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## Nucicla umbiliphora gen. et sp. nov.: a Quaternary peridinioid dinoflagellate cyst from the Antarctic margin

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#### **ABSTRACT**

In the southern high latitudes, dinoflagellate cysts are an important microfossil group for both biostratigraphic and palaeoenvironmental interpretations purposes. In light of this, the peridinioid dinoflagellate cyst Nucicla umbiliphora gen. et sp. nov. from the Antarctic margin is formally described. Nucicla is dorsoventrally compressed, has a rounded pentagonal outline in dorso-ventral view, an epicyst that is only half as high as the hypocyst, an unusual archaeopyle formed by the loss of the three anterior intercalary plates, and a posterior sulcal plate that is positioned at the antapex. The species N. umbiliphora is characterised by a scabrate cyst wall and possesses undulated and/or crenulated folds/ridges. It has been so far exclusively found in Quaternary sediments obtained from the East Antarctic continental shelf and the Ross Sea. Although the dinoflagellate producing this cyst is as yet unknown, its brown color and the lack of autofluorescence suggest that the motile cell is likely a heterotrophic *Protoperidinium* species. As such, N. umbiliphora might benefit from the phytoplankton blooms occurring close to the Antarctic margin after seasonal sea-ice retreat.

#### **KEYWORDS**

protoperidinioid; dinoflagellate cyst; East Antarctica; Antarctic shelf; Ouaternary

#### 1. Introduction

Although the Oligocene to Quaternary sediments of the Southern Ocean and Antarctic margin had long been thought to contain no dinoflagellate cysts (McMinn 1995), several endemic and bipolar species have been discovered during the last two decades. They are important for both biostratigraphical purposes and reconstructing high-latitude climatic evolution (Bijl et al. 2018; Marret & De Vernal 1997; Montresor et al. 1999; Esper & Zonneveld 2002; Prebble et al. 2013; Clowes et al. 2016).

Here we formally describe Nucicla umbiliphora gen. et sp. nov. This species has already been reported without formal description from four localities around Antarctica (Figure 1). These dinoflagellate cysts are now included in N. umbiliphora. It was first depicted as Dinocyst sp. A from Quaternary samples from the Cape Roberts Project drill core 1 (CRP-1), Ross Sea (Wrenn et al. 1998). Storkey (2006) reported the species from shelf surface sediments in Prydz Bay. Furthermore, the dinoflagellate cyst is depicted in Warny et al. (2006) as 'Lejeunecysta cf. sp. 1 and 5 of CRP' (i.e. cf. Hannah et al. 2000) from the Ross Sea shelf edge. Finally, the species has also been reported from a Holocene core from a small meromictic basin upstream Ellis Fjord, which lies at the eastern coastal margin of Prydz Bay (Boere et al. 2009). Here we add occurrences of this species in nine other East Antarctic marine sediment cores and surface sediments (Figure 1; Table 1).

#### 2. Material and methods

Samples obtained from nine cores (Table 1; Figure 1) were freeze-dried and crushed manually to small fragments in a mortar, after which a Lycopodium tablet was added with Agepon (1:200). Agepon was used to expand the shrivelled Lycopodium from the tablets, and the palynomorphs in general. A small amount of 30% cold hydrochloric acid (HCI) was added to remove carbonate, and to dissolve the Lycopodium tablet. Samples were treated with 38% cold hydrofluoric acid (HF) and shaken for 2 hours at 250 rpm. Thereafter, samples were diluted with tap water, allowed to settle for 24 hours, and decanted. Subsequently, 30% HCl was added to remove fluoride gels, tap water was added and the samples were centrifuged, after which the samples were decanted. No additional neutralisation steps were performed to remove excess HF. Tap water was added to the residue to prevent oxidation of the organic material. The treated material was sieved using a 10- $\mu$ m mesh sieve and kerogen clumps were ultrasonically fragmented. Heavy minerals were removed by pouring the residue from the sieve into a porcelain dish that is kept floating in the ultrasonic bath, and by decanting this back into the sieve after 5 minutes. The fraction > 10  $\mu$ m was concentrated into  $\sim$ 1 mL of glycerin-water mixture and a fraction thereof was mounted on a microscope slide using glycerine jelly.

Green autofluorescence of the cyst was tested on a Leica DM2500 LED fluorescence microscope. Images were taken using a Leica DM2500 LED microscope with mounted Leica MC170 HD camera, and for the images of Plate 1 the Live Image Builder (LIB) within the Leica Application Suite software 4.0 was used, which is a live z-stacking tool. Z-stacking constructs a two-dimensional image from a threedimensional object by combining the areas in focus from

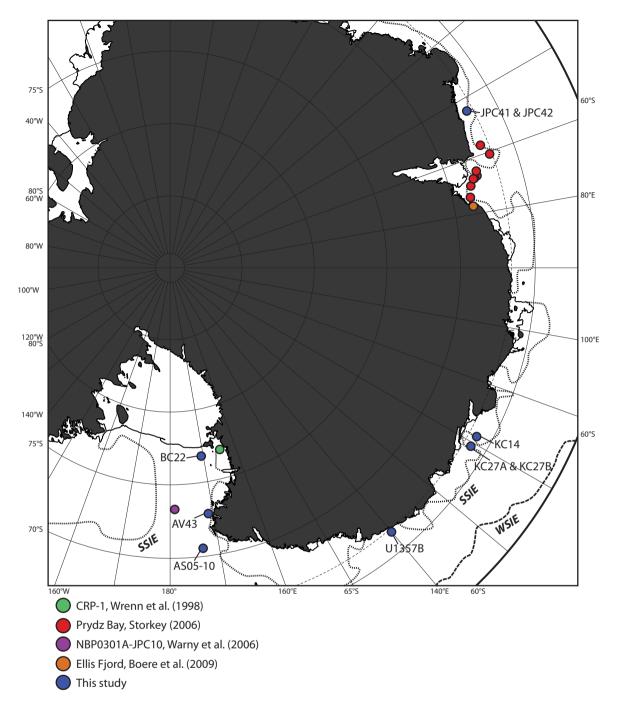


Figure 1. Previously published sites and the sites presented in this study from which samples are derived that contain *Nucicla umbiliphora*. The positions of the summer sea ice edge (SSIE) and winter sea ice edge (WSIE) are indicated by dotted and interrupted lines, respectively. The positions of the SSIE and WSIE are based on the figure by Arrigo et al. (2008), which shows the averaged satellite-derived, annual sea-ice cover for the period 1997–2006. SSIE: < 20 days/year sea-ice cover, and WSIE > 320 days/year sea-ice cover.

Table 1. Coordinates, core length and water depth of the sampled cores.

Core	Locality	Latitude	Longitude	Core length (m)	Water depth (mbsl)	Reference
AS05-10	Western Ross Sea	70°59.11′S	173°03.91′E	7.50	2377	This study
ANTA02-AV43	Western Ross Sea	74°08.45′S	166°04.97′E	2.20	218.5	Del Carlo et al. (2015)
BC22	Southwestern Ross Sea	76°41.59′S	169°04.68′E	0.37	790	This study
IODP U1357B	Adélie Basin	66°24.7990′S	140°25.5705′E	172.44	1017	Exp.318 Scientists (2011)
NBP0101-JPC41	MacRobertson Shelf	67°07.817′S	62°59.436′E	24.12	563	Leventer et al. (2001)
NBP0101-JPC42	MacRobertson Shelf	67°07.479′S	63°00.195′E	24.95	850	Leventer et al. (2001)
NBP1402-KC14	Sabrina Coast	66°52.3691′S	118°14.4022′E	2.63	643	Domack & Leventer (pers. comm.)
NBP1402-KC27A	Sabrina Coast	66°11.092′S	120°30.2403′E	2.952	544	Domack & Leventer (pers. comm.)
NBP1402-KC27B	Sabrina Coast	66°11.0907′S	120°30.2385′E	2.71	547	Domack & Leventer (pers. comm.)



multiple images, which is ideal for three-dimensional microscopic objects.

A scanning electron microscope (SEM) photograph was made using a JEOL NeoScope JCM-6000 Benchtop SEM, located at the 'Gemeenschappelijk Milieu Laboratorium' building at Utrecht University. For the SEM photo, dinoflagellate cysts were individually picked using a microinjection system, subsequently placed on a stub and coated with a thin (10 nm) layer of platinum.

Plate terminology follows the Kofoid tabulation system (see Kofoid 1911). Archaeopyle descriptive terms follow Bujak & Davies (1983).

#### 3. Results

Nucicla umbiliophora was found in core-top samples of all the examined cores, with the exception of cores AS05-10, NBP0101-JPC41 and -JPC42 for which no core-top samples were available. Dinoflagellate cysts of N. umbiliphora were occasionally found with the operculum still attached (Plate 1, figures 1, 2, 6) and in one case also containing cell contents (Plate 1, figure 10-11). The cysts of N. umbiliphora did not autofluoresce under fluorescence microscopy (Plate 1, figure 12).

For core ANTA02-AV43 all samples were taken above the interval 1.48-1.51 m below sea floor (mbsf), which has an age of 9.7  $\pm$  5.3 ka based on <sup>40</sup>Ar/<sup>39</sup>Ar dating (Del Carlo et al. 2015). No age models have been published yet for cores U1357B, BC22, NBP0101-JPC41 or NBP0101-JPC42. However, the latter two are likely of Holocene age as the nearby core NBP0101-JPC43B (23.95 m long) shows bottom <sup>14</sup>C ages of about 11.6 ka (Mackintosh et al. 2011). We encountered the species in JPC41 as deep as 17.52 mbsf. The new species has also been encountered throughout core BC22 (36.5 cm in length). Unpublished dinoflagellate cyst data from box core BC22 suggest a position for the Last Glacial Termination between 0.25 and 0.28 mbsf, from which the amount of dinoflagellate cysts per gram of dry sediment decreases strongly downcore. It has also been encountered sparsely in Hole U1357B, to a depth of 55.06 mbsf. The <sup>14</sup>C data from the nearby Hole U1357A provide an age of  $\sim$ 4.2 cal. kyr BP at 68.85 mbsf (Yamane et al. 2014). Samples from core AS05-10 have been retrieved from the interval with optimal dinoflagellate cyst preservation, which is associated with the onset of MIS5.5 (JD Hartman, pers. obs.). Apart from perhaps the CRP-1 core, for which the age model of the Quaternary section above 43 mbsf is not well resolved, the occurrence in core AS05-10 during MIS5.5 is the oldest record of Nucicla umbiliphora.

## 4. Systematic palaeontology

Division DINOFLAGELLATA (Bütschli 1885) Fensome et al. 1993

Subdivision DINOKARYOTA Fensome et al. 1993 Class DINOPHYCEAE Pascher 1914 Subclass PERIDINIPHYCIDAE Fensome et al. 1993 Order PERIDINIALES Haeckel 1894 Suborder PERIDINIINEAE Fensome et al. 1993 Family PROTOPERIDINIACEAE Bujak & Davies 1998 in Fensome et al. 1998

Subfamily PROTOPERIDINIOIDEAE Bujak & Davies 1983 Genus Nucicla gen. nov.

Type species. Nucicla umbiliphora Hartman, Sangiorgi, Bijl & Versteegh sp. nov.

**Derivation of the name.** From the Latin *nucicla*, meaning small nut, in reference to the cyst resembling a nut.

Diagnosis. Acavate, dorsoventrally compressed cyst with a rounded pentagonal outline, a hypocyst that is twice the size of the epicyst, an archaeopyle formed by the loss of three anterior intercalary plates, and a large sulcus with the posterior sulcal plate positioned at the antapex.

Differential diagnosis. This genus differs from all other peridinioid dinoflagellate cysts by its combination of (1) a consistent 31 archaeopyle, (2) a well-outlined cingulum and sulcus, (3) the absence of cavation, and (4) a large sulcus with a posterior sulcal plate at the antapex. The Late Cretaceous to Early Palaeocene genus Trithyrodinium Drugg 1967 also has a 31 archaeopyle, but is cavate. Vozzhennikovia Lentin & Williams 1976 has an I or 3I archaeopyle, and is also cavate. Although the number of archaeopyle plates in Brigantedinium Reid 1977 is not determined and therefore can include species with a 3I archaeopyle, Brigantedinium is spherical/ovoidal and lacks tabulation other than the archaeopyle. Other genera with dorsoventral compression, a pentagonal outline and consisting of an autophragm include Votadinium Reid 1977, Lejeunecysta Artzner & Dörhöfer 1978, Trinovantedinium Reid 1977 and Leipokatium Bradford 1975. Like Brigantedinium, Votadinium and Leipokatium can have an archaeopyle consisting of any number of intercalary plates, but Votadinium differs from Nucicla in having a shallow or deep depression between the antapical lobes, and lacks a well-defined cingulum. Leipokatium has very distinct antapical horns and a hypocyst much smaller than the epicyst. Both Lejeunecysta and Trinovantedinium may have a sulcus and/ or cingulum, but have an I archaeopyle. In addition, Trinovantedinium has non-tabular proximochorate processes and Lejeunecysta has an epicyst and hypocyst of approximately equal size. For all of the above-mentioned genera the position of the posterior sulcal plate is either unclear or not as posterior as in Nucicla.

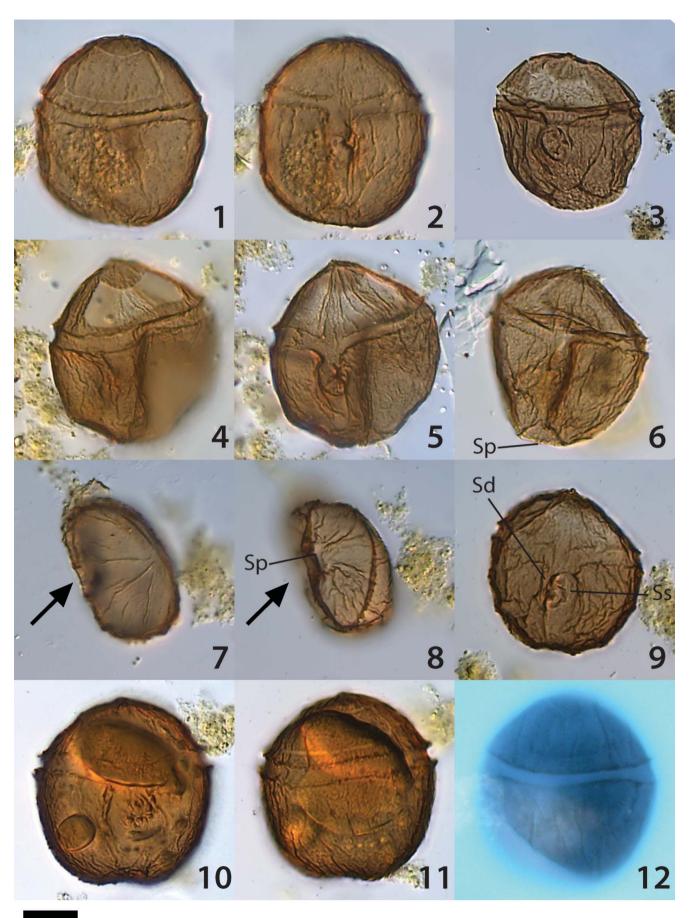
> Nucicla umbiliphora sp. nov. Plate 1, figures 1-12 and Plate 2, figures 1-6

#### Synononymy.

Dinocyst sp. A. Wrenn et al. 1998, p. 595, fig. 5 a-d. Protoperidinium sp. 2 Storkey, 2006, p. 49, plate 4, fig. 10-12. Lejeunecysta cf. sp. 1 and 5 of CRP Warny et al. 2006, p. 163, plate 3, figs 3-4.

Cyst type 1 Boere et al., 2009, p. 273, fig. 5D, E, (F?).

Holotype. Plate 1, figure 1-2. Cruise NBP0101, core JPC41, 108 cm depth, slide no. 1, England Finder (EF) coordinates: U29.2 down left corner.



20 μm



Repository. Stored in the collection of the Laboratory of Marine Palynology and Oceanography, Netherlands.

Type locality. Iceberg Alley, MacRobertson Shelf, East Antarctica.

Type stratum. Holocene.

Paratype. Plate 1, figure 6. Core ANTA02-AV43, 2-3 cm depth, slide no. 2, EF: F19.2 bottom side. Stored in the collection of the Laboratory of Marine Palynology and Oceanography, Utrecht, The Netherlands.

Stratigraphical range. Marine Isotope Stage 5.5 to Recent.

Derivation of the name. From the Latin umbilicus (navel) and the Ancient Greek suffix -phoros (bearing), with reference to its large flagellar scar, which resembles a navel.

**Diagnosis.** A species of *Nucicla* with a scabrate wall structure and with low, undulating or crenulating ridges with no apparent relation to plate boundaries with the exception of the cingulum and sulcus. The sulcus shows a large flagellar scar. Apart from the cingulum and sulcus, tabulation is only indicated by the clear 3I archaeopyle, of which the 2a intercalary is large and latideltaform. In the holotype the operculum is still attached; specimens are usually found without operculum.

#### Dimensions.

Holotype: Height = 70  $\mu$ m; width measured along the cingulum =  $62 \mu m$ .

Other specimens (n = 10): Height = 57-70  $\mu$ m, average = 64.5  $\mu$ m. Width = 52–63  $\mu$ m, average = 60.0  $\mu$ m.

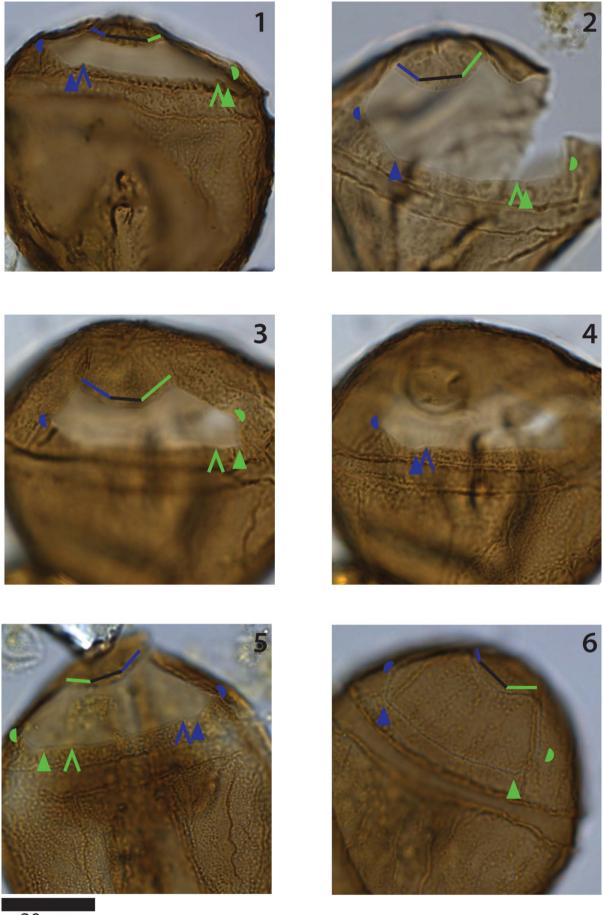
Description. A brown cyst with pentagonal outline, which is dorsoventrally compressed. Apical and antapical 'horns' are rounded and broad-based so that the cyst appears more rounded than pentagonal. A few specimens show a small acute apical horn (Plate 1, figures 3, 11). None of the specimens shows acute tips at the antapical 'horns'. The hypocyst is twice the height of the epicyst. The autophragm has folds that form low, undulating or crenulating ridges, which are predominantly longitudinal and can be dendritic. With the exception of the cingulum and sulcus, these ridges have no relation to sutures. The cyst wall is scabrate and the degree of scabration varies between specimens (Plate 2, figures 4-6). The clearly distinguishable cingulum is level or very slightly descending and

outlined by sutural ridges. Within the cingulum, low longitudinal ridges occur, but with no relation to cingular plate boundaries. The sulcus forms a clear depression, and is outlined by sinistral and dextral longitudinal sutural ridges. Some specimens also clearly show a posterior sutural ridge, which outlines the entire posterior sulcal plate (Sp; Plate 1, figures 6 and 8). The sutural ridge that outlines the Sp extends towards the dorsal side of the cyst, indicating the exceptional antapical position of the Sp. To our knowledge no other protoperidinioid cyst possesses an Sp that is positioned so far posteriorly, thereby pushing the antapical plates to the dorsal side. The sulcus shows a large flagellar scar, with low ridges converging towards it. Applying standard protoperidinioid tabulation N. umbiliphora, low ridges within the sulcus seem to outline the right sulcal plate (Sd) and the left sulcal plate (Ss) (Plate 1, figures 3 and 9). In some specimens (Plate 1, figures 3 and 9) a low ridge is present at the anterior margin of the left and right sulcal plates, indicating the position of the anterior sulcal plate (Sa) (Figure 2).

The 3I archaeopyle is relatively large and spans almost the entire dorsal side of the epicyst. It has a consistent shape with clear angles marking plate junctions, except for the boundaries between the intercalary plates, which are smooth and subtle. Nevertheless, both archaeopyle and operculum outline suggest a 3I archaeopyle, for the following reasons: (1) the upper margin of the archaeopyle is concave; although in dinoflagellate cyst species with a 2a archaeopyle such a concave upper margin could be the result of the inward folding of the apical plates, we are certain that this is not the case for N. umbiliphora, because the outline of the operculum is also concave (Plate 2, figure 6); (2) the upper margin of the archaeopyle is three-sided (particularly visible in Plate 2, figure 3), strongly suggesting that these are the three sides of the third apical plate (3') that border each of the anterior intercalaries (green, black and blue lines in Plate 2); and (3) in several instances the lower margin of the 2a intercalary appears slightly but nevertheless clearly elevated with respect to the lower margins of the 1a and 3a intercalaries (indicated by the ^ symbols in the figures of Plate 2). The position of the plate junctions in the lower margin of the archaeopyle suggests that the 1a and 3a intercalaries are very narrow, bordering a large latideltaform 2a intercalary. Although we cannot determine any further tabulation from the cyst of N. umbiliphora, the relatively low height of the epicyst in combination with the large 2a suggests that the height of the 3", 4", and 5" precingular plates is suppressed (Figure 2).

**Differential diagnosis.** This is currently the only species in the genus. Within the Protoperidiniaceae, Nucicla umbiliphora with

Plate 1. Light microscope photographs of Nucicla umbiliphora. Figure 1. Holotype, JPC42, 468 cm depth, slide 1, EF: U29.2, dorsal side up: dorsal view with operculum attached but archaeopyle outlined; 2. holotype, JPC42, 468 cm depth, slide 1, EF: U29.2, dorsal side up: ventral view (mirrored); 3. AS05-10, slice V-92, slide 1, EF: G24.1, dorsal side up: ventral view (mirrored), outline of archaeopyle visible; 4. JPC41, 108 cm depth, slide 1, EF: L35.4, dorsal side up: dorsal view; 5. JPC41, 108 cm depth, slide 1, EF: L35.4, dorsal side up: ventral view (mirrored) with clearly visible flagellar scar; 6. ANTA02-AV43, 2–3 cm depth, slide 2, EF: F19.2, ventral side up: ventral view, a low ridge indicates the position of the posterior sulcal plate; 7. JPC41, 556 cm depth, slide 1, EF: 042.1, antapical side up: apical view (mirrored), sulcus indicated by arrow; 8. JPC41, 556 cm depth, slide 1, EF: O42.1, antapical side up: antapical view, sulcus indicated by arrow, a low ridge can be distinguished at the posterior edge of the posterior sulcal plate; 9. JPC42, 295 cm depth, slide 2, EF: G32.4, ventral side up: ventral view, some of the low ridges within the sulcus may correspond to sulcal plate boundaries; 10. JPC42, 295 cm depth, slide 2, EF: N33.3, dorsal side up: ventral view (mirrored), cell contents visible; 11. JPC42, 295 cm depth, slide 2, EF: N33.3, dorsal side up: dorsal view, cell contents visible, operculum still attached, but outline visible; 12. Holotype, JPC42, 468 cm depth, slide 1, EF: U29.2, viewed under fluorescence microscope showing no autofluorescence.



20 μm

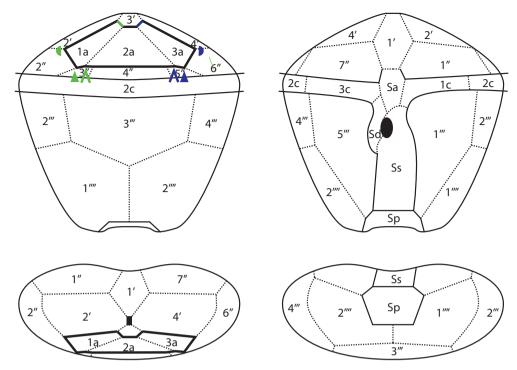


Figure 2. Schematic drawings of the plate boundaries of Nucicla umbiliphora. Top left: dorsal view. Top right: ventral view. Bottom left: apical view. Bottom right: antapical view. Uninterrupted lines indicate archaeopyle (thick), cingulum and sulcal outlines (thin). Dotted lines are hypothetical, based on the standard protoperidinioid plate configuration and deduced from the archaeopyle outline and presumed sutural sulcal ridges. Archaeopyle plate junctions are marked by symbols. The upper archaeopyle margin is indicated by coloured lines. This coding of lines and symbols corresponds to that used in Plate 2.

its pentagonal outline, limited tabulation, rounded antapical horns, scabration and brown colour most closely resembles Lejeunecysta rotunda Clowes et al. 2016. Instead of erecting a new genus one might consider emending Lejeunecysta to include N. umbiliphora. However, Lejeunecysta differs from Nucicla in more than just one aspect. Most importantly, Lejeunecysta has a consistent 2a archaeopyle and closely defined archaeopyle shape. Lejeunecysta also differs in having the epicyst and hypocyst of approximately equal length, symmetrically located horns which are small pointed and solid, a laevigate or chagrinate wall, and tabulation only indicated near the archaeopyle and cingulum, whereas the sulcus is only marked by a shallow depression. Because of the number and clarity of the differences, we found it necessary to erect a new genus. It differs from most other peridinioid genera by its 31 archaeopyle. Other known genera with a 3I archaeopyle are the Cretaceous to Early Palaeocene Trithyrodinium and Palaeocene to Oligocene Vozzhennikovia, but both these genera are cavate. Typically, the apical and antapical horns of these genera are made up of the periphragm, while N. umbiliphora only has an autophragm (Figure 3). Other brown scabrate dinoflagellate cyst species from the present-day Southern Ocean are Selenopemphix antarctica Marret & de Vernal 1997, Brigantedinium pynei Hannah

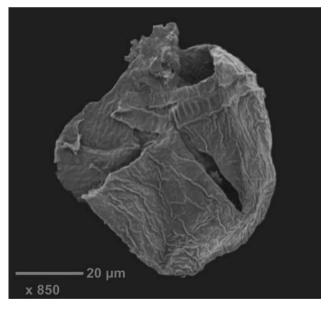


Figure 3. Benchtop scanning electron microscope (SEM) photograph of Nucicla umbiliphora (dorsal view). No tabulation pattern is visible except for the archaeopyle and cingulum. The autophragm with its folded ridges can be seen both externally and internally.

et al. 1998 and Cryodinium meridianum Esper & Zonneveld 2002. Cryodinium meridianum also has low, sometimes dendritic ridges, but most of these crests reflect tabulation. Furthermore, C. meridianum lacks the pentagonal outline and dorsoventral compression, and has a 21 archaeopyle. Brigantedinium pynei is reminiscent of C. meridianum, but its rugulose surface does not reflect tabulation and it has an I archaeopyle (Clowes et al. 2016). Selenopemphix antarctica does not have ridges, has an Itype archaeopyle, and the width of the cyst is much larger than its height, so that it typically appears in (ant)apical view on microscope slides. Nucicla umbiliphora was found in Quaternary sediments of the Ross Sea together with the similar-looking Dinocyst sp. B (Wrenn et al. 1998). Because Dinocyst sp. B of Wrenn et al. (1998) has antapical 'horns' with acute tips, and the number of intercalary plates that comprise the archaeopyle is uncertain, it is not included in N. umbiliphora.

#### 5. Discussion

#### 5.1. Taxonomy

We placed N. umbiliphora within the family Protoperidiniaceae based on the visible tabulation, the absence of cavation and the brown colour. It lacks plate boundaries between the cingular plates, which hampers definite placement within Protoperidiniaceae. However, several modern cysts with a pentagonal outline and an intercalary archaeopyle but without plate boundaries between the cingular plates do produce Protoperidinium thecae, such as Votadinium, Lejeunecysta, Selenopemphix and Trinovantedinium (Head 1996; Matsuoka & Head 2013; Mertens et al. 2017). In addition, N. umbiliphora does not show green autofluorescence, like many Protoperidinium cysts (Brenner & Biebow 2001; Anderson et al. 2003). Currently, the motile stage of N. umbiliphora is unknown.

### 5.2. Ecology

Nucicla umbiliphora occurrences are all near the Antarctic margin and, except for site AS05-10, only in sediments from the shelf or inland fjords. This strongly suggests it is endemic to the Antarctic shelf. All samples areas experience at least 9 months of yearly sea-ice cover (Figure 1; Arrigo et al. 2008). Considering that all modern dinoflagellates that produce brown cysts are heterotrophic, it is likely that the motile stage of N. umbiliphora is heterotrophic as well (Ellegaard et al. 2013). In the coastal waters of Antarctica, the phytoplankton blooms in the highly stratified surface waters after sea-ice retreat could be an important food source for N. umbiliphora (Kang & Fryxell 1993; Clarke & Leakey 1996; Arrigo et al. 1998; Smith Jr. et al. 2000; Hiscock et al. 2003; Smith Jr. et al. 2006; Peloguin & Smith Jr. 2007; Arrigo et al. 2008). At the Antarctic shelf, these conditions typically arise within the marginal ice zone in late summer (Fitch & Moore 2007; Arrigo et al. 2008) and within coastal polynyas (Arrigo et al. 1999; Arrigo & van Dijken 2003). Notably, the occurrence of N. umbiliphora in Hole U1357B confirms a preference for polynya environments, as U1357B was drilled directly downwind and downcurrent of the Mertz Glacier Polynya (Expedition 318 Scientists 2011).

#### 6. Conclusions

Nucicla umbiliphora gen. et sp. nov. (Peridiniales, Protoperidinioideae) occurs in Quaternary sediments from the East Antarctic margin and the Ross Sea. It has a 31 archaeopyle, which is unique among protoperidinoids. Furthermore, it has a rounded to pentagonal outline with a hypocyst twice as large as the epicyst, a sulcus with a distinct flagellar scar and a posterior sulcal plate positioned at the antapex. It has a scabrate wall ornamentation with low, somewhat crenulating ridges. The species is probably endemic to the Antarctic shelf environment during the Holocene and may prove to be confined to the high primary productivity after spring sea-ice retreat.

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#### **Disclosure statement**

No potential conflict of interest was reported by the authors.

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#### **Notes on contributors**



JULIAN HARTMAN started his PhD in the Marine Palynology and Palaeoceanography group, Utrecht University, The Netherlands, in January 2014 under the supervision of Henk Brinkhuis, Stefan Schouten, Francesca Sangiorgi and Peter Bijl. His PhD research focusses on reconstructing past surface-water conditions close to the Antarctic margin using palynology and organic geochemical proxies in order to reconstruct cryosphere

dynamics during periods of warming since the Oligocene. He is currently in the final stages of his PhD research.



FRANCESCA SANGIORGI earned a PhD in marine environmental sciences at the University of Bologna (Italy) and she is currently an assistant profes-Marine sor in the Palynology and Paleoceanography group, Department of Earth Sciences, Utrecht University. Her research mainly focuses on the ecology and palaeoecology of dinoflagellate cysts and on the Neogene climate evolution of polar oceans, where she combines

marine palynology with organic geochemical proxies. She works on several international projects dealing with the environmental and climatic evolution of the Southern Ocean.





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