






ORIGINAL RESEARCH

Plasma Markers of Alzheimer's Disease Pathology, Neuronal Injury, and Astrocytic Activation and MRI Load of Vascular Pathology and Neurodegeneration: The SMART-MR Study

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BACKGROUND: Two of the main causes for dementia are Alzheimer's disease (AD) and vascular pathology, with most patients showing mixed pathology. Plasma biomarkers for Alzheimer's disease-related pathology have recently emerged, including A β (amyloid-beta), p-tau (phosphorylated tau), NfL (neurofilament light), and GFAP (glial fibrillary acidic protein). There is a current gap in the literature regarding whether there is an association between these plasma biomarkers with vascular pathology and neurodegeneration.

METHODS AND RESULTS: Cross-sectional data from 594 individuals (mean [SD] age: 64 [8] years; 17% female) were included from the SMART-MR (Second Manifestations of Arterial Disease-Magnetic Resonance) study, a prospective cohort study of individuals with a history of arterial disease. Plasma markers were assessed using single molecular array assays (Quanterix). Magnetic resonance imaging markers included white matter hyperintensity volume, presence of infarcts (yes/no), total brain volume, and hippocampal volume assessed on 1.5T magnetic resonance imaging. Linear regressions were performed for each standardized plasma marker with white matter hyperintensity volume, total brain volume, and hippocampal volume as separate outcomes, correcting for age, sex, education, and intracranial volume. Logistic regressions were performed for the presence of lacunar and cortical infarcts. Higher p-tau181 was associated with larger white matter hyperintensity volume (b per SD increase=0.16 [95% CI, 0.06–0.26], $P=0.015$). Higher NfL ($b=-5.63$, [95% CI, -8.95 to -2.31], $P=0.015$) was associated with lower total brain volume and the presence of infarcts (odds ratio [OR], 1.42 [95% CI, 1.13–1.78], $P=0.039$). Higher GFAP levels were associated with cortical infarcts (OR, 1.45 [95% CI, 1.09–1.92], $P=0.010$).

CONCLUSIONS: Plasma biomarkers that have been associated with tau pathology, axonal injury, and astrocytic activation are related to magnetic resonance imaging markers of vascular pathology and neurodegeneration in patients with manifest arterial disease.

Key Words: dementia ■ neurodegeneration ■ neuroimaging ■ vascular

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RESEARCH PERSPECTIVE

What Is New?

- In this cross-sectional study in individuals all with manifest arterial disease, plasma biomarkers for tau pathology, axonal injury, and astrocytic activation were related to vascular pathology and neurodegeneration on magnetic resonance imaging.
- This study confirms the role of mixed pathology in individuals with manifest vascular disease.

What Question Should Be Addressed Next?

- Longitudinal studies should be performed to assess if individuals with both pathologies in plasma and magnetic resonance imaging have a higher risk of dementia.

Nonstandard Abbreviations and Acronyms

Aβ	amyloid-beta
GFAP	glial fibrillary acidic protein
NfL	neurofilament light
p-tau	phosphorylated tau
SMART-MR	Second Manifestations of Arterial Disease-Magnetic Resonance
TBV	total brain volume
WMH	white matter hyperintensity

Two of the main causes of dementia are Alzheimer's disease (AD) and vascular pathology.¹ The presence of AD-specific pathology, that is, the accumulation of A β (amyloid-beta) plaques and neurofibrillary tangles, can be established using cerebrospinal fluid or positron emission tomography (PET). Vascular pathology is typically assessed via MRI measures, such as white matter hyperintensities (WMHs) and lacunes.² Most patients with cognitive decline and dementia have mixed pathology,³ as well as hippocampal and global brain atrophy.⁴ The relationship between AD pathology with vascular pathology and neurodegeneration is not yet well known and has been hampered by the invasiveness and costs associated with cerebrospinal fluid and PET measurements.^{5–8}

Recent advancements in the development of high-sensitivity plasma assays have allowed for the assessment of biomarkers representing A β and p-tau (phosphorylated tau) accumulation, as well as axonal injury (NfL [neurofilament light]) and astrocytic activation (GFAP [glial fibrillary acidic protein])

in large-scale studies.⁹ Previous studies have highlighted a possible relation between these plasma biomarkers, specifically p-tau181, NfL, and GFAP with vascular pathology on MRI.^{10–15} However, other studies found no association.^{16–18} Regarding neurodegeneration on MRI, most studies have found that higher levels of p-tau181 and NfL are associated with greater atrophy, either globally^{19,20} or specifically in the hippocampus.^{10,16,18,19,21–23}

However, few studies focused on patients all with vascular, or arterial, disease. Neurodegeneration²⁴ and WMH are common in patients with arterial disease, and many patients show mixed AD and vascular pathology.^{25,26} By focusing on a population with arterial disease, it is possible to shed light on the role of AD pathology in the neurodegeneration and WMH observed in patients with arterial disease. We took as a starting point the SMART-MR (Second Manifestations of Arterial Disease-Magnetic Resonance) study cohort, a population all with manifest arterial disease. We aimed to examine if blood-based biomarkers (ie, A β 42/40, p-tau181, NfL, and GFAP) were associated with MRI measures of vascular pathology and neurodegeneration in individuals with manifest arterial disease and without dementia diagnosis. We hypothesized that NfL, as a general marker of axonal injury, would be related to global and hippocampal atrophy, whereas p-tau181, A β 42/40, and GFAP, as more specifically related to amyloid positivity on PET, would be related to vascular pathology based on previous studies in non-vascular populations.

METHODS

Design of Study and Sample

Data were obtained from the SMART-MR study. The SMART-MR study is a prospective cohort study that aimed to investigate brain MRI changes in patients independently living with symptomatic atherosclerotic disease.^{27,28} All patients that were recently referred to the University Medical Center Utrecht in the Netherlands with manifest arterial disease (ie, cerebrovascular disease, coronary artery disease, peripheral arterial disease, or an abdominal aortic aneurysm [prevalence shown in Table S1]) and without any MRI contraindications were invited to participate between May 2001 and December 2005. For this study, we used cross-sectional data from the second wave of the SMART-MR study (n=754).^{27,29} A further selection was done for biomarker assessment, particularly being 50 years or older, having a brain MRI scan, and available cognitive measurements (n=594). There were no significant differences between the selected group and the rest of the cohort regarding demographics (ie, sex, education, cardiovascular risk factors), other than the

selected group having a higher age. MRI brain scans, physical examinations, blood sampling, and questionnaires were all performed during a 1-day visit at the hospital. Written informed consent was obtained from all participants. A local ethics committee approved the SMART-MR study. The data that support the findings of this study are available from the corresponding author upon reasonable request.

This study was reported in accordance to the Strengthening the Reporting of Observational Studies in Epidemiology checklist.

Plasma Assessment

Briefly, participants underwent venipuncture under overnight-fasting conditions between 2006 and 2009. Plasma was then centrifuged for 10 minutes at 1800g within 2 hours. Then, in polypropylene tubes, plasma was aliquoted in 0.5-mL aliquots and stored at -80 °C until use. Plasma markers were analyzed in 2021. A β 40, A β 42, NfL, and GFAP were all assessed using the Neurology 4-plex E kit (Quanterix).³⁰ P-tau181 was assessed using the V2 Advantage kit (Quanterix). Measurements were performed according to manufacturer's instructions, using automated sample dilution on board of the Simoa HD-X analyzer. The Neurology 4-plex E kit was run in singlicates, and the p-tau181 V2 kit was run in duplicates. We calculated A β 42/40, to adjust for between-person differences in production rates and to correct for preanalytical sample handling effects.³¹ All plasma markers were Z score standardized for analysis.

MRI Protocol

Brain MRI was performed using a 1.5 Tesla whole-body system (Gyrosan ACS-NT, Philips Medical System, Best, The Netherlands).²⁹ A transversal T1-weighted gradient-echo (repetition time [TR]/echo time [TE]: 235/2 milliseconds), T2-weighted (TR/TE: 2200/11 milliseconds), fluid-attenuated inversion recovery (TR/TE/inversion time: 6000/100/2000 milliseconds), and T1-weighted inversion recovery (TR/TE/inversion time: 2900/22/410 milliseconds) sequences were acquired with a voxel size of 1.0×1.0×4.0 mm³ as part of the protocol.²⁹ For measurement of hippocampal volume, a sagittal T1-weighted 3D fast field-echo sequence was obtained (TR/TE: 7/3.2 milliseconds).³²

Brain Segmentation

WMH and brain volumes were segmented using an automated segmentation program on the T1-weighted gradient-echo, the inversion recovery sequence, and the fluid-attenuated inversion recovery sequences. More details regarding the probabilistic segmentation technique can be found here.^{33,34} The hippocampus

was manually outlined by 2 trained investigators, blinded to all clinical information.³² The hippocampus proper, subiculum, fimbria, alveus, and dentate gyrus were all included on an average of 40 slices.³⁵ TBV was defined as the sum of both gray and white matter, WMH, and infarct volumes. Total intracranial volume was the sum of both TBV as well as the volumes of sulcal and ventricular cerebrospinal fluid. Hippocampal volume was the sum of both right and left hippocampi, which were defined by multiplying the total number of voxels by the volume of a voxel (ie, 1.0×0.94×0.94 mm).

Infarcts and WMH

Infarcts were rated visually by both an investigator and a neuroradiologist. Both were blinded to any clinical characteristics, and scans were reevaluated in a consensus meeting.³⁶ WMHs were defined as periventricular and deep lesions and were summed to represent the total volume of WMHs using a fully automated technique and visually checked.^{29,36}

Infarcts were described as focal hyperintensities on T2-weighted images more than 3 mm in diameter and presumed of vascular origin.³⁷ If the T2 hyperintensities were within white matter, they also had to be hypointense on T1-weighted and fluid-attenuated inversion recovery images to be differentiated from WMHs. They were then characterized as lacunar or cortical infarcts. According to the Standards for Reporting Vascular Changes on Neuroimaging criteria,² lacunar infarcts were defined as focal lesions between 3 and 15 mm. Cortical infarcts were defined as being an area of tissue necrosis 4 mm or larger in cortical or cortico-subcortical areas.³⁸ Presence of any infarct was categorized as a dichotomous outcome (ie, yes/no). Lacunar and cortical infarcts were also categorized as any or none.

Covariates

Age, sex, education, smoking status, and alcohol use were given based on self-report. Education was categorized into less than high school education, at least some high school education, or college/university education based on the Dutch education system. Smoking status was categorized as never, former, or current smoker. Alcohol use was categorized as <1 drinks per week, 1 to 10 drinks per week, 11 to 20 drinks per week, and >20 drinks per week. Diabetes was defined as a (self-)reported history of diabetes, registered use of glucose-lowering medication (ie, insulin or oral antidiabetic drugs), or a glucose level of 7 mmol/L or higher. Hypertension was defined as self-reported use of antihypertensive medication, a mean systolic blood pressure of >140 mmHg, or a mean diastolic blood pressure of >90 mmHg.

Statistical Analysis

To address missing values (max: 10% missing on hippocampal volume), multiple imputation was performed using the *mice* package in R. Both outcomes and predictors were imputed if needed, using covariate information as predictors in the imputation.³⁹ We chose 10 imputed data sets, as a complete case analysis would be on 90% of the original sample size.⁴⁰ Pooled results are shown. Kruskal–Wallis tests were done to assess possible differences between sexes in plasma marker levels, and Pearson correlations were performed to assess the relationship between the plasma marker levels, brain volumes, and age. Linear regressions were performed to estimate the association between each standardized plasma marker and log-transformed WMH volume, TBV, and hippocampal volume, with age, sex, education, and intracranial volume as covariates. Logistic regressions were performed for the presence of infarcts (yes versus no), corrected for age, sex, and education. An additional model was performed correcting for further cardiovascular risk factors (ie, diabetes, hypertension, smoking status, and alcohol use). A sensitivity analysis was performed assessing the difference between lacunar and cortical infarcts by performing logistic regressions separately on cortical and lacunar infarcts. Another sensitivity analysis was done on the models of A β 42/40 by adding 1/A β 40 and A β 42 as main effects, as suggested by previous work on the complexities of using a ratio in regression analyses.⁴¹ We also report individual associations of A β 40 and A β 42 on all outcomes. Assumptions for both linear and logistic regressions were checked and met; therefore, no plasma markers were log-transformed. As there was moderate correlation between our outcomes, we used the Hommel method⁴² for multiple comparison adjustment.⁴³

RESULTS

The characteristics of the study population are shown in Table 1. The mean age of the study population was 64 \pm 8 years. Approximately 17% of the individuals were women, and 10% had an education level of a college/university degree. Eight percent met Petersen criteria⁴⁴ for mild cognitive impairment. A Pearson correlation matrix on age, the plasma markers, and the MRI markers can be found in Figure S1. Kruskal–Wallis tests showed no sex differences in A β 42/40 and p-tau181. Higher levels of NfL and GFAP were seen in women compared with men.

All plasma markers were nonnormally distributed. The respective median (10%–90% range) for A β 42/40, p-tau181, GFAP, and NfL were 0.06 (0.05–0.07), 1.37 (0.85–2.54), 86.33 (47.28–151.97), and

Table 1. Baseline Characteristics of the Study Population

	Study sample (n=594)
	Mean (SD) or n (%)
Demographics	
Age, y	64 (8)
Sex, women	101 (17%)
Education, college/university	57 (10%)
Current smoker	127 (21%)
Alcoholic drinks per week	1.14 (0.97)
Diabetes	131 (22%)
Hypertension	401 (68%)
Currently taking antihypertensive medication	461 (78%)
Petersen criteria	48 (8%)
Body mass index, kg/m ² *	27 (5)
High-density lipoprotein, mmol/L*	1.22 (0.47)
Low-density lipoprotein, mmol/L*	2.40 (0.90)
Plasma levels, pg/mL	
A β 40	113.19 (30.64)*
A β 42	6.79 (1.94)*
A β 42/40	0.06 (0.01)*
Phosphorylated-tau181	1.37 (0.79)*
Neurofilament light	13.82 (9.56)*
Glial fibrillary acidic protein	86.33 (53.98)*
Magnetic resonance imaging markers	
White matter lesion volume, mL	1.24 (2.50)*
Hippocampal volume, mL	5.98 (0.75)
Total brain volume, mL	1137.43 (105.14)
Infarct presence	206 (35%)
Lacunar infarct presence	137 (23%)
Cortical infarct presence	83 (14%)

1% missing on education, 1% missing on smoking, 1% missing on alcohol use, 1% missing on diabetes, 1% missing on hypertension, 3% missing on Petersen criteria, and 10% missing on hippocampal volume. A β indicates amyloid-beta.

*Shown as median (interquartile range).

13.82 (7.86–28.59) (shown in pg/mL). For the MRI markers, only WMH volume was nonnormally distributed. The mean (SD) for hippocampal volume and TBV were 5.98 mL (0.75) and 1137 mL (105), respectively. The median (10%–90% range) for WMH volume was 1.24 (0.29–8.12) mL. Thirty-five percent of our study population had at least 1 infarct, with 23% having at least 1 lacunar infarct and 14% having at least 1 cortical infarct.

Associations with MRI Markers of Vascular Pathology

Linear regression analysis showed that higher plasma p-tau181 was associated with higher WMH volume (*b* per SD increase: 0.16 [95% CI, 0.06–0.26], *P*=0.015), which remained after further correction (Table 2).

Table 2. Associations Between the Plasma AD Markers and MRI Markers of Vascular Pathology and Neurodegeneration

Plasma levels (per SD increase)	WMH volume, B (95% CI), adjusted P value	TBV, B (95% CI), adjusted P value	HV, B (95% CI), adjusted P value
Model 1			
Aβ42/40	-0.03 (-0.13 to 0.06), P=0.827	-1.39 (-4.00 to 1.23), P=0.827	0.01 (-0.04 to 0.07), P=0.827
p-tau181	0.16 (0.06 to 0.26), P=0.015*	-2.21 (-5.05 to 0.63), P=0.750	-0.02 (-0.07 to 0.05), P=0.827
NfL	0.17 (0.06 to 0.29), P=0.052	-5.63 (-8.95 to -2.31), P=0.015*	-0.05 (-0.12 to 0.02), P=0.781
GFAP	0.07 (-0.04 to 0.18), P=0.827	-1.14 (-4.31 to 2.04), P=0.827	-0.04 (-0.11 to 0.03), P=0.827
Model 2			
Aβ42/40	-0.04 (-0.13 to 0.05), P=0.785	-1.26 (-3.81 to 1.29), P=0.785	0.01 (-0.04 to 0.07), P=0.785
p-tau181	0.15 (0.06 to 0.25), P=0.028*	-1.76 (-4.54 to 1.01), P=0.785	-0.01 (-0.07 to 0.05), P=0.785
NfL	0.17 (0.05 to 0.29), P=0.052	-5.50 (-8.75 to -2.24), P=0.016*	-0.05 (-0.12 to 0.02), P=0.785
GFAP	0.09 (-0.03 to 0.20), P=0.738	-2.07 (-5.23 to 1.09), P=0.785	-0.05 (-0.12 to 0.02), P=0.738

Model 1 is adjusted for age, sex, education, and intracranial volume. Model 2 adds diabetes, hypertension, smoking status, and alcohol use. WMH volume is log-transformed. P values are adjusted using the Hommel method. Aβ indicates amyloid-beta; AD, Alzheimer’s disease; GFAP, glial fibrillary acidic protein; HV, hippocampal volume; MRI, magnetic resonance imaging; NfL, neurofilament light; p-tau, phosphorylated tau; TBV, total brain volume; and WMH, white matter hyperintensity.

*P values shown are corrected using the Hommel method, with P<0.05 denoting significance.

Higher NfL showed a trend toward higher WMH volume, but it did not survive correction for multiple comparisons. No other biomarkers were associated with WMH, also when adding 1/Aβ40 and Aβ42 as main effects or assessed individually (Table S2).

Regarding infarcts, higher plasma NfL was associated with higher odds of having an infarct (OR, 1.42 [95% CI, 1.13–1.78], P=0.039); however, this did not remain after further covariate adjustment (Table 3). When looking specifically at lacunar infarcts, higher plasma NfL was associated with higher odds of a lacunar infarct (OR, 1.36 [95% CI, 1.06–1.73], P=0.014). Regarding cortical infarcts, both higher plasma NfL and GFAP were associated with higher odds of having a cortical infarct (respectively OR, 1.58 [95% CI, 1.20–2.08], P=0.001 and OR, 1.45 [95% CI, 1.09–1.92], P=0.010) (Table 3). No other plasma markers were associated with the presence of infarcts (Table 3; Table S3).

Associations with MRI Markers of Neurodegeneration

Higher plasma NfL was associated with lower TBV (b: -5.63 [95% CI, -8.95 to -2.31], P=0.015), which remained after further covariate adjustment (Table 2). There were no associations between plasma markers and hippocampal volume, albeit an association was found for higher Aβ40 and lower hippocampal volume (Table S2).

DISCUSSION

In a sample of individuals with manifest arterial disease and WMH, we found that higher NfL was associated with infarcts and global brain atrophy, higher p-tau181 was associated with more WMH volume, and GFAP was related to the presence of 1 or more cortical

Table 3. Associations Between the Plasma AD Markers and Infarcts

Plasma levels (per SD increase)	Infarcts, OR (95% CI), adjusted P value	Lacunar infarcts, OR (95% CI), P value	Cortical infarcts, OR (95% CI), P value
Model 1			
Aβ42/40	1.18 (0.86–1.63), P=0.827	1.30 (0.84–2.02), P=0.240	1.04 (0.85–1.26), P=0.723
p-tau181	1.16 (0.96–1.40), P=0.729	1.22 (1.00–1.50), P=0.055	1.17 (0.92–1.50), P=0.202
NfL	1.42 (1.13–1.78), P=0.039*	1.36 (1.06–1.73), P=0.014*	1.58 (1.20–2.08), P=0.001*
GFAP	0.98 (0.79–1.21), P=0.827	0.86 (0.68–1.10), P=0.224	1.45 (1.09–1.92), P=0.010*
Model 2			
Aβ42/40	1.17 (0.83–1.64), P=0.785	1.27 (0.82–1.95), P=0.288	1.01 (0.84–1.23), P=0.883
p-tau181	1.13 (0.93–1.37), P=0.785	1.19 (0.97–1.47), P=0.101	1.13 (0.88–1.45), P=0.342
NfL	1.41 (1.12–1.79), P=0.052	1.36 (1.05–1.74), P=0.018*	1.54 (1.16–2.03), P=0.003*
GFAP	1.03 (0.82–1.30), P=0.785	0.87 (0.68–1.13), P=0.297	1.56 (1.16–2.10), P=0.003*

Model 1 is adjusted for age, sex, and education. Model 2 adds diabetes, hypertension, smoking status, and alcohol use. P values are adjusted using the Hommel method. Aβ indicates amyloid-beta; AD, Alzheimer’s disease; GFAP, glial fibrillary acidic protein; NfL, neurofilament light; OR, odds ratio; and p-tau, phosphorylated tau.

*P values shown are corrected using the Hommel method, with P<0.05 denoting significance.

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infarcts. None of the biomarkers were associated with hippocampal volume.

Higher levels of plasma NfL were associated with infarcts, which is in line with previous literature.¹² As NfL has been linked to axonal damage,⁴⁵ the relationship between NfL and infarcts could be explained by persisting axonal damage due to vascular pathology.⁴⁶ Additionally, we found an association between higher NfL and lower TBV, which is in line with a previous longitudinal study.²⁰ However, we did not find an association of plasma NfL with hippocampal atrophy, whereas some studies found an association.^{18,22,23,47} Further, in this cohort, memory has not been associated with hippocampal volume.⁴⁸ Hippocampal atrophy in vascular patients may have a different pathological mechanism than global atrophy and may not show a clear relation to memory decline. As this is the first study to our knowledge focusing on plasma AD biomarkers and brain volume in individuals with arterial disease, this needs to be validated in other studies of a similar population. We did not find an association after correction for multiple comparisons between NfL and WMH, which is in line with a former study on a population with cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy.¹⁷ In contrast, studies in populations with mild cognitive impairment and AD have reported such an association.^{13,14,49,50} This apparent contradiction may be explained by being in a later stage of cognitive impairment or may be due to a loss of power in the current study, as our findings were approaching significance. As studies assessing NfL and MRI markers of vascular pathology are scarce, further studies need to be performed to replicate this result.

However, higher p-tau181 was associated with more WMH volume, which is in line with previous studies.^{10,11} One study found that amyloid PET pathology was associated with WMH in the general population, with cerebral amyloid angiopathy possibly explaining this role.⁵¹ As p-tau181 in plasma is associated with amyloid PET positivity,⁵² cerebral amyloid angiopathy could explain the relation between AD-specific biomarkers and WMH. Surprisingly, we did not find an association with p-tau181 and hippocampal volume, even though many previous studies have found an association.^{10,16,19,21,22} However, 2 studies also found a null association.^{53,54} Hippocampal atrophy in this population with manifest arterial disease may be independent to AD-specific neuronal loss and solely of vascular origin.

Additionally, GFAP was associated with cortical infarcts. This is in line with a previous study on serum GFAP that found an association of GFAP with infarcts, but not with subcortical vascular pathology such as WMHs.⁴⁶ The specificity to cortical infarcts was also reflected in a previous study, highlighting that the site of injury may determine if GFAP is released into the blood.⁵⁵ However, as this is the first study to our

knowledge assessing GFAP in plasma with brain infarcts, future research should validate these findings. A β and p-tau181 were not associated with infarcts, which is in line with a previous study.⁵ Infarcts may not have a direct relationship to AD pathology, possibly due to the anatomical location of infarcts compared with WMH.⁵⁶

The current study had important strengths. We assessed multiple plasma markers using an ultrasensitive Simoa assay. Further, we used multiple imputation to account for missing data, corrected for multiple covariates, and also used a strict correction for multiple comparisons to prevent any Type I errors. However, the study also had limitations. The study population was relatively young and healthy, as the included participants were a subsample of the SMART-MR study that participated in the first follow-up assessment. Further, participants were predominantly White and male; thus, the generalizability to other populations is low. Other studies have highlighted that plasma p-tau181 and NfL may not accurately represent brain amyloidosis in Black adults compared with White individuals.⁵⁷ Future studies need to be done on marginally underrepresented individuals to assess any differences regarding plasma AD biomarkers and vascular pathology in these populations. Further, plasma biomarker assessment was performed more than 12 years after blood was collected and stored. Although studies have shown the reliability of blood biomarker assessment after long-term storage, slight increases in variability can be expected after 14 years.⁵⁸ Additionally, we did not have information on microbleeds, so we unfortunately could not assess the relation between the plasma AD markers on cerebral microbleeds.

CONCLUSIONS

Plasma biomarkers that have been associated with tau accumulation, axonal injury, and astrocytic activation are related to MRI markers of vascular pathology and neurodegeneration in patients with manifest arterial disease. The current study suggests a relationship between AD-related and vascular pathology in individuals with manifest arterial disease, highlighting the role of mixed pathology in these individuals. Future longitudinal studies should explore if individuals with both of these pathologies are at an increased risk of dementia.

ARTICLE INFORMATION

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Disclosures

None.

Supplemental Material

Tables S1–S3
Figure S1

REFERENCES

- van der Flier WM, Skoog I, Schneider JA, Pantoni L, Mok V, Chen CLH, Scheltens P. Vascular cognitive impairment. *Nat Rev Dis Primers*. 2018;4:18003. doi: [10.1038/nrdp.2018.3](https://doi.org/10.1038/nrdp.2018.3)
- Wardlaw JM, Smith EE, Biessels GJ, Cordonnier C, Fazekas F, Frayne R, Lindley RI, O'Brien JT, Barkhof F, Benavente OR, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol*. 2013;12:822–838. doi: [10.1016/s1474-4422\(13\)70124-8](https://doi.org/10.1016/s1474-4422(13)70124-8)
- Jellinger KA. Neuropathology of the Alzheimer's continuum: an update. *Free Neuropathol*. 2020;1:1. doi: [10.17879/freeneuropathology-2020-3050](https://doi.org/10.17879/freeneuropathology-2020-3050)
- Karas GB, Scheltens P, Rombouts SA, Visser PJ, van Schijndel RA, Fox NC, Barkhof F. Global and local gray matter loss in mild cognitive impairment and Alzheimer's disease. *Neuroimage*. 2004;23:708–716. doi: [10.1016/j.neuroimage.2004.07.006](https://doi.org/10.1016/j.neuroimage.2004.07.006)
- Kester MI, Goos JDC, Teunissen CE, Benedictus MR, Bouwman FH, Wattjes MP, Barkhof F, Scheltens P, van der Flier WM. Associations between cerebral small-vessel disease and Alzheimer disease pathology as measured by cerebrospinal fluid biomarkers. *JAMA Neurol*. 2014;71:855–862. doi: [10.1001/jamaneurol.2014.754](https://doi.org/10.1001/jamaneurol.2014.754)
- Kim HW, Hong J, Jeon JC. Cerebral small vessel disease and Alzheimer's disease: a review. *Front Neurol*. 2020;11:927. doi: [10.3389/fneur.2020.00927](https://doi.org/10.3389/fneur.2020.00927)
- Hertze J, Palmqvist S, Minthon L, Hansson O. Tau pathology and parietal white matter lesions have independent but synergistic effects on early development of Alzheimer's disease. *Dement Geriatr Cogn Dis Extra*. 2013;3:113–122. doi: [10.1159/000348353](https://doi.org/10.1159/000348353)
- Twait EL, Min B, Beran M, Vonk JMJ, Geerlings MI. The cross-sectional association between amyloid burden and white matter hyperintensities in older adults without cognitive impairment: a systematic review and meta-analysis. *Ageing Res Rev*. 2023;88:101952. doi: [10.1016/j.arr.2023.101952](https://doi.org/10.1016/j.arr.2023.101952)
- Benussi A, Cantoni V, Rivolta J, Archetti S, Micheli A, Ashton N, Zetterberg H, Blennow K, Borroni B. Classification accuracy of blood-based and neurophysiological markers in the differential diagnosis of Alzheimer's disease and frontotemporal lobar degeneration. *Alzheimers Res Ther*. 2022;14:155. doi: [10.1186/s13195-022-01094-5](https://doi.org/10.1186/s13195-022-01094-5)
- Wang YL, Chen J, Du ZL, Weng H, Zhang Y, Li R, Jia Z, Sun M, Jiang J, Wang FZ, et al. Plasma p-tau181 level predicts neurodegeneration and progression to Alzheimer's dementia: a longitudinal study. *Front Neurol*. 2021;12:695696. doi: [10.3389/fneur.2021.695696](https://doi.org/10.3389/fneur.2021.695696)
- Mielke MM, Frank RD, Dage JL, Jeromin A, Ashton NJ, Blennow K, Karikari TK, Vanmechelen E, Zetterberg H, Algeciras-Schimnich A, et al. Comparison of plasma phosphorylated tau species with amyloid and tau positron emission tomography, neurodegeneration, vascular pathology, and cognitive outcomes. *JAMA Neurol*. 2021;78:1108–1117. doi: [10.1001/jamaneurol.2021.2293](https://doi.org/10.1001/jamaneurol.2021.2293)
- Qu Y, Tan CC, Shen XN, Li HQ, Cui M, Tan L, Dong Q, Yu JT. Association of plasma neurofilament light with small vessel disease burden in non-demented elderly: a longitudinal study. *Stroke*. 2021;52:896–904. doi: [10.1161/strokeaha.120.030302](https://doi.org/10.1161/strokeaha.120.030302)
- Elahi FM, Casaletto KB, La Joie R, Walters SM, Harvey D, Wolf A, Edwards L, Rivera-Contreras W, Karydas A, Cobigo Y, et al. Plasma biomarkers of astrocytic and neuronal dysfunction in early- and late-onset Alzheimer's disease. *Alzheimers Dement*. 2020;16:681–695. doi: [10.1016/j.jalz.2019.09.004](https://doi.org/10.1016/j.jalz.2019.09.004)
- Sun Y, Tan L, Xu W, Wang ZT, Hu H, Li JQ, Dong Q, Tan L, Yu JT. Plasma neurofilament light and longitudinal progression of White matter hyperintensity in elderly persons without dementia. *J Alzheimers Dis*. 2020;75:729–737. doi: [10.3233/jad-200022](https://doi.org/10.3233/jad-200022)
- Shir D, Graff-Radford J, Hofrenning EI, Lesnick TG, Przybelski SA, Lowe VJ, Knopman DS, Petersen RC, Jack CR Jr, Vemuri P, et al. Association of plasma glial fibrillary acidic protein (GFAP) with neuroimaging of Alzheimer's disease and vascular pathology. *Alzheimers Dement (Amst)*. 2022;14:e12291. doi: [10.1002/dad2.12291](https://doi.org/10.1002/dad2.12291)
- Chong JR, Ashton NJ, Karikari TK, Tanaka T, Saridin FN, Reilhac A, Robins EG, Nai YH, Vrooman H, Hilal S, et al. Plasma P-tau181 to Aβ42 ratio is associated with brain amyloid burden and hippocampal atrophy in an Asian cohort of Alzheimer's disease patients with concomitant cerebrovascular disease. *Alzheimers Dement*. 2021;17:1649–1662. doi: [10.1002/alz.12332](https://doi.org/10.1002/alz.12332)
- Chen CH, Cheng YW, Chen YF, Tang SC, Jeng JS. Plasma neurofilament light chain and glial fibrillary acidic protein predict stroke in CADASIL. *J Neuroinflammation*. 2020;17:124. doi: [10.1186/s12974-020-01813-5](https://doi.org/10.1186/s12974-020-01813-5)
- Mattsson N, Cullen NC, Andreasson U, Zetterberg H, Blennow K. Association between longitudinal plasma neurofilament light and neurodegeneration in patients with Alzheimer disease. *JAMA Neurol*. 2019;76:791–799. doi: [10.1001/jamaneurol.2019.0765](https://doi.org/10.1001/jamaneurol.2019.0765)
- Chen SD, Huang YY, Shen XN, Guo Y, Tan L, Dong Q, Yu JT. Longitudinal plasma phosphorylated tau 181 tracks disease progression in Alzheimer's disease. *Transl Psychiatry*. 2021;11:356. doi: [10.1038/s41398-021-01476-7](https://doi.org/10.1038/s41398-021-01476-7)
- Benedet AL, Leuzy A, Pascoal TA, Ashton NJ, Mathotaarachchi S, Savard M, Therriault J, Kang MS, Chamoun M, Schöhl M, et al. Stage-specific links between plasma neurofilament light and imaging biomarkers of Alzheimer's disease. *Brain*. 2020;143:3793–3804. doi: [10.1093/brain/awaa342](https://doi.org/10.1093/brain/awaa342)
- Karikari TK, Pascoal TA, Ashton NJ, Janelidze S, Benedet AL, Rodriguez JL, Chamoun M, Savard M, Kang MS, Therriault J, et al. Blood phosphorylated tau 181 as a biomarker for Alzheimer's disease: a diagnostic performance and prediction modelling study using data from four prospective cohorts. *Lancet Neurol*. 2020;19:422–433. doi: [10.1016/s1474-4422\(20\)30071-5](https://doi.org/10.1016/s1474-4422(20)30071-5)
- Huang Y, Li Y, Xie F, Guo Q. Associations of plasma phosphorylated tau181 and neurofilament light chain with brain amyloid burden and cognition in objectively defined subtle cognitive decline patients. *CNS Neurosci Ther*. 2022;28:2195–2205. doi: [10.1111/cns.13962](https://doi.org/10.1111/cns.13962)

23. Barker W, Quinonez C, Greig MT, Behar R, Chirinos C, Rodriguez RA, Rosselli M, Rodriguez MJ, Cid RC, Rundek T, et al. Utility of plasma neurofilament light in the 1Florida Alzheimer's Disease Research Center (ADRC). *J Alzheimers Dis*. 2021;79:59–70. doi: [10.3233/jad-200901](https://doi.org/10.3233/jad-200901)
24. Tan L, Xing J, Wang Z, Du X, Luo R, Wang J, Zhao J, Zhao W, Yin C. Study of gray matter atrophy pattern with subcortical ischemic vascular disease-vascular cognitive impairment no dementia based on structural magnetic resonance imaging. *Front Aging Neurosci*. 2023;15:1051177. doi: [10.3389/fnagi.2023.1051177](https://doi.org/10.3389/fnagi.2023.1051177)
25. Attems J, Jellinger KA. The overlap between vascular disease and Alzheimer's disease—lessons from pathology. *BMC Med*. 2014;12:206. doi: [10.1186/s12916-014-0206-2](https://doi.org/10.1186/s12916-014-0206-2)
26. Jellinger KA, Attems J. Prevalence and pathogenic role of cerebrovascular lesions in Alzheimer disease. *J Neurol Sci*. 2005;229–230:37–41. doi: [10.1016/j.jns.2004.11.018](https://doi.org/10.1016/j.jns.2004.11.018)
27. Geerlings MI, Appelman AP, Vincken KL, Algra A, Witkamp TD, Mali WP, van der Graaf Y. Brain volumes and cerebrovascular lesions on MRI in patients with atherosclerotic disease. The SMART-MR Study. *Atherosclerosis*. 2010;210:130–136. doi: [10.1016/j.atherosclerosis.2009.10.039](https://doi.org/10.1016/j.atherosclerosis.2009.10.039)
28. Castelijns MC, Helming MAG, Hageman SHJ, Asselbergs FW, de Borst GJ, Bots ML, Cramer MJ, Dorresteijn JAN, Emmelot-Vonk MH, Geerlings MI, et al. Cohort profile: the Utrecht Cardiovascular Cohort-Second Manifestations of Arterial Disease (UCC-SMART) study—an ongoing prospective cohort study of patients at high cardiovascular risk in the Netherlands. *BMJ Open*. 2023;13:e066952. doi: [10.1136/bmjopen-2022-066952](https://doi.org/10.1136/bmjopen-2022-066952)
29. Ghaznawi R, Geerlings MI, Jaarsma-Coes M, Hendrikse J, de Bresser J. Association of white matter hyperintensity markers on MRI and long-term risk of mortality and ischemic stroke: the SMART-MR study. *Neurology*. 2021;96:e2172–e2183. doi: [10.1212/wnl.00000000000011827](https://doi.org/10.1212/wnl.00000000000011827)
30. Thijssen EH, Verberk IMW, Kindermans J, Abramian A, Vanbrabant J, Ball AJ, Pijnenburg Y, Lemstra AW, van der Flier WM, Stoops E, et al. Differential diagnostic performance of a panel of plasma biomarkers for different types of dementia. *Alzheimers Dement (Amst)*. 2022;14:e12285. doi: [10.1002/dad2.12285](https://doi.org/10.1002/dad2.12285)
31. Hansson O, Lehmann S, Otto M, Zetterberg H, Lewczuk P. Advantages and disadvantages of the use of the CSF amyloid β (A β) 42/40 ratio in the diagnosis of Alzheimer's disease. *Alzheimers Res Ther*. 2019;11:34. doi: [10.1186/s13195-019-0485-0](https://doi.org/10.1186/s13195-019-0485-0)
32. Knoops AJG, van der Graaf Y, Appelman APA, Mali WPTM, Geerlings MI. Total cerebral blood flow and hippocampal volume in patients with arterial disease. The SMART-MR study. *J Cereb Blood Flow Metab*. 2009;29:1727–1733. doi: [10.1038/jcbfm.2009.91](https://doi.org/10.1038/jcbfm.2009.91)
33. Anbeek P, Vincken KL, van Bochove GS, van Osch MJ, van der Grond J. Probabilistic segmentation of brain tissue in MR imaging. *Neuroimage*. 2005;27:795–804. doi: [10.1016/j.neuroimage.2005.05.046](https://doi.org/10.1016/j.neuroimage.2005.05.046)
34. Ghaznawi R, Zwartbol MH, Zuithoff NP, Bresser J, Hendrikse J, Geerlings MI. Reduced parenchymal cerebral blood flow is associated with greater progression of brain atrophy: the SMART-MR study. *J Cereb Blood Flow Metab*. 2021;41:1229–1239. doi: [10.1177/0271678x20948614](https://doi.org/10.1177/0271678x20948614)
35. Knoops AJ, Gerritsen L, van der Graaf Y, Mali WP, Geerlings MI. Basal hypothalamic pituitary adrenal axis activity and hippocampal volumes: the SMART-MR study. *Biol Psychiatry*. 2010;67:1191–1198. doi: [10.1016/j.biopsych.2010.01.025](https://doi.org/10.1016/j.biopsych.2010.01.025)
36. Rissanen I, Lucci C, Ghaznawi R, Hendrikse J, Kappelle LJ, Geerlings MI. Association of ischemic imaging phenotype with progression of brain atrophy and cerebrovascular lesions on MRI: the SMART-MR study. *Neurology*. 2021;97:e1063–e1074. doi: [10.1212/wnl.00000000000012539](https://doi.org/10.1212/wnl.00000000000012539)
37. Ghaznawi R, Vonk JMJ, Zwartbol MHT, Bresser J, Rissanen I, Hendrikse J, Geerlings MI. Low-grade carotid artery stenosis is associated with progression of brain atrophy and cognitive decline. The SMART-MR study. *J Cereb Blood Flow Metab*. 2022;43:309–318. doi: [10.1177/0271678x221133859](https://doi.org/10.1177/0271678x221133859)
38. Zwartbol MH, van der Kolk AG, Kuijff HJ, Witkamp TD, Ghaznawi R, Hendrikse J, Geerlings MI. Intracranial vessel wall lesions on 7T MRI and MRI features of cerebral small vessel disease: the SMART-MR study. *J Cereb Blood Flow Metab*. 2021;41:1219–1228. doi: [10.1177/0271678x20958517](https://doi.org/10.1177/0271678x20958517)
39. Groenewold RH, Donders AR, Roes KC, Harrell FE Jr, Moons KG. Dealing with missing outcome data in randomized trials and observational studies. *Am J Epidemiol*. 2012;175:210–217. doi: [10.1093/aje/kwr302](https://doi.org/10.1093/aje/kwr302)
40. White IR, Royston P, Wood AM. Multiple imputation using chained equations: issues and guidance for practice. *Stat Med*. 2011;30:377–399. doi: [10.1002/sim.4067](https://doi.org/10.1002/sim.4067)
41. Kronmal RA. Spurious correlation and the fallacy of the ratio standard revisited. *J R Stat Soc A Stat*. 1993;156:379–392. doi: [10.2307/2983064](https://doi.org/10.2307/2983064)
42. Hommel G. A stagewise rejective multiple test procedure based on a modified Bonferroni test. *Biometrika*. 1988;75:383–386. doi: [10.1093/biomet/75.2.383](https://doi.org/10.1093/biomet/75.2.383)
43. Vickerstaff V, Omar RZ, Ambler G. Methods to adjust for multiple comparisons in the analysis and sample size calculation of randomised controlled trials with multiple primary outcomes. *BMC Med Res Methodol*. 2019;19:129. doi: [10.1186/s12874-019-0754-4](https://doi.org/10.1186/s12874-019-0754-4)
44. Petersen RC. Mild cognitive impairment as a diagnostic entity. *J Intern Med*. 2004;256:183–194. doi: [10.1111/j.1365-2796.2004.01388.x](https://doi.org/10.1111/j.1365-2796.2004.01388.x)
45. Teunissen CE, Khalil M. Neurofilaments as biomarkers in multiple sclerosis. *Mult Scler*. 2012;18:552–556. doi: [10.1177/1352458512443092](https://doi.org/10.1177/1352458512443092)
46. Gatringer T, Pinter D, Enzinger C, Seifert-Held T, Kneihls M, Fandler S, Pichler A, Barro C, Gröbke S, Voortman M, et al. Serum neurofilament light is sensitive to active cerebral small vessel disease. *Neurology*. 2017;89:2108–2114. doi: [10.1212/wnl.0000000000004645](https://doi.org/10.1212/wnl.0000000000004645)
47. Chen Y, Theriault J, Luo J, Ba M, Zhang H. Neurofilament light as a biomarker of axonal degeneration in patients with mild cognitive impairment and Alzheimer's disease. *J Integr Neurosci*. 2021;20:861–870. doi: [10.31083/j.jin2004088](https://doi.org/10.31083/j.jin2004088)
48. Wisse LEM, de Bresser J, Geerlings MI, Reijmer YD, Portegies MLP, Brundel M, Kappelle LJ, van der Graaf Y, Biessels GJ. Global brain atrophy but not hippocampal atrophy is related to type 2 diabetes. *J Neurol Sci*. 2014;344:32–36. doi: [10.1016/j.jns.2014.06.008](https://doi.org/10.1016/j.jns.2014.06.008)
49. Walsh P, Sudre CH, Fiford CM, Ryan NS, Lashley T, Frost C, Barnes J. The age-dependent associations of white matter hyperintensities and neurofilament light in early- and late-stage Alzheimer's disease. *Neurobiol Aging*. 2021;97:10–17. doi: [10.1016/j.neurobiolaging.2020.09.008](https://doi.org/10.1016/j.neurobiolaging.2020.09.008)
50. Chong JR, Hilal S, Ashton NJ, Karikari TK, Reilhac A, Vrooman H, Schöll M, Zetterberg H, Blennow K, Chen CP, et al. Brain atrophy and white matter hyperintensities are independently associated with plasma neurofilament light chain in an Asian cohort of cognitively impaired patients with concomitant cerebral small vessel disease. *Alzheimers Dement (Amst)*. 2023;15:e12396. doi: [10.1002/dad2.12396](https://doi.org/10.1002/dad2.12396)
51. Graff-Radford J, Arenaza-Urquijo EM, Knopman DS, Schwarz CG, Brown RD, Rabinstein AA, Gunter JL, Senjem ML, Przybelski SA, Lesnick T, et al. White matter hyperintensities: relationship to amyloid and tau burden. *Brain*. 2019;142:2483–2491. doi: [10.1093/brain/awz162](https://doi.org/10.1093/brain/awz162)
52. Shen XN, Huang YY, Chen SD, Guo Y, Tan L, Dong Q, Yu JT. Plasma phosphorylated-tau181 as a predictive biomarker for Alzheimer's amyloid, tau and FDG PET status. *Transl Psychiatry*. 2021;11:585. doi: [10.1038/s41398-021-01709-9](https://doi.org/10.1038/s41398-021-01709-9)
53. Chatterjee P, Pedrini S, Ashton NJ, Tegg M, Goozee K, Singh AK, Karikari TK, Simrén J, Vanmechelen E, Armstrong NJ, et al. Diagnostic and prognostic plasma biomarkers for preclinical Alzheimer's disease. *Alzheimers Dement*. 2022;18:1141–1154. doi: [10.1002/alz.12447](https://doi.org/10.1002/alz.12447)
54. Hu H, Chen KL, Ou YN, Cao XP, Chen SD, Cui M, Dong Q, Tan L, Yu JT. Neurofilament light chain plasma concentration predicts neurodegeneration and clinical progression in nondemented elderly adults. *Aging (Albany NY)*. 2019;11:6904–6914. doi: [10.18632/aging.102220](https://doi.org/10.18632/aging.102220)
55. Foerch C, Niessner M, Back T, Bauerle M, De Marchis GM, Ferbert A, Grehl H, Hamann GF, Jacobs A, Kastrup A, et al. Diagnostic accuracy of plasma glial fibrillary acidic protein for differentiating intracerebral hemorrhage and cerebral ischemia in patients with symptoms of acute stroke. *Clin Chem*. 2012;58:237–245. doi: [10.1373/clinchem.2011.172676](https://doi.org/10.1373/clinchem.2011.172676)
56. Román GC, Erkinjuntti T, Wallin A, Pantoni L, Chui HC. Subcortical ischaemic vascular dementia. *Lancet Neurol*. 2002;1:426–436. doi: [10.1016/s1474-4422\(02\)00190-4](https://doi.org/10.1016/s1474-4422(02)00190-4)
57. Schindler SE, Karikari TK, Ashton NJ, Henson RL, Yarasheski KE, West T, Meyer MR, Kirmess KM, Li Y, Saef B, et al. Effect of race on prediction of brain amyloidosis by plasma A β 42/A β 40, phosphorylated tau, and neurofilament light. *Neurology*. 2022;99:e245–e257. doi: [10.1212/wnl.0000000000000358](https://doi.org/10.1212/wnl.0000000000000358)
58. Schubert CR, Paulsen AJ, Pinto AA, Merten N, Cruickshanks KJ. Effect of long-term storage on the reliability of blood biomarkers for Alzheimer's disease and neurodegeneration. *J Alzheimers Dis*. 2022;85:1021–1029. doi: [10.3233/jad-215096](https://doi.org/10.3233/jad-215096)