Synthesis of a selectively protected trisaccharide building block that is part of xylose-containing carbohydrate chains from N-glycoproteins

János Kerékgyártó ^a, Jos G.M. van der Ven ^b, Johannis P. Kamerling ^b, András Lipták ^a and Johannes F.G. Vliegenthart ^b

^a Institute of Biochemistry, L. Kossuth University, P.O. Box 55, H-4010 Debrecen (Hungary) ^b Bijvoet Center, Department of Bio-Organic Chemistry, Utrecht University, P.O. Box 80.075, NL-3508 TB Utrecht (Netherlands)

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ABSTRACT

The synthesis is reported of ethyl 4-O-[3-O-allyl-4,6-O-isopropylidene-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranosyl]-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (16), a key intermediate in the synthesis of xylose-containing carbohydrate chains from N-glycoproteins. Condensation of ethyl 3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (5) with 2,4,6-tri-O-acetyl-3-O-allyl- α -D-glucopyranosyl bromide, using silver triflate as a promoter, gave the β -linked disaccharide derivative 8 (84%). O-Deacetylation of 8 and then isopropylidenation afforded 10, which was converted via oxidation-reduction into ethyl 4-O-(3-O-allyl-4,6-O-isopropylidene- β -D-mannopyranosyl)-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (12). Silver triflate-promoted condensation of 12 with 2,3,4-tri-O-acetyl- α -D-xylopyranosyl bromide gave 16 (71%). The Xyl p unit in 16 and in de-isopropylidenated 16 (17) existed in the ${}^{1}C_{4}(D)$ conformation, but that in O-deacetylated 17 (18) existed in the ${}^{4}C_{1}(D)$ conformation.

INTRODUCTION

Xylose-containing N-linked carbohydrate chains occur in glycoproteins of plant and animal origin. Invariably β -D-Xyl p is $(1 \rightarrow 2)$ -linked to β -D-Man p of the core structure. If α -L-Fuc p is attached to the asparagine-linked D-GlcpNAc, it is $(1 \rightarrow 3)$ -linked in plants and $(1 \rightarrow 6)$ -linked in animals¹, as in 1 for the proteolytic enzyme bromelain from pineapple stem² and in 2 for the α -hemocyanin of *Helix* pomatia³.

For the study of the conformation^{4,5} and biosynthesis⁶ of these glycans, the oligosaccharide glycosides β -D-Xyl p-(1 \rightarrow 2)- β -D-Man p-OMe, β -D-Xyl p-(1 \rightarrow 2)- β -D-Man p-OMe, β -D-Xyl p-(1 \rightarrow 2)- β -D-Xyl p

Correspondence to: Professor Dr. J.P. Kamerling, Bijvoet Center, Department of Bio-Organic Chemistry, Utrecht University, P.O. Box 80.075, NL-3508 TB Utrecht, Netherlands.

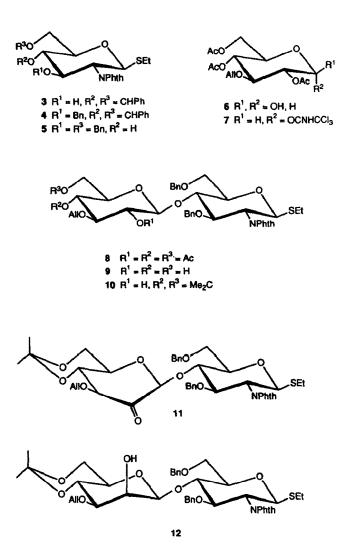
 $[\alpha$ -D-Man p-(1 \rightarrow 6)]- β -D-Man p-OMe, β -D-Xyl p-(1 \rightarrow 2)- $[\alpha$ -D-Man p-(1 \rightarrow 3)]- β -D-Man p-OMe, and β -D-Xyl p-(1 \rightarrow 2)- $[\alpha$ -D-Man p-(1 \rightarrow 3)][α -D-Man p-(1 \rightarrow 6)]- β -D-Man p-OMe have been synthesised⁷. We now describe the synthesis of the glycosyl donor ethyl 4-O-[3-O-allyl-4,6-O-isopropylidene-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranosyl]-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (16), an essential building block for the preparation of 1 and 2.

RESULTS AND DISCUSSION

The crucial step in the synthesis of 16 is the creation of the β -D-Man p bond. This type of linkage can be synthesised "directly" by using a C-1-activated D-Man p donor, having a non-participating group at C-2 with a reactive acceptor⁸⁻¹⁰, and "indirectly" by epimerisation of a β -D-Glc p unit either by an inter- or intra-molecular S_N2 reaction or via oxidation-reduction^{7,11-14}. Of these methods, the oxidation-reduction procedure was chosen because, after reduction, the β -D-Man p residue generated has a blocking pattern ideal for the introduction of β -D-Xyl p and α -D-Man p units.

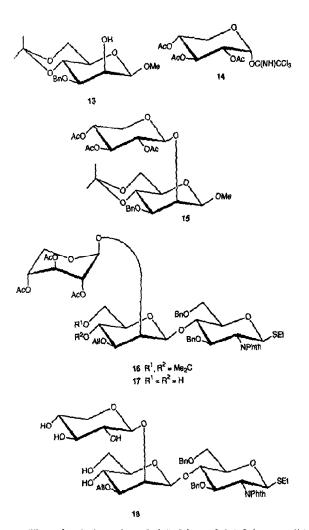
Ethyl 4,6-O-benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside¹⁵ (3) was treated with benzyl bromide in the presence of sodium hydride in boiling tetrahydrofuran to give 4 (90%). Regioselective reductive opening of the 4,6-O-benzylidene ring in 4 with the borane-trimethylamine complex and aluminium(III) chloride¹⁶ in tetrahydrofuran yielded the acceptor 5 (84%). Since HO-4 in D-GlcN derivatives has a low reactivity¹⁷, especially in compounds with the ⁴C₁ conformation, forcing conditions are needed to promote the reaction.

When 2,4,6-tri-O-acetyl-3-O-allyl- α -D-glucopyranosyl trichloroacetimidate (7), obtained from 1,2,4,6-tetra-O-acetyl-3-O-allyl- β -D-glucopyranoside¹⁸ via treatment with hydrazine acetate¹⁹ (\rightarrow 6, 81%) and trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene²⁰(\rightarrow 7, 74%), was used as the donor²¹, condensation with 5 in dichloromethane using trimethylsilyl triflate as a promoter at -40° C gave only 19% of the desired disaccharide derivative 8, and the many UV-absorbing products (TLC) indicated degradation of 5. However, coupling of 5 and 2,4,6-tri-O-acetyl-3-O-allyl- α -D-glucopyranosyl bromide¹⁸ in dichloromethane –toluene in the presence of silver triflate as a promoter afforded 84% of 8.



Zemplén O-deacetylation of 8 (\rightarrow 9, 99%), isopropylidenation with 2,2-dimethoxypropane²² (\rightarrow 10, 82%), and oxidation of HO-2 using methylsulfoxide-acetic anhydride afforded 11 (88%), without affecting the thioethyl and allyl groups.

Although borohydride reduction of β -D-arabino-hexopyranosyl-2-ulose to β -D-Man p derivatives can be achieved in 1:1 dichloromethane-methanol^{12,13,23,24} or 12:1 dioxane-water²⁵, phthalimido groups may be removed²⁶. However, when the borohydride reduction of 11 was carried out in dichloromethane, no degradation was observed, and the manno (12) and gluco (10) isomers were formed in the ratio ~ 7:3. A similar low stereoselectivity without decomposition occurred in 1:2 dichloromethane-2-propanol [\rightarrow 12 (53%) and 10 (38%)]. The isomers 10 and 12 were separated easily by column chromatography, so that 10 could be recycled.



The xylosylation of methyl 3-O-benzyl-4,6-O-isopropylidene- β -D-mannopyranoside (13) with 2,3,4-tri-O-acetyl- α -D-xylopyranosyl bromide in toluene-nitromethane in the presence of Hg(CN)₂ has been reported⁷ but, although the coupling proceeded with a good yield, an α , β -ratio of 1:9 was obtained. However, when 2,3,4-tri-O-acetyl- α -D-xylopyranosyl trichloroacetimidate²⁷ (14) was reacted with 13 in dichloromethane in the presence of trimethylsilyl triflate as a promoter, only the desired β -(1 \rightarrow 2)-linked disaccharide derivative⁷ 15 was formed. Application of the latter method to 12 gave a complex mixture of products, but the coupling of 12 and 2,3,4-tri-O-acetyl- α -D-xylopytanosyl bromide²⁸, catalysed by

silver triflate at -40° C in toluene-dichloromethane, yielded only the desired β -linked trisaccharide derivative 16 (70%).

In the ¹³C NMR spectrum of 16, the resonance of C-1 of the β -D-Xylp unit appeared at relatively high field (δ 98.66), whereas the ¹H NMR spectrum revealed relatively small coupling constants for the Xylp ring ($J_{1,2}$ 3.0, $J_{2,3}$ 5.0, $J_{3,4}$ 4.9 Hz). Based on the findings for ring protons in gauche-gauche arrangements and the chemical shift for the H-5 resonance²⁹, it is concluded that the Xylp unit in 16 has the ¹C₄ conformation. This behaviour is not uncommon for protected β -Xylp derivatives³⁰⁻³² and it has been suggested^{30,31} that the anomeric effect counters the destabilising effect of the three axially oriented O-acetyl groups. Removal of the isopropylidene group from 16 (\rightarrow 17, 97%) did not change the conformation of the Xylp unit (¹³C, C-1 δ 98.92; ¹H, $J_{1,2}$ 3.5, $J_{2,3}$ 5.5, $J_{3,4}$ 5.5 Hz). However, after saponification of 17 (\rightarrow 18, 95%), the NMR parameters completely changed (¹³C, C-1 δ 103.32; ¹H, $J_{1,2}$ 6.7 Hz). The usual ⁴C₁(D) chair conformation was observed for 15.

EXPERIMENTAL

General methods.—The ¹H (200 and 360 MHz) and ¹³C (APT, 50 MHz) NMR spectra were recorded at 25°C with a Bruker WP 200 SY, WP 200, or HX 360 spectrometer. Chemical shifts (δ) are given in ppm relative to the signal for internal Me₄Si (CDCl₃) for ¹H and for ¹³C (indirectly to CDCl₃, δ 76.9). Column chromatography was performed on Kieselgel 60 (Merck, <230 mesh) and fractions were monitored by TLC on Kieselgel 60 F₂₅₄ (Merck) by detection with UV light and then charring with H₂SO₄. Optical rotations were measured for solutions in CHCl₃ at 20°C with a Perkin–Elmer 241 polarimeter, using a 10-cm 1-mL cell. Melting points (uncorrected) were determined on a Kofler apparatus. Solvents were evaporated under reduced pressure at 40°C (bath). All solvents were distilled from the appropriate drying agents.

Ethyl 3-O-benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (4).—A solution of 3^{15} (6.5 g, 14.7 mmol) and benzyl bromide (2.7 mL, 22.7 mmol) in tetrahydrofuran (75 mL) was added dropwise to NaH (700 mg, 29.17 mmol), and the mixture was boiled under reflux. After 1 h, TLC (95:5 CH₂Cl₂acetone) indicated the reaction to be complete. The mixture was cooled, diluted with EtOAc (300 mL), filtered through Celite, and washed with water (3 × 30 mL), and the combined washings were extracted with EtOAc (50 mL). The combined organic phases were washed with water (50 mL), dried (MgSO₄), filtered, and concentrated. Column chromatography (95:5 CH₂Cl₂-acetone) of the residue gave 4, isolated as a syrup (7.1 g, 90%); $[\alpha]_D$ +97° (c 1); R_f 0.60. NMR data (CDCl₃): ¹H, δ 7.87-6.84 (m, 14 H, 2 Ph and Phth), 5.635 (s, 1 H, PhCH), 5.348 (d, 1 H, J_{1,2} 10.6 Hz, H-1), 4.799 and 4.516 (2 d, each 1 H, PhCH₂O), 2.657 (m, 2 H, CH₃CH₂S), 1.169 (t, 3 H, CH₃CH₂S); ¹³C, δ 137.6, 137.1, 133.7, 131.4, 128.8-127.2, 125.9, and 123.1 (2 C) (C₆H₅CH₂O, C₆H₅CH, and Phth), 101.1 (PhCH), 82.8, 81.6, 75.2, and 70.2 (C-1,3,4,5), 74.0 and 68.5 (C-6 and PhCH₂O), 54.5 (C-2), 23.8 (CH₃CH₂S), 14.7 (CH₃CH₂S). Anal. Calcd for $C_{30}H_{29}NO_6S$: C, 67.78; H, 5.50. Found: C, 67.76; H, 5.37.

Ethyl 3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio-β-p-glycopyranoside (5).—A mixture of the borane-trimethylamine complex (5.7 g, 78.1 mmol), powdered 4A molecular sieves (5 g), 4 (2.6 g, 4.9 mmol), and tetrahydrofuran (50 mL) was stirred for 1 h. Aluminium(III) chloride (10.5 g, 78.7 mmol) was added and the mixture was stirred for 5 h in the dark. TLC (9:1 CH₂Cl₂-EtOAc) then showed the conversion of 4 (R_f 0.80) into 5 (R_f 0.46). The mixture was diluted with CH₂Cl₂ (250 mL), filtered through Celite, washed with cold 0.5 M H₂SO₄, water, aq 5% NaHCO₃, and water, dried ($MgSO_4$), filtered, and concentrated. Column chromatography (9:1 CH₂Cl₂-EtOAc) of the residue gave 5 (2.2 g, 84%), which crystallised on storage and had mp 110°C (from EtOH); $[\alpha]_D$ +43° (c 1.5); R_f 0.46. NMR data (CDCl₃): ¹H, δ 7.83-6.94 (m, 14 H, 2 Ph and Phth), 5.275 (d, 1 H, $J_{1,2}$ 10.0 Hz, H-1), 4.750 and 4.543 (2 d, each 1 H, PhC H_2 O), 4.639 and 4.583 (2 d, cach 1 H, PhC H_2 O), 2.949 (d, 1 H, $J_{HO,4}$ 2.5 Hz, HO-4), 2.623 (m, 2 H, CH_3CH_2S), 1.160 (t, 3 H, CH_3CH_2S); ¹³C, δ 168.0 and 167.4 (CO Phth), 138.0, 137.4, 133.7, 131.5, 128.4–127.3, 123.4, and 123.2 (2 C₆H₅CH₂O and Phth), 81.0, 79.4, 77.4, and 74.5 (C-1,3,4,5), 74.3, 73.7, and 70.8 (C-6 and 2 PhCH₂O), 54.3 (C-2), 23.9 (CH₃CH₂S), 14.8 (CH₃CH₂S). Anal. Calcd for C₃₀H₃₁NO₆S: C, 67.52; H, 5.86. Found: C, 68.04; H, 5.78.

2,4,6-Tri-O-acetyl-3-O-allyl- α -D-glucopyranosyl trichloroacetimidate (7).—A solution of 1,2,4,6-tetra-O-acetyl-3-O-allyl- β -D-glucopyranoside¹⁸ (3.88 g, 9.99 mmol) in N,N-dimethylformamide (10 mL) was stirred with hydrazine acetate (1.1 g, 11.9 mmol) at 50°C for 15 min, cooled, diluted with EtOAc (250 mL), and washed with aq 5% NaCl (3 × 30 mL), and the combined washings were extracted with EtOAc (30 mL). The combined organic phases were washed with water (30 mL), dried (MgSO₄), filtered, and concentrated. Column chromatography (9:1 CH₂Cl₂-acetone) of the residue gave 6, isolated as a syrup (2.8 g, 81%); $[\alpha]_D + 47^\circ$ (c 0.5); R_f 0.40 (9:1 CH₂Cl₂-acetone).

To a solution of 6 (1.04 g, 3.00 mmol) in dry CH₂Cl₂ (10 mL) and trichloroacetonitrile (3.0 mL, 29.9 mmol) was added 1,8-diazabicyclo[5.4.0]undec-7-ene (0.56 mL, 3.74 mmol). The mixture was stirred for 20 min, when the reaction was complete (TLC), then concentrated. Column chromatography [3:2 light petroleum (bp 40-60°C)-EtOAc] of the residue gave 7 (1.09 g, 74%), which crystallised on storage and had mp 54°C (from ether); $[\alpha]_D + 64^\circ$ (c 0.5); R_f 0.58. NMR data (CDCl₃): ¹H, δ 8.63 (s, 1 H, NH), 6.48 (d, 1 H, $J_{1,2}$ 3.6 Hz, H-1), 5.78 (m, 1 H, H₂C=CHCH₂O), 5.24-5.06 (m, 3 H, H_2 C=CHCH₂O and H-4), 5.00 (dd, 1 H, $J_{2,3}$ 10.0 Hz, H-2), 2.06, 2.03, and 2.02 (3 s, each 3 H, 3 Ac). Anal. Calcd for C₁₂H₂₂Cl₃NO₉: C, 41.61; H, 4.52. Found: C, 42.03; H, 4.43.

Ethyl 3,6-di-O-benzyl-2-deoxy-2-phthalimido-4-O- $(2,4,6-tri-O-acetyl-3-O-allyl-\beta-D-glucopyranosyl)-1-thio-\beta-D-glucopyranoside (8).—A solution of 5 (1.15 g, 2.16 mmol) and 2,4,6-tri-O-acetyl-3-O-allyl-<math>\alpha$ -D-glucopyranosyl bromide¹⁸ (2.6 g, 6.4

mmol) in CH₂Cl₂ (20 mL) and toluene (20 mL) containing powdered 4A molecular sieves (7.5 g) was stirred for 30 min under Ar. A solution of silver triflate (2.2 g, 8.6 mmol) in toluene (40 mL) was added dropwise in the dark during 2 h at -45° C and stirring was continued for 2 h at -40°C. TLC [3:2 light petroleum (bp 40-60°C)-EtOAc] then showed the absence of 5 (R_f 0.54) and the formation of 8 $(R_f 0.47)$. Pyridine (5 mL) was added, and the mixture was diluted with CH₂Cl₂ (400 mL), filtered through Celite, washed with aq 10% sodium thiosulfate (3×50 mL) and water (3×50 mL), dried (MgSO₄), filtered, and concentrated. Column chromatography [3:2 light petroleum (bp $40-60^{\circ}$ C)-EtOAc] of the residue gave 8, isolated as a syrup (1.56 g, 84%); $[\alpha]_{D}$ + 24° (c 0.5); R_{f} 0.47. NMR data (CDCl₃): ¹H, δ 7.80–6.75 (m, 14 H, 2 Ph and Phth), 5.767 (m, 1 H, H₂C=CHCH₂O), 5.207 and 5.136 (2 m, 2 H, H₂C=CHCH₂O), 5.197 (d, 1 H, J₁₂ 9.8 Hz, H-1), 4.804 and 4.526 (2 d, each 1 H, PhCH₂O), 4.786 and 4.410 (2 d, each 1 H, PhCH₂O), 4.587 (d, 1 H, J_{1',2'} 8.1 Hz, H-1'), 2.617 (m, 2 H, CH₃CH₂S), 2.067, 2.056, and 1.963 (3 s, each 3 H, 3 Ac), 1.162 (t, 3 H, CH_3CH_2S); ¹³C, δ 170.5–166.8 (COCH₃ and CO Phth), 138.3, 137.8, 134.0, 133.5, 131.4, 128.3–127.6, 126.8, and 123.1 (C₆H₅CH₂O, Phth, and H₂C=CHCH₂O), 116.5 (H₂C=CHCH₂O), 100.1 (C-1'), 80.9, 79.8, 78.8, 78.0, 77.4, 72.6, 71.5, and 69.3 (C-1,3,4,5,2',3',4',5'), 74.5, 73.4, 72.5, 67.6, and 61.7 (C-6,6', 2 PhCH₂O, and H₂C=CHCH₂O), 54.5 (C-2), 23.7 (CH₃CH₂S), 23.7, 20.7, and 20.4 (3 COCH₃), 14.7 (CH₃CH₂S). Anal. Calcd for C₄₅H₅₁NO₁₄S: C, 62.71; H, 5.96. Found: C, 62.33, H, 6.07.

Ethyl 4-O-(3-O-allyl-4,6-O-isopropylidene- β -D-glucopyranosyl)-3,6-di-O-benzyl-2deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (10).—A solution of 8 (2.0 g, 2.3 mmol) and NaOMe (250 mg, 4.63 mmol) in MeOH (50 mL) was stirred overnight. TLC (9:1 CH₂Cl₂-MeOH) then showed a complete O-deacetylation (\rightarrow 9, R_f 0.58). The solution was neutralised with Amberlite IR-120 (H⁺) resin, filtered, and concentrated, and CH₂Cl₂ (2 × 10 mL) was evaporated from the residue to afford amorphous 9 (1.7 g, 99%).

To a solution of **9** (1.50 g, 2.04 mmol) in 2,2-dimethoxypropane (15 mL, 122 mmol) was added *p*-toluenesulfonic acid monohydrate (150 mg, 0.79 mmol). After 30 min, TLC (95:5 CH₂Cl₂-acetone) indicated the reaction to be complete, and solid NaHCO₃ was added. The mixture was diluted with CH₂Cl₂ (250 mL), washed with water, dried (MgSO₄), filtered, and concentrated. Column chromatography (95:5 CH₂Cl₂-acetone) of the residue gave **10**, isolated as a white glass (1.30 g, 82%); $[\alpha]_D$ + 50° (*c* 0.4); R_f 0.42. NMR data (CDCl₃): ¹H, δ 7.80–6.84 (m, 14 H, 2 Ph and Phth), 5.935 (m, 1 H, H₂C=CHCH₂O), 5.296 and 5.183 (2 m, 2 H, H_2 C=CHCH₂O), 5.199 (d, 1 H, $J_{1,2}$ 10.4 Hz, H-1), 4.765 and 4.402 (2 d, each 1 H, PhCH₂O), 4.724 and 4.592 (2 d, each 1 H, PhCH₂O), 4.604 (d, 1 H, $J_{1',2'}$ 7.6 Hz, H-1'), 2.631 (m, 2 H, CH₃CH₂S), 1.416 and 1.369 (2 s, each 3 H, CMe₂), 1.169 (t, 3 H, CH₃CH₂S); ¹³C, δ 138.2, 137.7, 135.0, 133.6, 131.4, 128.2–126.9, and 123.1–123.0 (C_6H_5 CH₂O, Phth, and H₂C=CHCH₂O), 116.7 (H₂C=CHCH₂O), 103.1 (C-1'), 98.9 (Me₂C), 80.8, 80.5, 78.9, 78.6, 78.4, 74.6, 73.8, and 66.9 (C-1,3,4,5,2',3',4',5'), 74.5, 73.2, 73.0, 68.1, and 61.9 (C-6,6', 2 PhCH₂O, and H₂C=CHCH₂O), 54.5

(C-2), 28.9 and 18.9 [2 (CH_3)₂C], 23.4 (CH_3CH_2S), 14.7 (CH_3CH_2S). Anal. Calcd for $C_{42}H_{49}NO_{11}S$: C, 65.02; H, 6.37. Found: C, 64.61; H, 6.58.

Ethyl 4-O-(3-O-allyl-4,6-O-isopropylidene-β-D-arabino-hexopyranosyl-2-ulose)-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (11).—A solution of 10 (1.10 g, 1.42 mmol) in 1:2 acetic anhydride–Me₂SO (45 mL) was stirred for 16 h at room temperature, then concentrated, and toluene (3 × 50 mL) was evaporated from the residue. Column chromatography (95:5 CH₂Cl₂-acetone) then gave 11, isolated as a white glass (955 mg, 87%); $[\alpha]_D$ +52° (c 1); R_f 0.42-0.62 (tailing, 3:1 toluene–acetone). NMR data (CDCl₃): ¹H, δ 7.80-6.80 (m, 14 H, 2 Ph and Phth), 5.920 (m, 1 H, H₂C=CHCH₂O), 5.230 (d, 1 H, J_{1,2} 10.5 Hz, H-1), 4.940 (s, 1 H, H-1'), 2.637 (m, 2 H, CH₃CH₂S), 1.545 and 1.415 (2 s, each 3 H, CMe₂), 1.170 (t, 3 H, CH₃CH₂S); ¹³C, δ 187.3 (C-2' as carbonyl), 102.6 and 101.7 (C-1' as carbonyl and gem-diol), 93.9 (C-2' as gem-diol), 99.2 and 99.0 (Me₂C as carbonyl and gem-diol), 54.6 (C-2), 23.6 (CH₃CH₂S), 28.7 and 18.7 [2 (CH₃)₂C], 14.7 (CH₃CH₂S). Anal. Calcd for C₄₂H₄₇NO₁₁S · H₂O: C, 63.70; H, 6.24. Found: C, 63.95; H, 6.18.

Ethyl 4-O-(3-O-allyl-4,6-O-isopropylidene-β-D-mannopyranosyl)-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (12).—To a solution of 11 (1.03 g, 1.33 mmol) in CH₂Cl₂ (5 mL) was added a solution of NaBH₄ (200 mg, 5.29 mmol) in 2-propanol (10 mL) at 0°C. After 10 min, TLC (3:1 toluene-acetone) showed the absence of 9, and the presence of the gluco- (10, R_f 0.62) and manno-(12, R_f 0.42) derivatives in the ratio ~ 3:7. The mixture was diluted with CH₂Cl₂ (200 mL), washed with water, dried (MgSO₄), filtered, and concentrated. Column chromatography (3:1 toluene-acetone) of the residue gave 12, isolated as a white glass (547 mg, 53%); $[\alpha]_{D}$ +41° (c 0.2); and 10 (392 mg, 38%), partially contaminated with 12, which was recycled. NMR data (CDCl₃) for 12: ¹H, δ 7.80–6.85 (m, 14 H, 2 Ph and Phth), 5.892 (m, 1 H, H₂C=CHCH₂O), 5.231 (d, 1 H, J₁₂ 10.5 Hz, H-1), 5.280 and 5.184 (2 m, 2 H, H₂C=CHCH₂O), 4.824 and 4.449 (2 d, each 1 H, PhCH₂O), 4.737 and 4.527 (2 d, each 1 H, PhCH₂O), 4.661 (s, 1 H, H-1'), 2.975 (m, 1 H, H-5'), 2.642 (m, 2 H, CH₃CH₂S), 1.432 and 1.379 (2 s, each 3 H, CMe₂), 1.175 (t, 3 H, CH_3CH_2S); ¹³C, δ 138.2, 137.6, 134.6, 133.6, 131.4, 128.3–126.9, and 123.3 (C₆H₅CH₂O), Phth, and H₂C=CHCH₂O), 116.9 (H₂C=CHCH₂O), 100.6 (C-1'), 99.3 (Me₂C), 80.9, 78.6, 78.4 (2 C), 77.1, 70.4, 69.3, 67.6 (C-1,3,4,5,2',3',4',5'), 74.5, 73.4, 71.0, 68.4, and 61.8 (C-6,6', 2 PhCH₂O, and H₂C=CHCH₂O), 54.5 (C-2), 29.0 and 19.0 [(CH₃)₂C], 23.5 (CH₃CH₂S), 14.7 (CH₃CH₂S). Anal. Calcd for C₄₂H₄₉NO₁₁S: C, 65.02; H, 6.37. Found: C, 64.50; H, 6.60.

Methyl 3-O-benzyl-4,6-O-isopropylidene-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranoside⁷ (15).—A solution of methyl 3-O-benzyl-4,6-O-isopropylidene- β -D-mannopyranoside⁷ (13; 80 mg, 0.25 mmol) and 2,3,4-tri-O-acetyl- α -D-xylopyranosyl trichloroacetimidate²⁷ (14; 126 mg, 0.30 mmol) in CH₂Cl₂ (4 mL) containing 4A molecular sieves (1 g) was stirred for 30 min under Ar. A solution of trimethylsilyl triflate (55 μ L, 0.30 mmol) in CH₂Cl₂ (2 mL) was added dropwise at -40°C. TLC (85:10:5 CH₂Cl₂-EtOAc-acetone) showed that **15** (R_f 0.56) was formed within 10 min. Pyridine (1 mL) was added, the mixture was filtered through Celite and concentrated, and toluene $(3 \times 5 \text{ mL})$ was evaporated from the residue. Column chromatography (85:10:5 CH₂Cl₂-EtOAc-acetone) then gave 15, isolated as a syrup (102 mg, 70%), $[\alpha]_D - 90^\circ$ (c 1); R_f 0.56. For the ¹H NMR data, see ref. 7.

Ethyl 4-O-[3-O-allyl-4,6-O-isopropylidene-2-O-(2,3,4-tri-O-acetyl-B-D-xylopyranosyl)-B-p-mannopyranosyl]-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio-B-p-glucopyranoside (16).--A solution of 12 (300 mg, 0.39 mmol) and 2,3,4-tri-O-acetyl- α -Dxylopyranosyl bromide²⁸ (264 mg, 0.78 mmol) in CH₂Cl₂ (3 mL) and toluene (3 mL) containing powdered 4A molecular sieves (900 mg) was stirred for 30 min under Ar. A solution of silver triflate (400 mg, 1.56 mmol) in toluene (4 mL) was added dropwise in the dark at -45° C during 40 min, and stirring was continued for 20 min at -45°C. TLC [1:1 light petroleum (bp 40-60°C)-EtOAc] then showed the absence of 12 (R_f 0.29) and the formation of 16 (R_f 0.48). Pyridine was added, and the mixture was diluted with CH₂Cl₂ (100 mL), filtered through Celite, washed with aq 10% sodium thiosulfate $(3 \times 10 \text{ mL})$ and water $(3 \times 10 \text{ mL})$, dried (MgSO₄), filtered, and concentrated. Column chromatography [1:1 light petroleum (bp $40-60^{\circ}$ C)-EtOAc] of the residue gave 16, isolated as a white glass (283 mg, 70%); $[\alpha]_{\rm D} = 25^{\circ} (c \ 0.5); R_f \ 0.48$. NMR data (CDCl₃): ¹H, δ 7.83–6.78 (m, 14 H, 2 Ph and Phth), 5.89 (m, 1 H, H₂C=CHCH₂O), 5.30 and 5.16 (2 m, 2 H, H₂C=CHCH₂O), 5.25 (d, 1 H, J_{1,2} 10.0 Hz, H-1), 5.14 (d, 1 H, J_{1",2"} 3.0 Hz, H-1"), 5.02 (t, 1 H, $J_{3''4''}$ 4.9 Hz, H-3"), 4.98 (dd, 1 H, $J_{2''3''}$ 5.0 Hz, H-2"), 4.85 (m, 1 H, $J_{4'',5''ax/eq}$ 3.3 and 3.7 Hz, H-4"), 4.85 and 4.35 (2 d, each 1 H, PhCH₂O), 4.73 and 4.51 (2 d, each 1 H, PhC H_2 O), 4.56 (s, 1 H, H-1'), 4.55 and 3.51 (2 dd, each 1 H, J_{5"ax,5"eq} - 12.9 Hz, H-5"ax, 5"eq), 4.34 (dd, 1 H, J_{3,4} 8.0 Hz, H-3), 4.24 (t, 1 H, J_{2,3} 10.0 Hz, H-2), 4.03 (d, 1 H, J_{2',3'} 3.0 Hz, H-2'), 3.99 (dd, 1 H, J_{4.5} 9.9 Hz, H-4), 3.84 (t, 1 H, $J_{4'.5'}$ 9.5 Hz, H-4'), 3.75 (1 H, $J_{6a,6b}$ –11.5 Hz, H-6a), 3.71 (1 H, $J_{6'a,6'b}$ -10.6 Hz, H-6'a), 3.60 (m, 1 H, $J_{5,6a} = J_{5,6b} \sim 2.5$ Hz, H-5), 3.39 (t, 1 H, H-6'b), 3.17 (dd, 1 H, J_{3'.4'} 9.5 Hz, H-3'), 2.95 (m, 1 H, J_{5'.6'a} 5.1, J_{5'.6'b} 10.3 Hz, H-5'), 2.65 (m, 2 H, CH₃CH₂S), 2.13, 2.10, and 2.00 (3 s, each 3 H, 3 Ac), 1.40 and 1.37 (2 s, each 3 H, CMe₂), 1.18 (t, 3 H, CH_3CH_2S); ¹³C, δ 169.6, 168.8, and 168.7 (COCH₃), 138.6, 137.6, 134.9, 133.5, 131.4, 128.5–126.6, and 123.1 (C₆H₅CH₂O, Phth, and $H_2C=CHCH_2O$, 116.1 ($H_2C=CHCH_2O$), 102.1 (C-1'), 99.2 (Me₂C), 98.4 (C-1"), 80.9, 80.3, 78.7, 78.2, 77.1, 74.0, 70.4, 68.5, 68.2, 68.1, and 67.8 (C-1,3,4,5,2',3',4',5',2",3",4"), 74.5, 73.4, 70.8, 68.8, 61.8, and 59.5 (C-6,6',5", 2 PhCH₂O, and H₂C=CHCH₂O), 54.6 (C-2), 28.9 and 18.9 [(CH₃)₂C], 23.4 (CH₃CH₂S), 20.7 (2 C) and 20.4 (3 COCH₃), 14.7 (CH₃CH₂S). Anal. Calcd for C₅₃H₆₃NO₁₈S: C, 61.56; H, 6.14. Found: C, 61.23; H, 6.09.

Ethyl 4-O-[3-O-allyl-2-O-(2,3,4-tri-O-acetyl- β -D-xylopyranosyl)- β -D-mannopyranosyl]-3,6-di-O-benzyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (17). —A solution of 16 (200 mg, 0.19 mmol) in 6:4 acetic acid-water (4 mL) was kept for 20 min at 60°C, then concentrated, and toluene (5 × 5 mL) was evaporated from the residue to afford 17, as a white glass (186 mg, 97%); [α]_D - 30° (c 0.3);

 $R_f 0.62 (7:3 \text{ CH}_2\text{Cl}_2\text{-acetone})$. NMR data (CDCl₃): ¹H, δ 7.82–6.83 (m, 14 H, 2 Ph and Phth), 5.91 (m, 1 H, H₂C=CHCH₂O), 5.29 (d, 1 H, J₁, 10.2 Hz, H-1), 5.28 and 5.19 (2 m, 2 H, H₂C=CHCH₂O), 5.06 (dd, 1 H, J_{3" 4"} 5.5 Hz, H-3"), 5.02 (d, 1 H, J_{1".2"} 3.5 Hz, H-1"), 4.94 (dd, 1 H, J_{2".3"} 5.5 Hz, H-2"), 4.91 and 4.43 (2 d, each 1 H, PhC H_2 O), 4.84 (m, 1 H, $J_{4'',5''ax/ea}$ 4.5 and 4.6 Hz, H-4"), 4.72 and 4.53 (2 d, each 1 H, PhCH₂O), 4.60 (s, 1 H, H-1'), 4.43 and 3.49 (2 dd, each 1 H, J_{5"ar,5"ea} -12.6 Hz, H-5"ax,5"eq), 4.38 (t, 1 H, J_{3.4} 8.4 Hz, H-3), 4.24 (t, 1 H, J_{2.3} 10.0 Hz, H-2), 4.13 and 3.88 (2 m, 2 H, CH₂=CHCH₂O), 4.02 (d, 1 H, J_{2'3'} 3.0 Hz, H-2'), 3.99 (t, 1 H, H-4), 3.50 (dd, 1 H, $J_{6'a,5'}$ 2.8, $J_{6a',6b'}$ – 11.6 Hz, H-6'a), 3.18 (m, 1 H, $J_{5',4'}$ 9.4, $J_{5',6'b}$ 6.0 Hz, H-5'), 3.07 (dd, 1 H, $J_{3',4'}$ 9.4 Hz, H-3'), 2.65 (m, 2 H, CH₃CH₂S), 2.11, 2.04, and 1.96 (3 s, each 3 H, 3 Ac), 1.18 (t, 3 H, CH₃CH₂S); 13 C, δ 169.7, 169.3, and 169.0 (3 COCH₃), 138.3, 137.8, 134.5, 133.6, 131.6, 128.4-127.0, and 123.2 (C₆H₅CH₂O, Phth, and H₂C=CHCH₂O), 117.1 (H₂C=CHCH₂O), 101.2 (C-1'), 98.9 (C-1"), 81.0, 80.4, 80.1, 78.8, 78.4, 76.1, 72.4, 69.3, 68.7, 68.1, and 66.4 (C-1,3,4,5,2',3',4',5',2",3",4"), 74.7, 73.5, 70.0, 69.4, 62.3, and 60.1 (C-6,6',5", 2 PhCH₂O, and H₂C=CHCH₂O), 54.9 (C-2), 23.6 (CH₃CH₂S), 20.5 (2 C) and 20.4 (COCH₃), 14.8 (CH₃CH₂S). Anal. Calcd for C₅₀H₅₉NO₁₈S · H₂O: C, 59.34; H, 6.07. Found: C, 59.68; H, 5.98.

Ethyl 4-O-(*3*-O-*allyl*-2-O-β-D-*xylopyranosyl*-β-D-*mannopyranosyl*)-*3*,6-*di*-O-*ben*-*zyl*-2-*deoxy*-2-*phthalimido*-1-*thio*-β-D-*glucopyranoside* (**18**).—A solution of **17** (60 mg, 60 μ mol) in methanolic M NaOMe (4 mL) was kept for 16 h, then neutralised with Dowex 50W (H⁺) resin, filtered, and concentrated to give **18**, as a white glass (50 mg, 95%); [α]_D - 26° (*c* 0.2); R_f 0.62 (85 : 15 CH₂Cl₂-MeOH). NMR data (CDCl₃): ¹H, δ 7.83–6.72 (m, 14 H, 2 Ph and Phth), 5.86 (m, 1 H, H₂C=CHCH₂O), 5.21 (d, 1 H, J_{1,2} 10.2 Hz, H-1), 4.77 and 4.49 (2 d, each 1 H, PhCH₂O), 4.58 (s, 1 H, H-1'), 4.49 (d, 1 H, J_{1",2"} 6.7 Hz, H-1"), 4.30 (t, 1 H, J_{3,4} 8.4 Hz, H-3), 4.17 (t, 1 H, J_{2,3} 10.0 Hz, H-2), 2.61 (m, 2 H, CH₃CH₂S), 1.15 (CH₃CH₂S); ¹³C, δ 138.5, 137.9, 134.4, 133.7, 131.6, 128.5–127.0, and 123.4 (C₆H₅CH₂O, Phth, and H₂C=CHCH₂O), 117.8 (H₂C=CHCH₂O), 103.3 (C-1"), 102.2 (C-1'), 81.0, 80.5, 80.1, 78.8 (2 C), 78.6, 76.5, 75.0, 73.0, 72.6, and 69.3 (C-1,3,4,5,2',3',4',5',2",3",4"), 74.8, 73.6 (2 C), 70.3, 69.2, and 61.0 (C-6,6',5", 2 PhCH₂O, and H₂C=CHCH₂O), 54.9 (C-2), 23.7 (CH₃CH₂S), 14.9 (CH₃CH₂S). Anal. Calcd for C₄₄H₅₃NO₁₅S · H₂O: C, 59.65; H, 6.26. Found: C, 59.98; H, 6.05.

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