃): δ 187.9 (C-2 as carbonyl) and 93.4 (C-2 as gem-diol).

To a solution of **21** (7.1 g, 22 mmol) in 1:1 CH₂Cl₂-MeOH (200 mL) containing Kieselgel 60 (70-230 mesh, 3.0 g) was added NaBH₄ (2.6 g, 68 mmol), as described for 7. After 30 min, TLC (95:5 CH₂Cl₂-acetone) showed **20** (R_f 0.51) and **22** (R_f 0.31) in the ratio ~ 1:6. The mixture was diluted with CH₂Cl₂ (300 mL), filtered, washed with water, dried (MgSO₄), filtered, and concentrated. Column chromatography [3:2 light petroleum (bp 40-60°C)-EtOAc] of the residue first gave **20** (800 mg, 11%), and then **22** (4.0 g, 56%), both isolated as a white solid (yields calculated from **20**). Compound **22**: $[\alpha]_D$ -16° (c 1); R_f 0.18. NMR data (CDCl₃): ¹H, δ 7.41-7.31 (m, 5 H, Ph), 5.889 (m, 1 H, H₂C=CHCH), 5.518 and 5.344 (2 m, each 1 H, H_2 C=CHCH), 5.050 (m, 1 H, H₂C=CHCH), 4.851 and 4.746 (2 d, each 1 H, PhC H_2 O), 4.386 (d, 1 H, $J_{1,2}$ 1.2 Hz, H-1), 4.229 (dd, 1 H, $J_{6a,5}$ 4.9, $J_{6a,6b}$ – 10.5 Hz, H-6a), 4.090 (m, 1 H, H-2), 3.973 (t, 1 H, $J_{4,5}$ 9.5 Hz, H-4), 3.739 (t, 1 H, $J_{6b,5}$ 10.4 Hz, H-6b), 3.592 (dd, 1 H, $J_{2,3}$ 3.3, $J_{3,4}$ 9.5 Hz, H-3), 3.540 (s, 3 H, CH₃O), 3.251 (m, 1 H, H-5), 2.488 (d, 1 H, $J_{HO-2,2}$ 1.5 Hz, HO-2); ¹³C, δ 133.6 (H₂C=CHCH), 128.4 and 127.8 ($C_6H_5CH_2O$), 118.9 (H₂C=CHCH), 101.3 and 100.6 (C-1 and H₂C=CHCH), 77.0, 76.6, 69.6, and 66.7 (C-2,3,4,5), 72.4 and 68.1 (C-6 and PhCH₂O), 57.2 (CH₃O). Anal. Calcd for C₁₇H₂₂O₆: C, 63.34; H, 6.88. Found: C, 63.42; H, 6.84.

Methyl 3-O-benzyl-4,6-O-prop-2-enylidene-2-O-(2,3,4-tri-O-acetyl-B-D-xylopyranosyl)-B-D-mannopyranoside (23).--A solution of 22 (2.1 g, 6.5 mmol) and 2,3,4-tri-O-acetyl- α -D-xylopyranosyl trichloroacetimidate¹⁴ (8; 3.6 g, 8.6 mmol) in CH₂Cl₂ (50 mL) containing powdered 4A molecular sieves (5.0 g) was stirred for 30 min under N₂. A solution of trimethylsilyl triflate (300 μ L) in CH₂Cl₂ (10 mL) was added dropwise at -40° C. After 75 min, when TLC showed the disappearance of 22 and a new compound 23 (R_f 0.45, 95:5 CH₂Cl₂-acetone), pyridine (1 mL) was added, the mixture was filtered through Celite and concentrated, and toluene $(3 \times 10 \text{ mL})$ was evaporated from the residue. Column chromatography [1:1 light petroleum (bp 40-60°C)-EtOAc] of the residue afforded 23, isolated as a white foam (3.4 g, 89%); $[\alpha]_{\rm D} = 86^{\circ} (c \ 1)$; $R_f \ 0.37$. NMR data (CDCl₃): ¹H, $\delta \ 7.39 = 7.29$ (m, 5 H, Ph), 5.872 (m, 1 H, H₂C=CHCH), 5.505 and 5.326 (2 m, each 1 H, H_2 C=CHCH), 5.166 (t, 1 H, $J_{3',2'} = J_{3',4'} = 7.7$ Hz, H-3'), 4.913 (d, 1 H, $J_{1',2'}$ 5.6 Hz, H-1'), 4.738 (s, 2 H, PhC H_2 O), 4.302 (dd, 1 H, $J_{5'eq,4'}$ 4.6, $J_{5'eq,5'ax}$ -12.0 Hz, H-5'eq), 4.276 (s, 1 H, H-1), 4.177 (dd, 1 H, $J_{6a,5}$ 4.8, $J_{6a,6b}$ – 10.4 Hz, H-6a), 4.129 (d, 1 H, $J_{2,3}$ 3.0 Hz, H-2), 3.861 (t, 1 H, $J_{4,3} = J_{4,5} = 9.6$ Hz, H-4), 3.657 (t, 1 H, $J_{6b,5}$ 10.3 Hz, H-6b), 3.506 (dd, 1 H, J_{3,4} 9.9 Hz, H-3), 3.441 (s, 3 H, CH₃O), 3.371 (dd, 1 H, J_{5'ax,4'} 7.4 Hz, H-5'ax), 3.181 (m, 1 H, H-5), 2.063, 2.054, and 2.028 (3 s, each 3 H, 3 Ac); ¹³C, δ 169.8 (COCH₃), 138.0 and 128.2–127.5 (C₆H₅CH₂O), 133.6 (H₂C=CHCH), 118.8 (H₂C=CHCH), 102.3 and 100.6 (2 C) (C-1,1' and H₂C=CHCH), 77.7, 76.0, 74.8, 70.2, 69.9, 68.9, and 67.4 (C-2,3,4,5,2',3',4'), 71.4, 68.1, and 61.4 (C-6,5' and PhCH₂O), 56.8 (CH₃O), 20.7 (COCH₃). Anal. Calcd for C₂₈H₃₆O₁₃: C, 57.93; H, 6.25. Found: C, 57.67; H, 6.28.

Methyl 6-O-allyl-3-O-benzyl-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -Dmannopyranoside (24).—A solution of 23 (3.2 g, 5.5 mmol) and sodium cyanoborohydride (4.2 g, 67 mmol) in tetrahydrofuran (100 mL) containing 3A molecular sieves (7.5 g) was stirred for 30 min under N₂. A saturated solution of HCl in diethyl ether was added until the evolution of gas ceased, and the mixture was stirred for 2 h. TLC (95:5 CH₂Cl₂-acetone) then showed the conversion of 23 (R_f 0.45) into 24 (R_f 0.23). The mixture was diluted with CH₂Cl₂ (250 mL), filtered through Celite, washed with water, aq 5% NaHCO₃, and water, dried (MgSO₄), filtered, and concentrated, and MeOH (3 × 25 mL) was evaporated from the residue. Column chromatography (9:1 CH₂Cl₂-acetone) of the residue gave 24, isolated as a white solid (2.2 g, 69%); $[\alpha]_D - 111^\circ$ (c 1); R_f 0.42. NMR data (CDCl₃): ¹H, δ 7.37-7.26 (m, 5 H, Ph), 5.897 (m, 1 H, H₂C=CHCH₂O), 5.280 and 5.174 (2 m, each 1 H, H_2 C=CHCH₂O), 5.181 (t, 1 H, $J_{3',4'}$ 8.0 Hz, H-3'), 5.002 (dd, 1 H, $J_{2',3'}$ 8.2 Hz, H-2'), 4.924 (m, 1 H, $J_{4',5'eq}$ 4.8, $J_{4',5'ax}$ 8.0 Hz, H-4'), 4.899 (d, 1 H, $J_{1',2'}$ 6.3 Hz, H-1'), 4.803 and 4.494 (2 d, each 1 H, PhC H_2 O), 4.260 (s, 1 H, H-1), 4.226 (dd, 1 H, $J_{5'eq,5'ax}$ –12.9 Hz, H-5'eq), 4.148 (d, 1 H, $J_{2,3}$ 2.9 Hz, H-2), 4.049 and 4.031 (2 m, each 1 H, H₂C=CHC H_2 O), 3.776 (dd, 1 H, $J_{6a,5}$ 3.4, $J_{6a,6b}$ –10.5 Hz, H-6a), 3.735 (m, 1 H, H-4), 3.620 (dd, 1 H, $J_{6b,5}$ 6.4 Hz, H-6b), 3.465 (s, 3 H, CH₃O), 3.287 (dd, 1 H, $J_{3,4}$ 9.3 Hz, H-3), 2.538 (d, 1 H, $J_{HO-4,4}$ 1.9 Hz, HO-4), 2.049 (s, 9 H, 3 Ac); ¹³C, δ 169.8 (2 C) and 169.4 (3 COCH₃), 137.5 and 128.4–127.8 (C_6H_5 CH₂O), 134.5 (H_2 C=CHCH₂O), 116.9 (H_2 C=CHCH₂O), 101.5 and 100.6 (C-1,1'), 72.3, 70.6, 70.4, and 61.5 (C-6,5, PhCH₂O, and H_2 C=CHCH₂O), 56.6 (CH₃O), 20.7 (COCH₃). Anal. Calcd for C₂₈H₃₈O₁₃: C, 57.72; H, 6.57. Found: C, 57.15; H, 6.42.

Methyl 4-O-acetyl-6-O-allyl-3-O-benzyl-2-O-(2,3,4-tri-O-acetyl-β-D-xylopyranosyl)-β-D-mannopyranoside (25).—A solution of 24 (2.1 g, 3.6 mmol) in 1:1 pyridine-acetic anhydride (50 mL) was stirred for 16 h, and toluene (3×20 mL), EtOH (3 \times 20 mL), and CH₂Cl₂ (3 \times 20 mL) were evaporated from the residue to give 25, isolated as a yellow solid (2.2 g, 98%); R_f 0.56 (95:5 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 7.35–7.27 (m, 5 H, Ph), 5.868 (m, 1 H, H₂C=CHCH₂O), 5.255 and 5.163 (2 m, each 1 H, H_2 C=CHCH₂O), 5.128 (t, 1 H, $J_{4,3} = J_{4,5} = 9.6$ Hz, H-4), 4.716 and 4.499 (2 d, each 1 H, PhCH₂O), 4.442 (dd, 1 H, $J_{5'ax/eaA'}$ 3.4, $J_{5'ax,5'eq}$ - 12.0 Hz, H-5'ax or H-5'eq), 4.281 (s, 1 H, H-1), 4.149 (d, 1 H, $J_{2,3}$ 3.0 Hz, H-2), 4.000 and 3.982 (2 m, each 1 H, H₂C=CHCH₂O), 3.469 (s, 3 H, CH₃O), 2.141, 2.075, 2.072, and 2.028 (4 s, each 3 H, 4 Ac); ¹³C, δ 169.7 (3 C) and 169.0 (4 COCH₃), 137.6, 128.3, 127.7, and 127.5 (C₆H₅CH₂O), 134.4 (H₂C=CHCH₂O), 117.1 (H₂C=CHCH₂O), 101.4 and 99.0 (C-1,1'), 77.3, 73.9, 72.8, 68.7, 68.2, and 67.8 (2 C) (C-2,3,4,5,2',3',4'), 72.4, 70.7, 70.0, and 59.6 (C-6,5', PhCH₂O, and H₂C=CHCH₂O), 56.8 (CH₃O), 20.7 (COCH₃). Anal. Calcd for C₃₀H₄₀O₁₄: C, 57.68; H, 6.45. Found: C, 57.28; H, 6.53.

Methyl 4-O-acetyl-3-O-benzyl-2-O-(2,3,4-tri-O-acetyl-β-D-xylopyranosyl)-β-Dmannopyranoside (**26-a**) and methyl 6-O-acetyl-3-O-benzyl-2-O-(2,3,4-tri-O-acetylβ-D-xylopyranosyl)-β-D-mannopyranoside (**26-b**).—To a mixture of **25** (746 mg, 1.2 mmol) and 1,4-diazabicyclo[2.2.2]octane (100 mg) in 8:3:1 EtOH-toluene-water (40 mL) was added tris(triphenylphosphine)rhodium(I) chloride (170 mg), and the mixture was boiled under reflux for 2 h, then cooled, and concentrated. A solution of the residue in acetone (45 mL) and M HCl (5 mL) was boiled under reflux for 30 min, when TLC (9:1 CH₂Cl₂-acetone) showed the conversion of the propenyl analogue of **25** (R_f 0.88) into **26-a** (R_f 0.14) to be complete. The mixture was neutralised with aq 5% NaHCO₃, concentrated, diluted with CH₂Cl₂ (100 mL), washed with water, dried (MgSO₄), filtered, and concentrated. Column chromatography (8:2 CH₂Cl₂-acetone) of the residue first gave **26-b** (280 mg, 40%); R_f 0.73, then **26-a** (371 mg, 53%); $[\alpha]_D - 108^{\circ}$ (c 1); R_f 0.57, both isolated as a white solid. NMR data (CDCl₃) for **26-a**: ¹H, δ 7.71–7.31 (m, 5 H, Ph), 5.243 (t, 1 H, J_{4,5} 9.8 Hz, H-4), 4.730 and 4.505 (2 d, each 1 H, PhCH₂O), 4.433 and 3.402 (2 dd, each 1 H, $J_{5'ax/eq,4'}$ 3.5 and 4.7, $J_{5'ax,5'eq}$ – 12.6 and – 12.4 Hz, H-5'eq,5'ax), 4.317 (s, 1 H, H-1), 4.164 (d, 1 H, $J_{2,3}$ 2.9 Hz, H-2), 3.487 (dd, 1 H, $J_{3,4}$ 9.8 Hz, H-3), 3.475 (s, 3 H, CH₃O), 3.312 (m, 1 H, H-5), 2.481 (m, 1 H, HO-6), 2.135, 2.082, 2.069, and 2.051 (4 s, each 3 H, 4 Ac); ¹³C, δ 169.9 and 169.2 (COCH₃), 137.5, 128.3, 127.7, and 127.4 ($C_6H_5CH_2O$), 101.5 and 99.2 (C-1,1'), 77.3, 74.5, 72.8, 68.5, 68.0 (2 C), and 67.5 (C-2,3,4,5,2',3',4'), 70.6, 61.5, and 59.7 (C-6,5' and PhCH₂O), 56.9 (CH₃O), 20.8 (3 C) and 20.6 (4 COCH₃). NMR data (CDCl₃) for **26-b**: ¹H, δ 7.39–7.27 (m, 5 H, Ph), 5.212 (t, 1 H, $J_{3',4'}$ 8.8 Hz, H-3'), 5.026 (dd, 1 H, $J_{2',3'}$ 9.0 Hz, H-2'), 4.957 (m, 1 H, $J_{4',5'ax}$ 8.8, $J_{4',5'eq}$ 5.1 Hz, H-4'), 4.863 (d, 1 H, $J_{1',2'}$ 7.0 Hz, H-1'), 4.809 and 4.479 (2 d, each 1 H, PhCH₂O), 4.256 (s, 1 H, H-1), 4.174 (dd, 1 H, H-5'eq), 4.158 (d, 1 H, $J_{2,3}$ 3.0 Hz, H-2), 3.759 (dt, 1 H, $J_{4,5}$ 9.5 Hz, H-4), 3.459 (s, 3 H, CH₃O), 3.336 (dd, 1 H, $J_{5'ax,5'eq}$ – 11.8 Hz, H-5'ax), 3.283 (dd, 1 H, $J_{3,4}$ 9.2 Hz, H-3), 2.512 (d, 1 H, $J_{HO-4,4}$ 2.6 Hz, HO-4), 2.124 and 2.038 (2 s, 3 and 9 H, 4 Ac). Anal. Calcd for C₂₇H₃₆O₁₄: C, 55.48; H, 6.21. Found **26-a**: C, 55.36; H, 6.26.

Methyl 4-O-acetyl-3-O-benzyl-6-O-(2,3,4,6-tetra-O-acetyl-α-D-mannopyranosyl)-2- $O-(2,3,4-tri-O-acetyl-\beta-D-xylopyranosyl)-\beta-D-mannopyranoside$ (28).—A solution of **26-a** (309 mg, 0.53 mmol) and 2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl trichloroacetimidate⁸ (27; 345 mg, 0.70 mmol) in CH₂Cl₂ (10 mL), containing powdered 4A molecular sieves (1 g), was stirred for 30 min under N₂. A solution of trimethylsilyl triflate (50 μ L) in CH₂Cl₂ (5 mL) was added dropwise at -40°C, and after 1 h, when TLC showed the disappearance of 26-a and a new product 28 (R_f 0.53, 9:1 CH₂Cl₂-acetone), pyridine (1 mL) was added, the mixture was filtered through Celite and concentrated, and toluene $(3 \times 10 \text{ mL})$ was evaporated from the residue. Column chromatography (9:1 CH_2Cl_2 -acetone) of the residue gave 28, isolated as a white solid (359 mg, 77%); $[\alpha]_D = 65^\circ$ (c 1.5). NMR data (CDCl₃): ¹H, δ 7.36–7.25 (m, 5 H, Ph), 5.199 (dd, 1 H, $J_{2'',3''}$ 3.0 Hz, H-2"), 4.790 (d, 1 H, $J_{1'',2''}$ 1.8 Hz, H-1"), 4.718 and 4.506 (2 d, each 1 H, PhCH₂O), 4.401 and 3.398 (2 dd, each 1 H, $J_{5'eq/ax,4'}$ 3.5 and 4.9, $J_{5'eq,5'ax}$ – 12.3 Hz, H-5'ax,5'eq), 4.311 (s, 1 H, H-1), 4.157 (d, 1 H, J_{2.3} 3.0 Hz, H-2), 3.481 (s, 3 H, CH₃O), 2.147, 2.113, 2.104, 2.066, 2.052, 2.014, and 1.978 (7 s, 3,3,3,6,3,3,3 H, 8 Ac); 13 C, δ 170.6–168.8 (COCH₃), 137.5 and 128.3-127.5 (C₆H₅CH₂O), 101.3, 99.2, and 96.9 (C-1,1',1"), 77.5, 72.9, 72.7, 69.3, 68.9, 68.7, 68.5 (2 C), 68.0 (2 C), and 65.9 (C-2,3,4,5,2',3',4',2",3",4",5"), 70.9, 67.6, 62.1, and 59.8 (C-6,5',6" and PhCH₂O), 56.7 (CH₃O), 20.7 (COCH₃). Anal. Calcd for C₄₁H₅₄O₂₃: C, 53.83; H, 5.95. Found: C, 53.29; H, 6.01.

Methyl 4-O-acetyl-6-O-(2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl)-2-O-(2,3,4tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranoside (29).—To a solution of 28 (330 mg, 0.37 mmol) in 1:1 EtOH-EtOAc (12 mL) were added acetic acid (0.3 mL) and 10% Pd-C (250 mg). Hydrogenolysis, and work-up as described for 12, gave 29, isolated as a white solid (300 mg, 98%); R_f 0.11 (9:1 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 5.228 (dd, 1 H, $J_{2'',3''}$ 3.2 Hz, H-2''), 4.944 (dd, 1 H, $J_{2',3''}$ 8.6 Hz, H-2'), 4.836 (d, 1 H, $J_{1'',2''}$ 1.8 Hz, H-1''), 4.750 (d, 1 H, $J_{1',2'}$ 6.7 Hz, H-1'), 4.385 (s, 1 H, H-1), 4.001 (d, 1 H, $J_{2,3}$ 3.2 Hz, H-2), 3.504 (s, 3 H, CH₃O), 3.388 (dd, 1 H, $J_{5'ax,4'}$ 8.8, $J_{5'ax,5'eq}$ – 11.9 Hz, H-5'ax), 2.156, 2.121, 2.113, 2.064, 2.041, 2.010, and 1.986 (7 s, 3,3,3,3,6,3,3 H, 8 Ac); ¹³C, δ 170.5–169.4 (COCH₃), 100.9 (2 C) and 96.8 (C-1,1',1"), 77.9, 72.6, 71.0, 70.3, 70.0 (2 C), 69.3, 68.8, 68.3 (2 C), and 65.8 (C-2,3,4,5,2',3',4',2",3",4",5"), 66.8, 62.0, and 61.7 (C-6,5',6"), 56.8 (CH₃O), 20.5 (COCH₃). Anal. Calcd for C₃₄H₄₈O₂₃: C, 49.52; H, 5.87. Found: C, 49.16; H, 5.81.

Methyl 4-O-acetyl-6-O-(2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl)-3-O-(2,4,6tri-O-acetyl-3-O-methyl-α-D-mannopyranosyl)-2-O-(2,3,4-tri-O-acetyl-β-D-xylopyranosyl)-B-D-mannopyranoside (30).—A solution of 29 (109 mg, 0.13 mmol) and 18 (140 mg, 0.3 mmol) in CH_2Cl_2 (10 mL) containing powdered 4A molecular sieves (1 g) was stirred for 30 min under N₂. A solution of trimethylsilyl triflate (30 μ L) in CH_2Cl_2 (5 mL) was added dropwise at -40°C, and after 1.5 h, when TLC showed the disappearance of 29 and a new compound 30 (R_f 0.49, 9:1 CH₂Cl₂-acetone), pyridine (1 mL) was added. The mixture was filtered through Celite and concentrated, and toluene $(3 \times 10 \text{ mL})$ was evaporated from the residue. Column chromatography (9:1 CH_2Cl_2 -acetone) of the residue afforded 30, isolated as a white solid (90 mg, 61%); $[\alpha]_{\rm D} = 19^{\circ} (c \ 1)$. NMR data (CDCl₃): ¹H, δ 4.926 (d, 1 H, $J_{1'',2''}$ or J_{1'",2'"} 1.7 Hz, H-1" or H-1""), 4.367 (s, 1 H, H-1), 4.308 (dd, 1 H, J_{5'eg,4'} 5.2 Hz, H-5'eq), 4.002 (d, 1 H, J₂₃ 3.0 Hz, H-2), 3.505 and 3.386 (2 s, each 3 H, 2 CH₃O), 3.346 (dd, 1 H, J_{5'ax,4'} 8.2, J_{5'ax,5'eg} -11.6 Hz, H-5'ax), 2.155, 2.128, 2.123, 2.085, 170.6–168.9 (COCH₃), 101.1, 101.0, 99.9, and 96.9 (C-1,1',1",1"), 67.3, 63.0, 62.1, and 61.3 (C-6,5',6",6"), 58.0 and 56.8 (2 CH₃O), 20.5 (COCH₃). Anal. Calcd for C47H66O31: C, 50.09; H, 5.90. Found: C, 49.15; H, 5.82.

Methyl 6-O-α-D-mannopyranosyl-3-O-(3-O-methyl-α-D-mannopyranosyl)-2-O-β-D-xylopyranosyl-β-D-mannopyranoside (3).—Deacetylation of 30 (24 mg, 22 μ mol), as described for 2, gave 3, isolated as a white powder (14 mg, 96%); $[\alpha]_D + 2^\circ$ (*c* 0.5, H₂O); R_f 0.47 (2:1:1 1-butanol-MeOH-water). ¹³C NMR data (D₂O): δ 106.5, 103.3, 102.6, and 100.9 (C-1,1',1",1"), 58.2 and 57.4 (2 CH₃O). For ¹H NMR data, see Table I.

Methyl 4-O-acetyl-3-O-benzyl-6-O-(2,4,6-tri-O-acetyl-3-O-methyl- α -D-mannopyranosyl)-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranoside (31). —Compound 25 (200 mg, 0.32 mmol) was converted into 26a as described above, but the residue after work-up was not purified by column chromatography. A solution of the residue (26-a) and 18 (250 mg, 0.54 mmol) in CH₂Cl₂ (10 mL) containing powdered 4A molecular sieves (1 g) was stirred for 30 min under N₂. A solution of trimethylsilyl triflate (50 μ L) in CH₂Cl₂ (5 mL) was added dropwise at -40°C, and after 1 h, when TLC showed the disappearance of 26-a and a new product 31 (R_f 0.23, 9:1 CH₂Cl₂-acetone), pyridine (1 mL) was added, the mixture was filtered through Celite and concentrated, and toluene (3 × 10 mL) was evaporated from the residue. Column chromatography (9:1 CH₂Cl₂-acetone) of the residue gave 31, isolated as a white solid (155 mg, 51%); [α]_D -63° (c 0.3). NMR data (CDCl₃): ¹H, δ 7.37–7.27 (m, 5 H, Ph), 5.371 (dd, 1 H, $J_{2'',3''}$ 3.4 Hz, H-2"), 4.995 (dd, 1 H, $J_{2',3'}$ 6.5 Hz, H-2'), 4.947 (d, 1 H, $J_{1',2'}$ 4.7 Hz, H-1'), 4.811 (d, 1 H, $J_{1'',2''}$ 1.8 Hz, H-1"), 4.727 and 4.480 (2 d, each 1 H, PhCH₂O), 4.298 (s, 1 H, H-1), 4.260 (dd, 1 H, $J_{5'eq,4'}$ 5.5, $J_{5'eq,5'ax}$ –12.3 Hz, H-5'eq), 4.167 (d, 1 H, $J_{2,3}$ 3.0 Hz, H-2), 3.481 and 3.365 (2 s, each 3 H, 2 CH₃O), 2.134, 2.099, 2.083, 2.078, 2.059, and 2.034 (6 s, 3,3,3,6,3,3 H, 7 Ac); ¹³C, δ 170.3–168.8 (COCH₃), 137.5, 128.3, and 127.8–127.6 ($C_6H_5CH_2O$), 101.6, 99.8, and 97.4 (C-1,1',1''), 67.4, 70.6, 66.7, 62.5, and 60.4 (C-6,5',6'' and PhCH₂O), 57.7 and 56.7 (2 CH₃O), 20.7 (COCH₃).

Methyl 4-O-acetyl-6-O-(2,4,6-tri-O-acetyl-3-O-methyl-α-D-mannopyranosyl)-2-O-(2,3,4-tri-O-acetyl-β-D-xylopyransyl)-β-D-mannopyranoside (32).—To a solution of 31 (145 mg, 0.16 mmol) in 1:1 EtOH–EtOAc (16 mL) were added acetic acid (0.4 mL) and 10% Pd–C (125 mg). Hydrogenolysis, and work-up as described for 12, gave 32, isolated as a white solid (127 mg, 98%); R_f 0.17 (9:1 CH₂Cl₂-acetone). ¹³C NMR data (CDCl₃): δ 170.6, 170.3, 170.0, 169.9, 169.8, 169.6, and 169.0 (7 COCH₃), 101.4, 101.2, and 97.5 (C-1,1',1"), 77.7, 76.4, 72.5, 71.3, 70.8, 70.1, 69.3, 68.6 (2 C), 67.9, and 67.4 (C-2,3,4,5,2',3',4',2",3",4",5"), 66.1, 62.5, and 62.1 (C-6,5',6"), 57.7 and 56.8 (2 CH₃O), 20.9 (2 C), 20.7 (3 C), 20.6, and 20.4 (7 COCH₃).

Methyl 4-O-acetyl-3-O-(2,3,4,6-tetra-O-acetyl-α-D-mannopyranosyl)-6-O-(2,4,6 $tri-O-acetyl-3-O-methyl-\alpha-D-mannopyranosyl)-2-O-(2,3,4-tri-O-acetyl-\beta-D-xylopyran-b-xylo$ osyl)-B-D-mannopyranoside (33).-To a solution of 32 (95 mg, 0.12 mmol) and 2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl trichloroacetimidate⁸ (27; 65 mg, 0.13) mmol) in CH₂Cl₂ (10 mL) containing powdered 4A molecular sieves (1 g) was added dropwise a solution of trimethylsilyl triflate (50 μ L) in CH₂Cl₂ (5 mL) at -40°C, and the mixture was stirred for 1.5 h. TLC (9:1 CH_2Cl_2 -acetone) then indicated the disappearance of 32 and a new product 33 (R_f 0.41), and pyridine (1 mL) was added. The mixture was filtered through Celite and concentrated, and toluene $(3 \times 10 \text{ mL})$ was evaporated from the residue. Column chromatography $(85:15 \text{ CH}_2\text{Cl}_2\text{-acetone})$ of the residue gave 33, isolated as a white solid (92 mg, 70%); $[\alpha]_{\rm D} = -27^{\circ}$ (c 1); R_f 0.31. NMR data (CDCl₃): ¹H, δ 5.421 and 5.042 (2 dd, each 1 H, $J_{2'',3''} = J_{2''',3'''} = 3.4$ Hz, H-2",2"'), 5.401 (dd, 1 H, $J_{3''',4'''}$ 10.0 Hz, H-3"'), 4.949 and 4.816 (2 d, each 1 H, $J_{1'',2''} = J_{1''',2''} = 1.7$ Hz, H-1",1""), 4.921 (m, 1 H, $J_{4',3'} = J_{4',5'ax} = 7.1, J_{4',5'ea}$ 4.4 Hz, H-4'), 4.873 (d, 1 H, $J_{1',2'}$ 5.8 Hz, H-1'), 4.365 (s, 1 H, H-1), 4.168 (d, 1 H, J₂₃ 4.0 Hz, H-2), 3.494 and 3.398 (2 s, each 3 H, 2 CH₃O), H, 11 Ac); ¹³C, δ 170.6–168.7 (COCH₃), 101.4, 100.5, 99.3, and 97.5 (C-1,1',1'',1''), 62.5 (3 C) and 61.0 (C-6,5',6",6"), 57.8 and 56.7 (2 CH₃O), 20.9-20.5 (COCH₃). Anal. Calcd for C₄₇H₆₆O₃₁: C, 50.09; H, 5.90. Found: C, 50.31; H, 6.04.

Methyl 3-O- α -D-mannopyranosyl-6-O-(3-O-methyl- α -D-mannopyranosyl)-2-O- β -D-xylopyranosyl- β -D-mannopyranoside (4).—Deacetylation of 33 (40 mg, 36 μ mol), as described for 2, gave 4, isolated as a white powder (24 mg, 96%); $[\alpha]_D$ +4° (c 0.5, H₂O); R_f 0.46 (2:1:1 1-butanol–MeOH–water). ¹³C NMR data

(D₂O): δ 106.3, 103.3, 102.5, and 100.8 (C-1,1',1",1"), 58.2 and 57.6 (2 CH₃O). For ¹H NMR data, see Table I.

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REFERENCES

- 1 J.A. van Kuik, R.P. Sijbesma, J.P. Kamerling, J.F.G. Vliegenthart, and E.J. Wood, Eur. J. Biochem., 160 (1986) 621-625.
- 2 J.A. van Kuik, R.P. Sijbesma, J.P. Kamerling, J.F.G. Vliegenthart, and E.J. Wood, Eur. J. Biochem., 169 (1987) 399-411.
- 3 G.Z. Lederkremer and A.J. Parodi, J. Biol. Chem., 259 (1984) 12514-12518.
- 4 J.P. Kamerling, Pure Appl. Chem., 63 (1991) 465-472.
- 5 J.B. Bouwstra, J. Kerékgyártó, J.P. Kamerling, and J.F.G. Vliegenthart, *Carbohydr. Res.*, 186 (1989) 39-49.
- 6 B.R. Leeflang, J. Kerékgyártó, J.P. Kamerling, and J.F.G. Vliegenthart, *Carbohydr. Res.*, 208 (1990) 117-126.
- 7 H. Mulder, H. Schachter, M. de Jong-Brink, J.G.M. van der Ven, J.P. Kamerling, and J.F.G. Vliegenthart, *Eur. J. Biochem.*, 210 (1991) 459-465.
- 8 J. Kerékgyártó, J.P. Kamerling, J.B. Bouwstra, J.F.G. Vliegenthart, and A. Lipták, Carbohydr. Res., 186 (1989) 51-62.
- 9 J. Kerékgyártó, J.G.M. van der Ven, J.P. Kamerling, A. Lipták, and J.F.G. Vliegenthart, *Carbohydr. Res.*, 238 (1993) 135-145.
- 10 H. Paulsen, Angew. Chem. Int. Ed. Engl., 21 (1982) 155-173.
- 11 P.A. Finan and C.D. Warren, J. Chem. Soc., (1962) 3089-3092.
- 12 R.F. Butterworth and S. Hanessian, Synthesis, (1971) 70-88.
- 13 P.M. Pojer and S.J. Angyal, Aust. J. Chem., 31 (1978) 1031-1040.
- 14 M. Mori, Y. Ito, and T. Ogawa, Carbohydr. Res., 191 (1990) 199-224.
- 15 M. Ek, P.J. Garegg, H. Hultberg, and S. Oscarson, J. Carbohydr. Chem., 2 (1983) 305-311.
- 16 A.K. Sen, K.K. Sarkar, and N. Banerji, J. Carbohydr. Chem., 7 (1988) 645-654.
- 17 S. Sato, Y. Ito, T. Nukada, Y. Nakahara, and T. Ogawa, Carbohydr. Res., 167 (1987) 197-210.
- 18 P.J. Garegg, H. Hultberg, and S. Oscarson, J. Chem. Soc., Perkin Trans. 1, (1982) 2395-2397.
- 19 E.J. Corey and J.W. Suggs, J. Org. Chem., 38 (1973) 3224.
- 20 O. Kanie, T. Takeda, N. Hada, and Y. Ogihara, J. Carbohydr. Chem., 10 (1991) 561-581.
- 21 C.V. Holland, D. Horton, and J.S. Jewell, J. Org. Chem., 32 (1967) 1818-1821.
- 22 H. Paulsen, F. Garrido Espinose, W.P. Trautwein, and K. Heyns, Chem. Ber., 101 (1968) 179-185.
- 23 L.D. Hall and J.F. Manville, Carbohydr. Res., 4 (1967) 512-515.
- 24 A. de Bruyn, M. Anteunis, R. van Rijsbergen, M. Claeyssens, and P. Kovác, J. Carbohydr. Chem., 1 (1982/1983) 301-309.
- 25 P.L. Durette, D. Horton, and N.S. Bhacca, Carbohydr. Res., 10 (1969) 565-577.
- 26 P.L. Durette and D. Horton, Carbohydr. Res., 18 (1971) 57-80.
- 27 E.E. Lee, G. Keaveney, and P.S. O'Colla, Carbohydr. Res., 59 (1977) 268-273.