

Iron and manganese shuttles control the formation of authigenic phosphorus minerals in the euxinic basins of the Baltic Sea

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ABSTRACT

Microanalysis of epoxy resin-embedded sediments is used to demonstrate the presence of authigenic iron (Fe) (II) phosphates and manganese (Mn)- calcium (Ca)- carbonate-phosphates in the deep euxinic basins of the Baltic Sea. These minerals constitute major burial phases of phosphorus (P) in this area, elevating the total P burial rate above that expected for a euxinic depositional environment. Particle shuttles of Fe and Mn oxides into the deep euxinic basins act as drivers for P-bearing mineral authigenesis. While Fe (II) phosphates are formed continuously in the upper sediments following the sulfidization of Fe-oxyhydroxides and release of associated P, Mn-Ca-carbonate-phosphates are formed intermittently following inflow events of oxygenated North Sea water into the deep basins. The mechanism of Fe (II) phosphate formation differs from previously reported occurrences of vivianite formation in marine sediments, by occurring within, rather than below, the sulfate-methane transition zone. The spatial distribution of both authigenic phases in Baltic sediments varies in accordance with the periodic expansion of anoxia on centennial to millennial timescales. The results highlight the potential importance of authigenic P-bearing minerals other than carbonate fluorapatite for total P burial in euxinic basins.

1. INTRODUCTION

Phosphorus (P) is the ultimate limiting nutrient for marine biological production, due to the ability of pelagic cyanobacteria to fix nitrogen (N) from the atmosphere and thus to make up any deficit in the supply of nitrate (NO_3^-) relative to phosphate (HPO_4^{2-}) in surface waters (Tyrrell, 1999). Hence, the availability of P in marine systems controls the rate of organic matter production. Phosphorus availability, in turn, is controlled by the balance between the input of P from rivers and the atmosphere, internal recycling and exchange with adjacent marine basins, and finally removal through burial in sediments (Ruttenberg, 2003).

The flux of P to the oceans can vary naturally on geological timescales (Tsandev and Slomp, 2009), while on the shorter timescale of recent decades, anthropogenic loading of both P and N has led to the eutrophication of coastal systems worldwide (e.g. Elmgren, 2001; Turner et al., 2006). By influencing organic matter production, P also exerts a strong control on oxygen concentrations in the oceans due to the oxygen demand of organic matter respiration. Moreover, positive feedbacks within the P cycle amplify this relationship. Under low-oxygen conditions, P is preferentially remineralized (relative to carbon and nitrogen) from sedimentary organic matter. This process stimulates efficient P recycling to surface waters, fuelling further production and a high biological oxygen demand (Ingall et al., 1993; Jilbert et al., 2011). The reduction of sedimentary iron (Fe)-oxyhydroxides under low-oxygen conditions also lowers the potential of sediments to 'trap' P close to the sediment-water interface (Mortimer, 1941), further increasing the efficiency of P regeneration.

Sediment P burial represents the long-term output of the marine P cycle, and thus has the potential to break the feedback loop between P regeneration, high biological production

and low-oxygen conditions. Phosphorus may be buried in sediments within various distinct fractions. These include organic matter, detrital phosphate minerals and authigenic mineral phases (Ruttenberg, 1992). Authigenic P-bearing phases buried in sediments include P associated with Fe-oxyhydroxides which resist reduction in the upper sediments (Reed et al., 2011), calcium (Ca)-phosphates such as carbonate fluorapatite (e.g. Ruttenberg and Berner, 1993) and P associated with manganese (Mn)-Ca-carbonates (Mort et al., 2010; Suess, 1979), and finally Fe (II) phosphates such as vivianite (e.g. Burns, 1997).

Carbonate fluorapatite (CFA), $\text{Ca}_{10}(\text{PO}_4)_{6-x}(\text{CO}_3)_x(\text{OH},\text{F})_{2+x}$, forms either by direct nucleation from saturated porewaters (Van Cappellen and Berner, 1989), via precursor phases (Gunnars et al., 2004; Jahnke et al., 1983) or by conversion of calcite microfossils within the sediments (Manheim et al., 1975). CFA formation has been shown to occur in a range of coastal and deep-sea environments worldwide (Filippelli and Delaney 1996; Ruttenberg and Berner 1993; Slomp et al., 1996) and is considered the most important authigenic sink for P in the marine realm, accounting for an estimated 50% of global P burial (Ruttenberg, 1993).

In contrast to Ca-phosphates, the formation of vivianite, $\text{Fe}_3(\text{PO}_4)_2 \cdot 8(\text{H}_2\text{O})$, in marine sediments has been reported only in a few locations (Burns, 1997; März et al., 2008; Ruttenberg and Goni, 1997; Schulz et al., 1994). Mostly, vivianite formation has been observed in environments where porewaters are rich in dissolved Fe, implying supersaturation with respect to vivianite. Porewater Fe accumulation is most likely to occur below the sulfate-methane transition zone (SMT) of marine sediments, where sulfide production is absent. In the presence of hydrogen sulfide, dissolved Fe is expected to be rapidly sequestered into the solid phase, maintaining undersaturation of the porewaters with respect to vivianite.

Despite the established knowledge of enhanced P regeneration from organic matter under low-oxygen conditions (Ingall et al., 1993), the processes controlling sequestration of P into authigenic minerals in low-oxygen environments are poorly constrained. Improved knowledge of these processes is required to better constrain the global rate of P burial in marine sediments, and consequently the relationships with the global carbon and oxygen cycles (Van Cappellen and Ingall, 1996). The modern expansion of low-oxygen conditions in the coastal zone (Diaz and Rosenberg, 2008) and the open ocean (Stramma et al., 2008) adds a particular urgency to this need. Recent work has suggested that sedimentary bacteria may influence Ca-phosphate authigenesis in some low-oxygen coastal environments (Goldhammer et al., 2010; Schulz and Schulz 2005), but the global importance of this process is still unknown. Moreover, both the mechanisms and global importance of Fe (II) phosphate precipitation in low-oxygen sediments remain to be established.

Here we present data from one of the world's largest anoxic water bodies, the Baltic Sea, which suggest that Fe (II) phosphates play an important role in sediment P burial in euxinic basins. We present direct evidence for currently-forming sedimentary Fe-phosphate minerals, obtained from microanalysis of epoxy resin-embedded sediments. Using porewater chemical data we show how the formation mechanism of Fe (II) phosphates differs from those reported in earlier studies, suggesting that Fe (II) phosphate authigenesis may be more widely distributed in low-oxygen marine sediments than previously thought. We also confirm the importance of Mn-Ca-carbonates as a sink for P in Baltic sediments and demonstrate the role of specific microfossils in sequestering P as apatite. Finally, by analyzing sediment cores capturing the past ~8000 years, we show how authigenesis of P-bearing minerals has varied in the Baltic throughout the Holocene.

2. STUDY SITES AND CORE COLLECTION

Sediment cores were recovered using a multi-corer and gravity corer from three sites in the central Baltic Sea during multiple sampling campaigns (Fig. 1). Details of the sites and the data collected during each sampling campaign are given in Table 1. This study focuses principally on two sites (F80 and LL19) located within the deep central basins of the Baltic. One shallower site (LF1) is studied as a reference for processes occurring on the margins of the deep basins.

The sediments at all sites have accumulated during the current brackish phase of the Holocene history of the Baltic, referred to as the Littorina Sea stage (e.g. Winterhalter et al., 1981). The Littorina was initiated between 9000 and 7000 yr BP, and its sediments are characterized by elevated organic carbon contents (Sohlenius et al., 2001). Three distinct phases of enhanced Baltic-wide anoxia have been identified in the Littorina sedimentary record, corresponding to the Early Holocene (~8000 yr BP to 4000 yr BP), the Medieval Climate Anomaly (MCA, ~1100 yr BP to 650 yr BP) and the modern anoxic interval from the late 20th century to present (Zillen et al., 2008).

The expansion of modern anoxia in the Baltic is principally driven by anthropogenic nutrient loading (Conley et al., 2009). Bottom waters in the deep basins today are frequently euxinic, with hydrogen sulfide concentrations rising to > 20 µmol/L between major inflow events of more saline and oxygenated North Sea water (Baltic Environmental Database, <http://nest.su.se/models/bed.htm>). Both deep basin sites (F80 and LL19) were subject to an inflow in 2003 but were euxinic at the time of sampling. Hence, by the scheme of Mort et al. (2010) and Jilbert et al. (2011), these sites classify as ‘Group 2’ (permanently hypoxic/anoxic)

while the shallow reference site LF1 classifies as ‘Group 1’ (oxic/seasonally hypoxic, Table 1).

3. ANALYTICAL METHODS

3.1 Epoxy embedding and microanalysis

Mini sub-cores from sediment multicores were embedded, vertically intact, with epoxy resin in preparation for elemental microanalysis. Sediment was pushed upwards from the multicore tubes into a series of shorter (7 cm-length) tubes of equivalent diameter, which were separated by inserting horizontal dividers. Mini sub-cores were then taken from the 7 cm-length multicore sections using cutoff 1 cm-diameter centrifuge tubes. The cutoff centrifuge tubes were inverted, inserted into the sediment and capped at the base with a hollow cap, after insertion of a piece of filter cloth between cap and tube. The mini sub-cores were then transferred to acetone baths in a polyethylene argon-filled glovebox and subsequently embedded with Spurr’s epoxy resin (Jilbert et al., 2008, note that no desalination step was performed in the present study). Horizontally-oriented samples from gravity core sections were also embedded after extraction from the open core surface with aluminium trays.

After curing, epoxy-embedded samples were opened perpendicular to the plane of sedimentation with a rock saw, and the exposed internal surface was polished. The samples were then mounted inside an EDAX Orbis Micro XRF Analyzer to construct elemental maps at a spatial resolution of 30 μm (Rh tube at 30kV, 500 μA , no filter, 300 ms dwell time, 30 μm capillary beam). Selected zones were analyzed at higher resolution by electron microprobe energy-dispersive spectroscopy (EDS) elemental mapping (JEOL JXA 8600

Superprobe with Noran Voyager automation, 15kV accelerating voltage, Si/Li detector, 1 μm beam). General electron microprobe maps were produced at 4 μm spatial resolution, and subsequent targeted (1 μm -resolution) maps were made of the regions around selected particles. Additionally, microfossils located in the epoxy embedded samples and isolated from dried sediment powders were analyzed by scanning electron microscopy (SEM), to produce micrographs and SEM-EDS spot measurements of interior mineral concretions.

3.2 Porewater sampling and analysis

Multicores (~0-60 cm) and gravity cores (~0-4 m) were taken for porewater analysis. Porewater was analyzed for methane (CH_4 , one parallel core) and major element and nutrient chemistry (a second parallel core). All cores were pre-drilled with 1.5 cm diameter holes at regular depth spacing, and taped prior to coring. Upon recovery, cutoff syringes were inserted through the holes to extract wet sediment samples.

For the CH_4 analyses, precisely 10 ml wet sediment was extracted from each hole and transferred immediately to a 65 ml glass bottle filled with saturated sodium chloride (NaCl) solution. This bottle was then closed with a rubber stopper and screwcap, and a headspace of 10 ml N_2 gas was inserted. Methane concentrations in the headspace of the 65 ml glass bottles were determined by injection of a subsample into a Thermo Finnigan Trace GC gas chromatograph (flame ionization detector, Restek Q-PLOT column of 30m length, 0.32mm internal diameter, oven temperature 25 $^\circ\text{C}$). Data were then back-calculated to the original porewater concentrations (porosity was calculated from water content data in the multicore samples and assumed constant in the gravity core).

For the major element and nutrient analysis, approximately 20 ml wet sediment was extracted from each hole. The cutoff syringe was capped with parafilm and transferred to a nitrogen-filled glovebox, where the contents were transferred to a centrifuge tube. Samples were centrifuged at 2500 g for 10-30 min and transferred back to the glovebox. Supernatant porewaters were then filtered through 0.45 μm disposable filters and collected for subsampling.

One supernatant porewater subsample was taken for analysis of hydrogen sulfide (based on pH measurements at the depth of porewater sulfide maxima, assumed to be present as HS^-). A 0.5 ml subsample was immediately transferred to 2 ml of 2% zinc (Zn)-acetate solution after return to the glovebox, to precipitate zinc sulfide (ZnS), and stored at 5 $^{\circ}\text{C}$. Hydrogen sulfide concentrations were estimated spectrophotometrically by complexation of the ZnS precipitate in an acidified solution of phenylenediamine and ferric chloride (Strickland and Parsons, 1972).

A second supernatant porewater subsample was taken for analysis of P, Fe, Mn, Ca and sulfur (S) by ICP-OES (acidified to 1M HCl, stored at 5 $^{\circ}\text{C}$). Hydrogen sulfide was assumed to be released during the initial acidification, thus S data is assumed to represent sulfate (SO_4^{2-}) only. Comparison between ICP-OES-derived P and autoanalyzer-derived HPO_4^{2-} concentrations suggest a contribution from dissolved organic phosphorus (DOP) at all sites of $< 20 \mu\text{mol/L}$ (not shown). Hence, ICP-OES-derived P is assumed approximately equal to phosphate. ICP-OES-derived Fe, Mn and Ca data are assumed to represent Fe^{2+} , Mn^{2+} and Ca^{2+} , respectively.

A third supernatant porewater subsample was taken for analysis of ammonium (NH_4^+) and stored at -20°C . Ammonium was determined spectrophotometrically on an autoanalyzer using the phenol hypochlorite method (Riley, 1953). A fourth subsample was titrated for total alkalinity with 0.01M HCl.

3.3 Bulk sediment sampling and analysis

Multicores from all sites were sliced onboard in a N_2 -filled glovebox immediately after recovery. To obtain gravity core sediment samples from site F80, sediment residue was taken from the centrifuge tubes used for porewater extraction. Gravity core sediment samples from site LL19 were taken from a separate core sliced in a glovebox (Table 1). Sediment samples from all cores were stored at -20°C before being freeze-dried (including an estimate of water content for non-centrifuged samples), powdered and ground in an agate mortar inside an argon-filled glovebox.

From each sediment sample, aliquots of 0.5 g dried sediment were decalcified by shaking in excess 1M HCl, initially for 12 h and for a further 4 h after addition of new acid. Tests have shown that the amount of organic carbon hydrolyzed by this protocol is negligible (van Santvoort et al., 2002). The decalcified sediment was dried, ground in an agate mortar and analyzed by combustion for organic carbon (C_{org}) using a Fisons NA 1500 NCS (precision and accuracy $<2\%$ based on an atropine/acetanilide standard calibration and checked against internal laboratory standard sediments).

A second 0.1 g aliquot of dried sediment was dissolved in 2.5 ml HF (40 %) and 2.5 ml of a $\text{HClO}_4/\text{HNO}_3$ mixture, in a closed Teflon bomb at 90°C for 12 h. The acids were then evaporated at 190°C and the resulting gel was dissolved in 1M HNO_3 , which was analyzed

for P, Fe and Mn concentrations by ICP-OES (precision and accuracy <5 %, based on calibration to standard solutions and checked against internal laboratory standard sediments).

A third 0.1 g aliquot of dried sediment was subjected to the SEDEX sequential extraction procedure to determine the solid-phase partitioning of phosphorus (Ruttenberg, 1992). All water (H₂O) rinses were omitted from the original protocol (see Slomp et al., 1996), but three magnesium chloride (MgCl₂) rinses were included. The full speciation is shown in Table 2 with the expected P phases extracted in each step indicated. The initial 1M MgCl₂ and citrate- dithionite- bicarbonate (CDB) rinses were performed inside the glovebox, to eliminate the potential conversion of CFA to Fe-oxyhydroxide- bound P due to pyrite oxidation upon oxygen exposure (Kraal et al., 2009). Phosphate in all rinses was analyzed colorimetrically by the ammonium heptamolybdate - ascorbic acid method (precision and accuracy <2 %, based on calibration to standard solutions and checked against internal laboratory standard sediments), with the exception of the CDB rinse, in which P was analyzed by ICP-OES.

4. RESULTS

4.1 Microanalysis

The electron microscope-EDS maps of the shallow sediments from site F80 show a heterogeneous P distribution (Fig. 2a). At the 4 µm resolution of the general map, most P enrichments are visible as single pixels or small clusters of pixels (Fig. 2a, b). Many of the brightest pixels in the P maps are also bright in the corresponding maps of Fe (Fig. 2b), indicating the presence of Fe-phosphates in these sediments. A 1 µm-resolution map centered on one such enrichment (Fig. 2c) suggests 2D particle dimensions in the range of ~10 µm,

although the morphology of the particles remains difficult to resolve. Some of the Fe-P enrichments also coincide with S (box 1 in Fig. 2a). Similar Fe-P enrichments are present throughout the deeper sections of the sediments at F80, but are only infrequently observed at LL19.

At specific depths at both F80 and LL19, layers of globular 100-500 μm -scale enrichments of P coincide with Ca and Mn. These enrichments are large enough to be resolved at the 30 μm resolution of the Micro XRF maps (Fig. 3), and indicate P associated with Mn-Ca-carbonates (hereafter *Ca-rhodochrosites*). Similarly to the Fe-P enrichments, these Ca-Mn-P enrichments are substantially more concentrated in the sediments at F80 than in those at LL19.

Many circular 50-200 μm -scale enrichments of Ca and P are present in the Micro XRF maps at all depths at both F80 and LL19, associated with microfossils in the sediments (Fig. 4a). The largest observed enrichments are related to unidentified spherical palynomorphs of up to 200 μm in diameter (Fig. 4b). SEM-EDS analysis of these specimens reveals concretions within the palynomorphs which are rich in Ca, P and oxygen (O), suggesting a form of apatite (Fig. 4b).

4.2 Porewater analysis

At site F80, a pronounced SMT is present in the upper 70 cm of the sediments, as indicated by the opposing gradients of SO_4^{2-} and CH_4 and a strong maximum in HS^- at 24 cm depth (Fig. 5a). The addition of HCO_3^- and HS^- to the porewaters at this depth also generates a small peak in the alkalinity profile, which otherwise shows the convex asymptotic increase with depth typical of anaerobic, organic rich sediments. At site LL19, the SMT is not present

in the depth interval sampled. Sulfate concentrations decline with increasing depth but are not fully exhausted within the analyzed 5 m, while CH₄ concentrations are negligible throughout the core and no distinct peak is observed in HS⁻ (Fig. 5b).

Porewater trends for most other components are similar between sites F80 and LL19. Ammonium concentrations display a convex asymptotic increase with depth. Phosphorus and Mn²⁺ concentrations increase with depth until 70 cm but decline thereafter. Iron (Fe²⁺) displays low but stable values at all depths, while Ca²⁺ increases slightly with depth (Fig. 5a, b). Maximum NH₄⁺, P and Mn²⁺ values are all higher at F80 than at LL19.

4.3 Multicore P speciation

The SEDEX extraction scheme was applied to multicore sediments from both deep-basin sites (F80 and LL19) and from the shallow reference site LF1. The P distributions at each of the three sites differ markedly from one another. Site F80 shows the highest total P concentrations, principally contributed by Org-P and CDB-P (Fig. 6a). Notably at this site, CDB-P shows a slight decline from the sediment surface downwards but never falls below 10 µmol/g after ~5 cm depth. Site LL19 shows intermediate total P concentrations, with enrichments of Org-P and CDB-P present in the upper 10 cm of the sediments (Fig. 6b). LF1 shows the lowest total P concentrations, but a clear exponential decay in CDB-P from high surface values to a background of 3-4 µmol/g (Fig. 6c). Acetate-P concentrations are generally constant with depth at all sites. The only exception is a single large peak in Acetate-P at 29 cm depth at site F80 (Fig. 6a).

4.4 Gravity core P speciation and bulk sediment chemistry

The depth intervals corresponding to known periods of enhanced Baltic-wide anoxia during the Littorina Sea stage are indicated by C_{org} enrichments in the sediments of F80 and LL19 (Fig. 7). Due to the shorter absolute core length and higher sedimentation rate at F80, the Early Holocene anoxic interval is not captured at this site.

Throughout the gravity core records, total P concentrations are higher at F80 than at LL19, due to higher concentrations of Org-P, CDB-P and, intermittently, Acetate-P (Fig. 7). The maximum Org-P enrichments at both sites occur during intervals of enhanced Baltic-wide anoxia. However, while small CDB-P and Acetate-P maxima also occur during Baltic-wide anoxic intervals at LL19, far stronger CDB-P and Acetate-P maxima occur during the intervening oxic periods at F80 (Fig. 7).

Concentrations of Ca and Mn are substantially higher at F80 than at LL19, indicating higher concentrations of Mn-Ca-carbonates in the sediments at the former site (Fig. 7). Furthermore, Ca and Mn concentrations increase with depth at F80, with occasional maxima superimposed onto this trend. Both the trend and occasional maxima coincide with those observed in the profile of Acetate-P.

5. DISCUSSION

Our results suggest that three principal P-bearing authigenic mineral phases are present in the sediments of the deep basins of the Baltic. The following sections will discuss the formation mechanisms of each phase in turn, before addressing the spatial and temporal context of P-bearing mineral authigenesis in the Baltic, and the broader implications for understanding the controls on P burial in marine sediments.

5.1 Fe (II) phosphate authigenesis

The CDB buffer solution of the SEDEX procedure is expected to dissolve both Fe-oxyhydroxide associated P (Ruttenberg 1992) and Fe (II) phosphates such as vivianite (Nembrini et al., 1983). Due to our strict anoxic procedures, contamination of the CDB-P signal by neoformation of Fe-oxyhydroxide associated P during sample handling can be ruled out (see Kraal et al., 2009). Furthermore, oxidation of primary Fe (II) phosphates to Fe (III) phases such as kertschenite, koninckite and santabarbaraite (e.g. März et al., 2008) should also have been avoided by our procedures. At site F80, CDB-P shows a slight decline from the sediment surface downwards but never falls below 10 $\mu\text{mol/g}$ after ~5 cm depth (Fig. 6, 7). This pattern suggests the presence of Fe oxyhydroxide-associated P in the surface sediments, of which a portion undergoes dissolution, while the remainder is converted to Fe (II) phosphates and buried (Fig. 6). The alternative hypothesis, namely deep preservation of Fe oxyhydroxide-associated P, appears unlikely in the strongly euxinic sediments of F80, due to the high reactivity of most ferric Fe minerals in the presence of hydrogen sulfide (Raiswell and Canfield, 2012).

Our microanalysis results support the interpretation of Fe (II) phosphate authigenesis in the modern sediments at F80. The electron microprobe EDS maps show 10- μm -scale enrichments of Fe and P at 5 cm depth (Fig. 2). We counted 10 discrete Fe-P enrichments in the 1 mm^2 map of Fig. 2a. The mean molar Fe/P of these enrichments, estimated from count ratios in their EDS spectra, is 1.65. Although the microprobe-EDS approach may be regarded as semi-quantitative only, this value appears significantly lower than those of Fe (III) oxyhydroxide-phosphate complexes found in the redoxcline of the Baltic (Fe/P = 2.0-2.9, Dellwig et al., 2010), and closer to that of Fe (II) phosphates such as vivianite,

$\text{Fe}_3(\text{PO}_4)_2 \cdot 8(\text{H}_2\text{O})$, $\text{Fe}/\text{P} = 1.5$. We propose that the enrichments represent Fe (II) phosphate minerals precipitated *in situ* in the sediments and therefore constitute a permanent P burial phase. A rough extrapolation from the observed frequency of enrichments, assuming spherical 10 μm -diameter particles of pure vivianite and accounting for porosity and sediment density, yields an expected vivianite-P concentration of $\sim 15 \mu\text{mol/g}$ at 5 cm depth. This value is similar to the 10 $\mu\text{mol/g}$ background CDB-P observed at this depth.

Several previous observations of marine vivianite formation have cited bulk porewater vivianite saturation due to a buildup of dissolved Fe and P below the SMT (e.g. März et al., 2008; Schulz et al., 1994). However, both our SEDEX and microanalysis results suggest Fe (II) phosphate formation in the uppermost 10 cm of the sediment at F80, thus well within the SMT at this location (Fig. 5a). Furthermore, the high sedimentation rate of the upper sediments ($\sim 0.5 - 1.1 \text{ cm/yr}$, estimated from ^{210}Pb data, not shown) confirms that Fe (II) phosphate authigenesis at this depth has occurred very recently, implying a presently-active process. Porewater conditions at F80 remained stable over three sampling years from 2009 to 2011 (Fig. 5a) and have likely been similar throughout the deposition of the upper 10 cm of sediment at this site. Assuming the measured bulk porewater conditions to be representative of those at the time of Fe (II) phosphate authigenesis, we calculated the saturation state of the porewater with respect to vivianite at 5 cm depth (using PHREEQC, and applying the saturation product of Al Borno and Tomson (1994), data from Fig. 5, and measured pH, temperature and salinity). The results suggest that the porewater is in fact undersaturated with respect to vivianite at this depth, principally due to the lack of free Fe (II) in the porewater. This implies that Fe (II) phosphate authigenesis is not controlled by the composition of the bulk porewater, but rather is directly coupled to the reduction of Fe-oxyhydroxides and release of associated P, potentially in protected microenvironments within the sediments.

Under the strongly euxinic conditions of F80, rapid reductive dissolution by HS^- (i.e. sulfidization) is likely to dominate over dissimilatory reduction (coupled to organic matter degradation) as the major pathway of Fe-oxyhydroxide reduction in the sediments. If sulfidization occurs in microenvironments, the probability of subsequent Fe (II) phosphate precipitation may be determined by the highly local availability of Fe, P and S. The precipitation of pyrite (FeS_2) is favored over that of vivianite in sulfidic systems across a range of Eh conditions and HPO_4^{2-} activities (Nriagu, 1972). Hence, in most cases of microenvironment sulfidization, pyrite would be expected to form as the dominant authigenic phase. However, under high enough localized HPO_4^{2-} concentrations, or after exhaustion of the local S supply relative to that of Fe and P, the precipitation of Fe (II) phosphates such as vivianite may be favored. Authigenesis of these minerals coupled to Fe-oxyhydroxide sulfidization would explain the occasional co-occurrence of Fe, P and S enrichments in the microprobe maps (e.g. box 1 in Fig. 2a). Intriguingly, Fe (II) phosphate accumulations were recently observed within cells of Deltaproteobacteria, the dominant sulfate-reducers of many SMT environments (Milucka et al., 2012). Further study is required to determine whether the Fe (II) phosphate accumulations observed in our samples are related to these microbial processes.

A strongly contrasting upper-sediment CDB-P profile is observed at the shallow reference site LF1, where CDB-P decays exponentially from the surface to background values of 3-4 $\mu\text{mol/g}$ (Fig. 6). Here, dissolution apparently removes most Fe and P from the CDB-soluble pool with increasing sediment depth, with negligible conversion to Fe (II) phosphate phases. Dissimilatory reduction of Fe-oxyhydroxides coupled to organic matter degradation is likely quantitatively more important than sulfidization in the low-sulfide sediments at LF1.

Indeed, the dominance of the former process was shown by a recent modeling study of a site in the south-west Baltic which experiences similar redox conditions and organic matter fluxes to those at LF1 (Reed et al., 2011). Hence, despite the plentiful supply of Fe-oxyhydroxides at such sites, substantial Fe (II) phosphate authigenesis does not occur, implying that a high rate of sulfidization is also a prerequisite for this process. The upper-sediment CDB-P profile of LL19 appears somewhat intermediate to those of the other two sites (Fig. 6), with a low background concentration from 10-50 cm depth, but a recent shift towards higher concentrations which coincides with an increase in Org-P. This pattern suggests that Fe (II) phosphate autogenesis has recently been initiated at LL19, in response to the onset of the modern anoxic interval (see also Fig. 7).

Although vivianite is the principal Fe (II) phosphate mineral to have been reported in marine sediments, it should be noted that the series $\text{Fe}^{2+}_3(\text{PO}_4)_2 \cdot (\text{H}_2\text{O})_n$ has multiple stable phases, such as ludlamite $\text{Fe}_3(\text{PO}_4)_2 \cdot 4(\text{H}_2\text{O})$ and phosphoferrite $\text{Fe}_3(\text{PO}_4)_2 \cdot 3(\text{H}_2\text{O})$ (Moore, 1971). A series of hydroxylated Fe (II) phosphates also exists, including tinctite $\text{Fe}_3(\text{PO}_4)_2(\text{OH})_3 \cdot 3(\text{H}_2\text{O})$ and lipscombite $\text{Fe}_3(\text{PO}_4)_2(\text{OH})_2$ (Nriagu and Dell, 1974). The analytical approaches in the present study are unable to differentiate between these phases.

5.2 Ca-rhodochrosite-P authigenesis

Intermittent layers rich in 100-500 μm -scale globular Ca-Mn-P enrichments are observed in the micro XRF maps of the sediments at site F80 (Fig. 3), and to a lesser extent at site LL19. The gravity core data for F80 show that extreme maxima in solid-phase Mn ($\leq 14\%$) and Ca ($\leq 3\%$) consistently coincide with maxima in Acetate-P (Fig. 7), confirming the association of P with Ca-rhodochrosite at this site. Ca-rhodochrosite has been reported in many previous studies of Baltic Sea sediments (e.g. Jakobsen and Postma, 1989; Manheim,

1982; Suess, 1979), and is believed to form when Mn oxide (MnO_x) packets emplaced at the sediment-water interface during inflow events are converted to carbonates after the re-establishment of anoxic, high-alkalinity conditions (Heiser et al., 2001; Huckriede and Meischner, 1996; Sternbeck and Sohlenius, 1997). Prior to these inflow events, bottom waters are rich in both dissolved Mn^{2+} (Neretin et al., 2003) and HPO_4^{2-} (Gustafsson and Stigebrandt, 2007). The association of P with particulate Mn thus most likely occurs during inflow events, upon the initial massive precipitation of MnO_x . This is supported by recent observations of particulate MnO_x -P in the vicinity of the modern redoxcline (Dellwig et al., 2010). During the subsequent conversion of MnO_x to Ca-rhodochrosite, phosphate apparently substitutes for carbonate in the rhodochrosite structure, generating mixed Mn-Ca-carbonate-phosphate phases as reported by Suess (1979).

Background sedimentary concentrations of Mn and Ca also increase downcore at site F80 (Fig. 7a), implying syn-sedimentary precipitation of Ca-rhodochrosite as suggested previously for Baltic sediments by Carman and Rahm (1997) and Heiser et al. (2001). Background Acetate-P shows a slight concomitant increase (Fig. 7a), and porewater concentrations of both Mn and P decrease from ~50 cm to the base of the core (Fig. 5a). These observations suggest that some P is also incorporated into the syn-sedimentary precipitated rhodochrosite phase. We suspect that emplaced Ca-rhodochrosite layers undergo stepwise recrystallization after burial, allowing Mn and P to accumulate in the upper porewaters and be drawn down at greater depth. This implies that the Ca-rhodochrosites are only metastable at the time of their initial crystallization, as previously noted by various authors (e.g. Jakobsen and Postma 1989; Sternbeck and Sohlenius 1997) who showed that typical Baltic rhodochrosite Mn/Ca ratios fall within the large solid-solution miscibility gap in the system CaCO_3 - MnCO_3 - H_2O (Middelburg et al., 1987).

5.3 Apatite precipitation in microfossils

Our microanalysis results show the frequent presence of apatite concretions within spherical palynomorphs and other microfossils at both F80 and LL19 (Fig. 4). The concretions in the palynomorphs have a soft texture and can be easily cut with a knife, suggesting a poorly crystalline material possibly aggregated with organic matter. The SEM-EDS spectra of the concretions often show small peaks of fluorine (F) beside those of C and O (Fig. 4b), suggesting the apatite phase present to be CFA. However, since the measured C and O may derive from either carbonate in CFA, or from organic matter, deducing the precise stoichiometry of the apatite phase is not possible from these results. Sodium (Na), Mn and magnesium (Mg) are also detected in minor quantities, implying substitution of these elements for Ca in the apatite structure. Extrapolations to estimate the contribution of these concretions to total Acetate-P in the SEDEX extractions are sensitive to both the assumed concentration of CFA within the poorly crystalline material, and the assumed CFA stoichiometry. However, if CFA with the formula of $\text{Ca}_{10}(\text{PO}_4)_{4.6}(\text{CO}_3)_{1.32}\text{F}_{1.87}(\text{OH})_{1.45}$ (Perrone et al., 2002) were to occupy 30% of the volume of all such palynomorphs in their observed density in the sediments, this phase alone could account for the full ~10-15 $\mu\text{mol/g}$ background Acetate-P observed at both F80 and LL19 (Fig. 6, 7), implying a quantitatively significant burial flux of P. Except for the effects of Ca-rhodochrosite-P precipitation and recrystallization at F80 (described in Section 5.2), sedimentary Acetate-P remains roughly constant with depth at both sites. This suggests that precipitation of CFA within biogenic microenvironments occurs not in the sediments, as previously reported for conversion from calcite (Manheim et al., 1975), but rather in the water column or at the sediment-water interface. Furthermore, this burial flux of P has varied little throughout the Holocene.

Precipitation of CFA in seawater is inhibited by the presence of Mg^{2+} ions (Martens and Harriss, 1970) and undersaturation with respect to precursor phases such as octacalcium phosphate $Ca_8H(PO_4)_3 \cdot 2.5(H_2O)$, which may have higher saturation products than CFA itself (Gunnars et al., 2004). Hence, the biogenic microenvironments in our samples must have experienced significant localized enrichment of Ca^{2+} and/or HPO_4^{2-} to trigger the precipitation of CFA. We speculate that degradation of organic material within confined microenvironments may have led to enhanced localized HPO_4^{2-} concentrations and hence supersaturation with respect to octacalcium phosphate. Notably, we did not observe P enrichments within microenvironments associated with biogenic silica, suggesting that diatom-related polyphosphate accumulations (Diaz et al., 2008) are of minor importance to P burial in the Baltic. However, without suspended matter samples we cannot comment on the possible role of polyphosphates as precursors to the apatite precipitation we observe, as reported for the precipitation of hydroxyapatite in *Thiomargarita* bacteria (Schulz and Schulz, 2005). Interestingly, Acetate-P was not observed to be an important fraction of particulate P in the water column of a silled fjord in another recent study (Diaz et al., 2012). This suggests either that the apatite precipitation we observe takes place mainly at the sediment-water interface, or that the palynomorphs and other microfossils in our samples – and hence the available microenvironments for CFA precipitation – are specific to the Baltic.

5.4 Bathymetric controls on the distribution of authigenic P-bearing minerals

Both Fe (II) phosphates (indicated by CDB-P) and Ca-rhodochrosite-P (indicated by peaks in Acetate-P) are more prevalent in the sediments at the deepest studied site, F80, than at the slightly shallower LL19 (Fig. 6, 7). Due to its greater water depth (Table 1), F80 experiences the more severe and persistent bottom water euxinia of the two sites (Baltic Environmental Database, <http://nest.su.se/models/bed.htm>). This spatial variability in redox

conditions ultimately controls the distribution of authigenic P-bearing minerals in the sediments of the Baltic, with Fe (II) phosphates and Ca-rhodochrosite-P found primarily in the deepest basins. Continuous shelf to basin ‘shuttling’ of the oxides of Fe and Mn has been shown to focus these elements preferentially into the deepest parts of euxinic basins (Lyons and Severman, 2006). In the Baltic, the spatial scale is apparently small enough, and the downslope Fe-oxyhydroxide flux great enough, that deep basin sites such as F80 can accumulate a surface-sediment pool of Fe-oxyhydroxides which have resisted dissolution in the water column. This pool carries with it associated P, and under the euxinic conditions in the deep basins, sulfidization triggers its partial conversion to Fe (II) phosphates (Section 5.1). In contrast, MnO_x largely dissolves upon entering the euxinic basins, leading to accumulation of Mn in the deep waters and intermittent massive reprecipitation of Mn oxides during inflow events (Section 5.2).

The variable distribution of redox conditions in the Baltic during the Holocene has also impacted the distribution of authigenic P-bearing minerals in the sediments on centennial to millennial timescales. When comparing the gravity core records of F80 and LL19 (Fig. 7), it can be seen that the maximum accumulation of CDB-P and Acetate-P at F80 occurred during periods of oxic conditions throughout much of the Baltic. In contrast, slightly enhanced CDB-P and Acetate-P burial at LL19 occurred during the Baltic-wide anoxic intervals of the Early Holocene, MCA and late 20th century (the slight increase of CDB-P burial at LL19 during the late 20th century is also visible in the upper 10 cm of the multicore from this site, Fig. 6). The contrasting histories of authigenic P-bearing mineral accumulation at the two sites can be explained in terms of the source area to basin sink area (S/B) concept used by Raiswell and Anderson (2005) to explain variable rates of Fe enrichment in anoxic basins. During generally oxic intervals of Baltic history, deep basin sites such as F80

apparently still experienced localized anoxia, due to their relatively restricted hydrography and the oxygen demand of focused organic material. These ‘pockets’ of low-oxygen conditions must have undergone massive focusing of Fe and Mn oxides - precursors to the eventual authigenic P-bearing minerals - due to the large source area of the surrounding sediments relative to the sink area of the basins themselves (i.e. high S/B conditions). In contrast, during periods of expanded anoxia such as the MCA, the source area of Fe and Mn oxides was relatively small, while the potential basin sink area was much greater (Fig. 8). These ‘low S/B’ conditions led to a more homogeneous accumulation of authigenic P-bearing minerals across a much wider area, including at site LL19.

5.5 Quantitative importance of authigenic P-bearing minerals to P burial in the Baltic

The complete suite of authigenic P-bearing minerals, including Fe (II) phosphates, Ca-rhodochrosite-P and microfossil CFA, account for a substantial fraction of total P burial in the deep basins of the Baltic. At F80, the authigenic contribution to total P during a typical ‘oxic’ interval - such as the period between the MCA and modern anoxic intervals - is 50% (Table 3). The net Baltic-wide burial rate of authigenic P-bearing minerals during oxic intervals may be roughly estimated by assuming that F80 is representative for all sediments below ~175 m water depth, equivalent to a total sedimentary area of ~5000 km². A mean accumulation rate of approximately 1760 tonnes P/yr into authigenic minerals is thus calculated for oxic intervals (Table 3), while total P burial in the Baltic is estimated at around 14000 tonnes P/yr (Bo Gustafsson, personal communication). This confirms that the authigenic component is a quantitatively significant burial pathway for P during oxic intervals. Hence, the shuttling of Fe and Mn oxide precursor phases into the deep basins may have helped to maintain generally oxic conditions between the MCA and modern anoxic interval, by efficiently sequestering P in a few euxinic deep basins.

During Baltic-wide anoxic intervals such as today, the spatial extent of authigenic P-bearing mineral formation increases to include site LL19 and likely all sediments below the redoxcline at ~100 m water depth (Fig. 8). However, the concentrations of Fe (II) phosphates and Ca-rhodochrosite-P in the sediments at F80 are lower at such times because of the low S/B conditions (Section 5.4). Due to the less focused occurrence of P-bearing mineral authigenesis, and the higher rate of Org-P burial, we calculate an authigenic contribution of just 21% to total P burial during the MCA at F80 (Table 3). Yet, if F80 is considered representative for 60000 km² of sediments during anoxic intervals, the net burial rate of P into authigenic minerals throughout the Baltic is 2540 tonnes P/yr (vs. 1760 tonnes P/yr in oxic intervals). Due to the uncertainties involved, the most important conclusion of this exercise is that the burial rate of authigenic P-bearing minerals is of a similar order of magnitude during both oxic and anoxic intervals, but that its contribution to total P burial declines during anoxic intervals due to enhanced burial of Org-P. Nevertheless, the significant burial of authigenic P-bearing minerals we report during both oxic and anoxic intervals contrasts with a previous study which suggested P burial in authigenic phases to be limited in the Baltic (Mort et al., 2010). We suggest that the small number of true deep-basin sites investigated in that study obscured the importance of authigenic phases to total P burial.

5.6 Wider implications for global sedimentary P burial

Major uncertainties still exist regarding the controls on P burial rates in marine sediments, principally related to poor understanding of the processes which regulate the sequestration of P into authigenic mineral phases. It is well established that low-oxygen conditions increase the efficiency of P release during organic matter breakdown, as recorded by higher sedimentary organic carbon to organic phosphorus ($C_{org}:P_{org}$) ratios under low-

oxygen conditions (Ingall et al., 1993; Jilbert et al., 2011; Slomp and Van Cappellen, 2007). Furthermore, a similar negative correlation exists between bottom-water oxygen concentrations and sedimentary organic carbon to total phosphorus ($C_{org}:P_{tot}$) ratios (Algeo and Ingall, 2007), suggesting that P released from organic matter in low-oxygen environments is not generally retained in the sediments. However, many of the datasets presented in the compilation of Algeo and Ingall (2007) contain a large degree of scatter along the $C_{org}:P_{tot}$ axis, implying that P mineral authigenesis rates vary between environments of similar redox conditions, exerting a secondary control on the total burial rate of P.

Authigenic apatite phases such as CFA are commonly found in ocean margin sediments (Filippelli and Delaney 1996; Rittenberg and Berner 1993; Slomp et al., 1996), and in some environments may act as a precursor to phosphorite deposits (Föllmi, 1996). Apatite formation has been suggested to be accelerated by sulfur bacteria such as *Thiomargarita* and *Beggiatoa* (Schulz and Schulz, 2005; Goldhammer et al., 2010), but the global distribution of these genera is not yet known. Potentially, sedimentary environments of similar redox conditions but contrasting bacterial assemblages may thus record different rates of CFA formation, accounting for some of the scatter in the global $C_{org}:P_{tot}$ data of Algeo and Ingall (2007). Alternatively, the bacteria may be abundantly present but the environmental conditions in the sediment may not support CFA formation. Our results for the Baltic show that the rate of Fe (II) phosphate and Ca-rhodochrosite-P authigenesis varies spatially and temporally in connection with the shuttling of precursor Fe and Mn oxide phases into euxinic basins. Total P burial in euxinic basins may thus depend upon the magnitude and distribution of these particle shuttles. Although the flux of Fe and Mn oxides from surrounding continents is the primary driver of submarine particle shuttles, the spatial distribution of low-oxygen conditions in such basins modifies their magnitude and trajectory, and hence the burial rate of

P. For example at our deepest studied site, F80, localized euxinic conditions generally favor intense P mineral authigenesis, raising the rate of total P burial relative to that expected under the euxinic bottom-water conditions. However, the rate of authigenesis also decreases markedly at this site during intervals of more widespread anoxia, despite the constancy of euxinia in the deep basins themselves. Hence, site-specific redox conditions alone cannot be used to predict the total rate of P burial without an understanding of their wider spatial context.

6. CONCLUSIONS

Iron (II) phosphate authigenesis occurs in the sediments of the deep euxinic basins of the Baltic Sea. In contrast to previously reported occurrences of Fe (II) phosphates in marine sediments, authigenesis of these minerals in the Baltic is not driven by bulk porewater saturation below the SMT. Rather, authigenesis occurs within the SMT in the upper sediments, most likely during the rapid reductive dissolution of Fe-oxyhydroxides and release of associated P by reaction with hydrogen sulfide. Phosphorus is also incorporated into Mn-Ca-carbonates, which form after inflow events into the deep euxinic basins have precipitated massive deposits of Mn oxides in the surface sediments. Throughout the Holocene, the burial rate of P burial in these two authigenic phases has been greatest in the deepest basins, where euxinia has persisted even during intervals of generally oxic conditions elsewhere in the Baltic. Such conditions favor intense shuttling of Fe and Mn oxide precursor phases into a restricted depositional area. A third authigenic P-bearing mineral, CFA, precipitates within biogenic microenvironments in the water column or at the sediment-water interface and contributes a relatively constant burial flux of P throughout the Baltic and throughout the Holocene. Our observations suggest that total P burial in euxinic basins may be strongly influenced by the distribution of submarine particle shuttles, partly accounting for the

observed scatter in predicted relationships between P burial rate and bottom-water redox conditions worldwide.

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TABLES

Table 1. Sampling sites in the Baltic Sea presented in this study.

Site name and classification	Cruise	Co-ordinates (deg.-dec.)	Water depth (m)	Multicore analyses	Gravity core analyses
F80 Permanently hypoxic/anoxic (Group 2*)	R/V Aranda May/June 2009	19.8968 E 58.0000 N	191	-Sediment	
	R/V Heincke June/July 2010	19.8899 E 57.9928 N	181	-Porewater	-Porewater -Sediment
	R/V Pelagia May 2011	19.8969 E 57.9997 N	194	-Epoxy embedding and microanalysis	
LL19 Permanently hypoxic/anoxic (Group 2*)	R/V Aranda May/June 2009	20.3108 E 58.8807 N	169	-Sediment	-Sediment -Epoxy embedding and microanalysis
	R/V Heincke June/July 2010	20.3112 E 58.8762 N	173	-Porewater	-Porewater
	R/V Pelagia May 2011	20.3122 E 58.8771 N	175	-Epoxy embedding and microanalysis	
LF1 Oxic/seasonally hypoxic (Group 1*)	R/V Aranda May/June 2009	21.2807 E 57.9825 N	67	-Sediment	

* Classifications from Mort et al. (2010) and Jilbert et al. (2011).

Table 2. Stages of the SEDEX sequential extraction procedure (Ruttenberg, 1992). Expected extracted phosphorus phases are listed, with references indicated when these differ from Ruttenberg (1992).

Stage and code	Extractant	pH	Rinsing time (h)	Expected P phases	Reference
1 Ex-P	1M MgCl ₂	8	0.5	-Exchangeable, loosely sorbed P	
2 CDB-P	Citrate-dithionite-bicarbonate (CDB)	7.5	8	-Fe oxyhydroxide-bound P -Vivianite	a
3 (CDB-P)	1M MgCl ₂	8	0.5	-Any P resorbed during stage 2	
4 Acetate-P	Na acetate buffer	4	6	-Carbonate fluorapatite (CFA) -Hydroxyapatite -Ca-rhodochrosite-P	b
5 (Acetate-P)	1M MgCl ₂	8	0.5	-Any P resorbed during stage 4	
6 Detrital-P	1M HCl	0	24	-Detrital apatite minerals	
7	(ashing at 550 °C for 2 h)				
8 Organic P	1M HCl	0	24	-Organic P	

^a Nembrini et al., 1983

^b Suess, 1979

Table 3. Estimated burial rates of authigenic P ([CDB-P + Acetate-P] – [0.5*surface sediment Acetate-P]) during the Medieval Climate Anomaly (MCA) and the period between the MCA and the modern anoxic interval. Estimates include Fe (II) phosphates, Ca-rhodochrosite-P and microfossil CFA (subtraction of half the surface-sediment Acetate-P concentration is based on an assumed 50% contribution of microfossil-CFA to total Acetate-P).

Time interval	Authigenic P burial rate at F80 ($\mu\text{mol}/\text{cm}^2/\text{yr}$)	Total P burial rate at F80 ($\mu\text{mol}/\text{cm}^2/\text{yr}$)	Contribution of authigenic P to total P at F80 (%)	Area of similar sedimentation (km^2)	Baltic-wide authigenic P burial rate (tonnes/yr)
End of MCA to onset of modern anoxic interval	1.14	2.27	50	5000	1760
MCA	0.14	0.65	21	60000	2540

FIGURE CAPTIONS

Figure 1. Map of the Baltic Proper, showing the principal sub-basins and the locations of the sites presented in this study. Bathymetric and coastline data are presented in Miller cylindrical projection, taken from the General Bathymetric Chart of the Oceans (GEBCO) Digital Atlas (Intergovernmental Oceanographic Commission et al., 2003).

Figure 2. (a) General electron microprobe-EDS maps of Fe, S and P on the internal surface of an epoxy-embedded sediment block from F80 (5 cm depth). The maps are presented in true vertical orientation and comprise a matrix of individual data points at 4 μm resolution. (b) Zooms of the numbered boxes indicated in the Fe and P maps in *a*. The arrows indicate examples of coincident enrichments of Fe and P. (c) Targeted electron microprobe-EDS maps (at 1 μm resolution) in the vicinity of one Fe-and P-enriched particle located in *a*. The particle is indicated by the arrow. A spot-measurement electron microprobe-EDS spectrum on the same particle is shown to the right. Color scaling in all microprobe-EDS maps is determined by the count intensity range within the mapped area, and subsequent adjustment of brightness and contrast to optimize the visualization of features.

Figure 3. Micro XRF maps of Ca, P and Mn on the internal surface of an epoxy resin-embedded sediment block from F80 (50 cm depth). The maps are presented in true vertical orientation and comprise a matrix of individual data points at 30 μm resolution. The histograms below each map indicate the distribution of pixels (data points) over the full range of measured count intensities. The vertical axes have been expanded and truncated to highlight the long-tailed distribution of the histograms; in reality, the modal peak for each

element extends several orders of magnitude beyond the depicted range. For each element, the majority of the pixels thus record relatively low count intensities, while the majority of the count range is represented by single pixels within ‘hotspots’ of extreme enrichment. Color scaling in the maps is determined by the count intensity range within the mapped area, and subsequent adjustment of brightness and contrast to optimize the visualization of features.

Figure 4. (a). Micro XRF maps of Ca and P on the internal surface of an epoxy resin-embedded sediment block from LL19 (325 cm depth). The maps are presented in true vertical orientation and comprise a matrix of individual data points at 30 μm resolution. The histograms below each map indicate the distribution of pixels (data points) over the full range of measured count intensities. The vertical axes have been expanded and truncated to highlight the long-tailed distribution of the histograms; in reality, the modal peak for both elements extends several orders of magnitude beyond the depicted range. For both elements, the majority of the pixels thus record relatively low count intensities, while the majority of the count range is represented by single pixels within ‘hotspots’ of extreme enrichment. Color scaling in the maps is determined by the count intensity range within the mapped area, and subsequent adjustment of brightness and contrast to optimize the visualization of features.

(b). SEM micrographs of spherical palynomorphs in epoxy-embedded sediments from F80, ~2.5cm depth (left) and isolated from dried sediment samples from LL19, ~300 cm depth (right). A spot-measurement SEM-EDS spectrum performed on the material within the palynomorph in the right-hand micrograph is shown below. The exact position of the measurement is indicated by the asterisk.

Figure 5. Porewater data from sites F80 and LL19. Multicore sulfate profiles from three consecutive years (left) are shown to illustrate the lack of spatial and temporal variability

between the slightly offset sampling locations of each year (see Table 1). In the complete profiles (right), filled circles indicate multicore samples, open circles indicate gravity core samples.

Figure 6. SEDEX phosphorus speciation results for multicore sediments from F80, LL19 and LF1. The stacked lines represent the concentrations of P extracted in each step of the SEDEX procedure, while the white-filled circles represent total P as determined by ICP-OES.

Figure 7. Complete solid-phase sediment profiles from F80 and LL19. In F80 data, filled circles indicate multicore samples, open circles indicate gravity core samples. LL19 data are higher resolution and presented in line form only. Grey bars indicate C_{org} excursions associated with widespread anoxic conditions in the Baltic during the Early Holocene (EH), Medieval Climate Anomaly (MCA), and modern (Mod.) intervals.

Figure 8. Proposed bathymetric contours corresponding to the redoxcline during generally oxic intervals of the Holocene history of the Baltic (left, 175 m) and during intervals of widespread anoxia such as the MCA and today (right, 100 m). Sediments shallower than the redoxcline act as a source for Fe and Mn oxide particle shuttles, while sediments deeper than the redoxcline act as a sink. The rate of particle shuttling to a given basin controls the formation of authigenic P-bearing minerals (see Section 5.4). Note the higher source area to basin sink area (S/B) ratio during generally oxic intervals.

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Figure 1.pdf

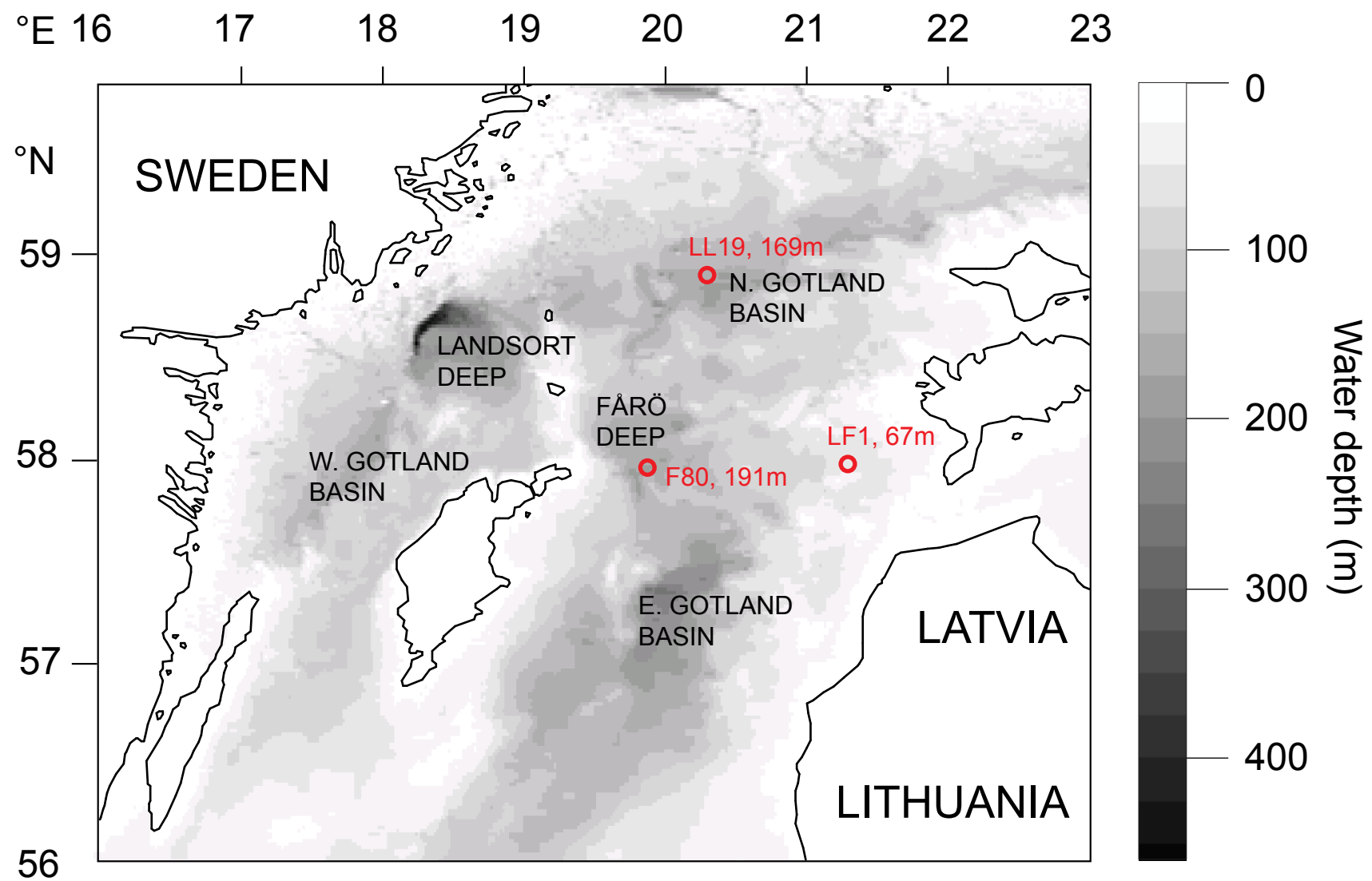


Figure 2.pdf

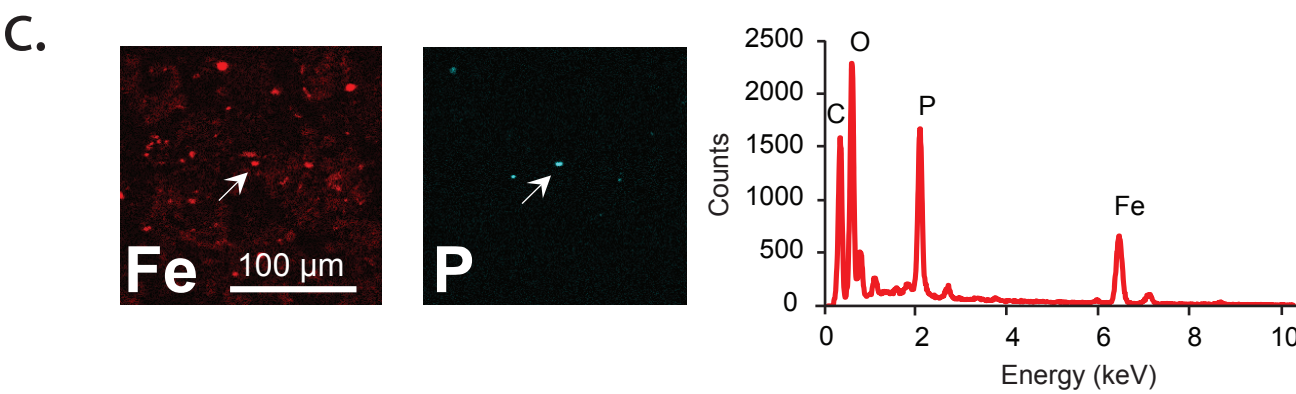
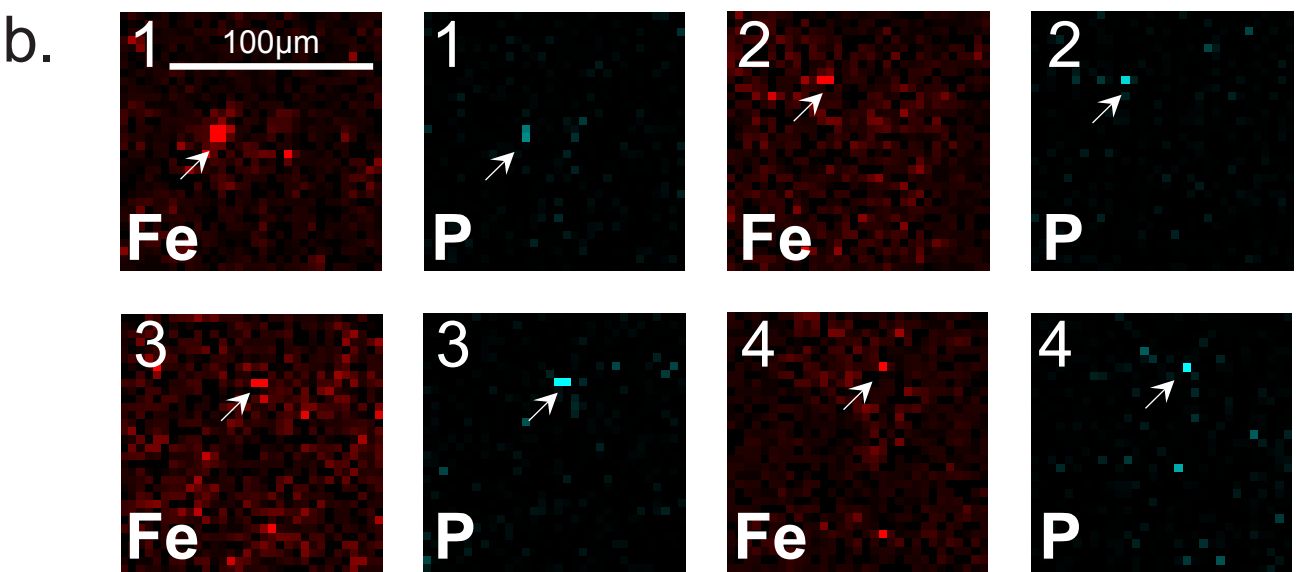
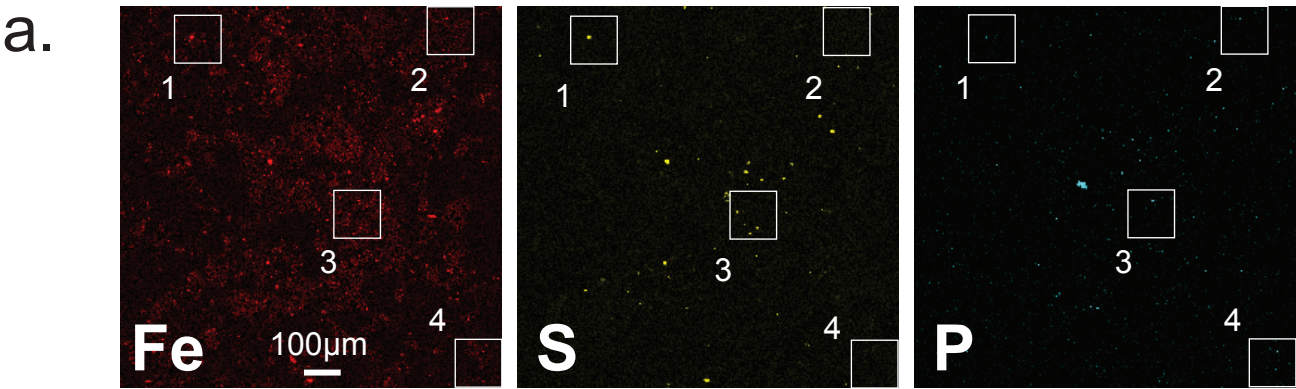


Figure 3.pdf

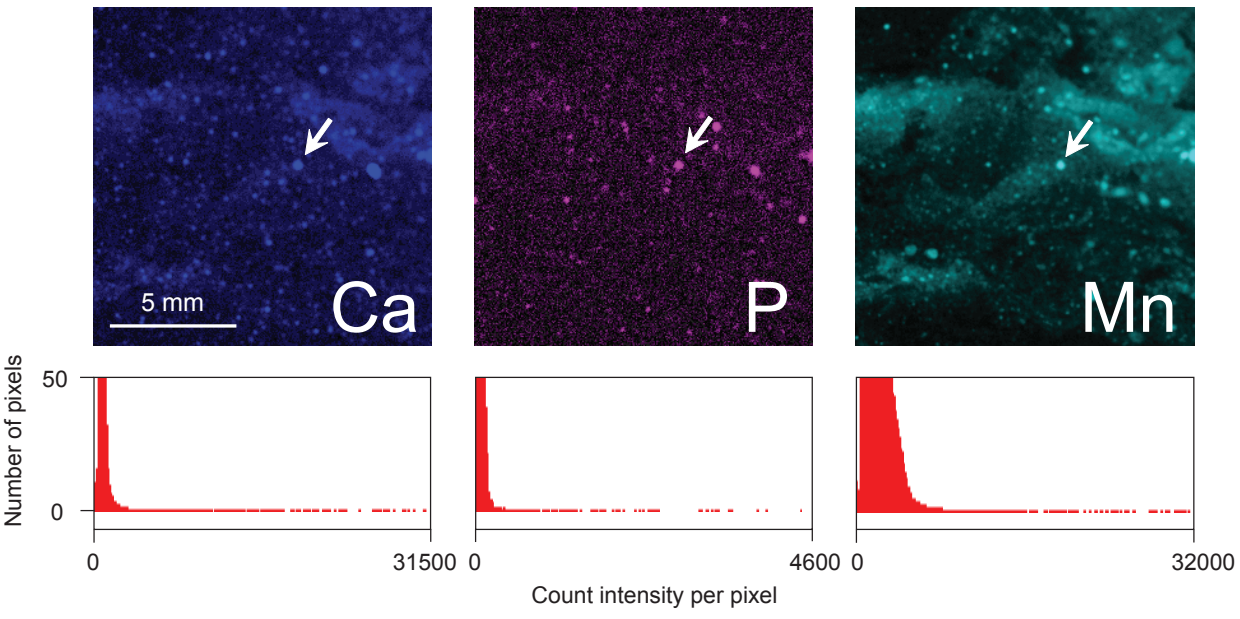
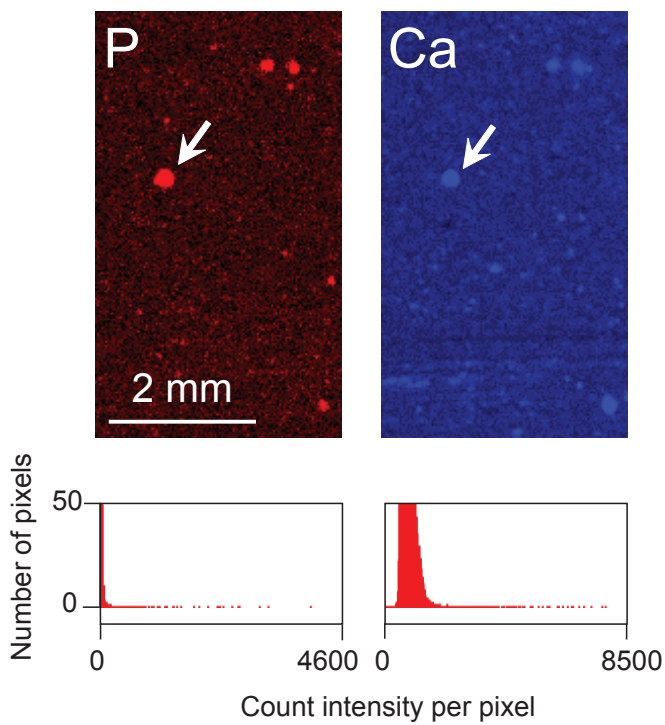


Figure 4.pdf

a.



b.

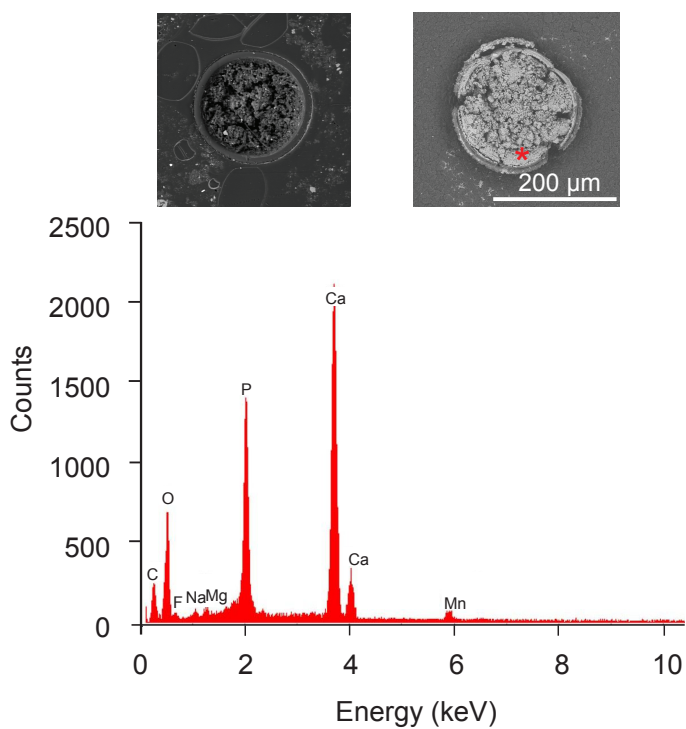
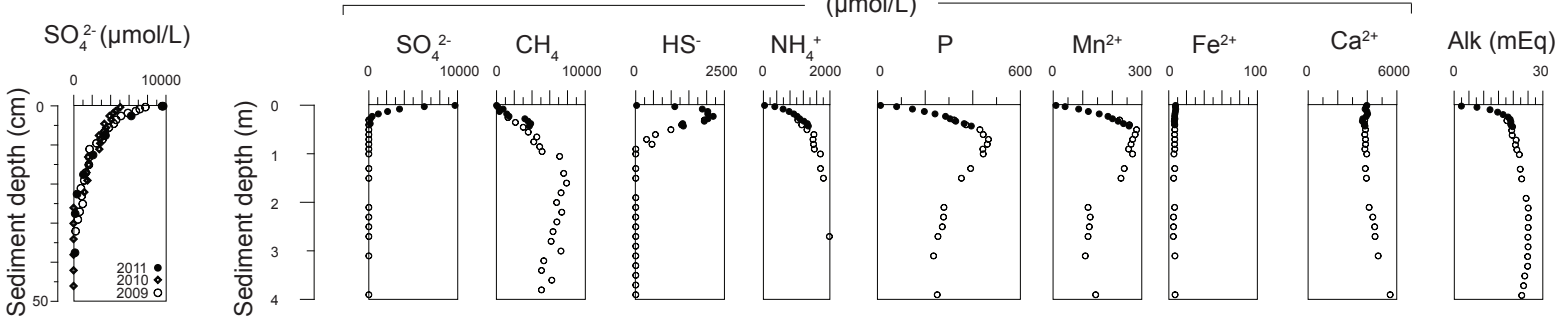


Figure 5.pdf

a. F80



b. LL19

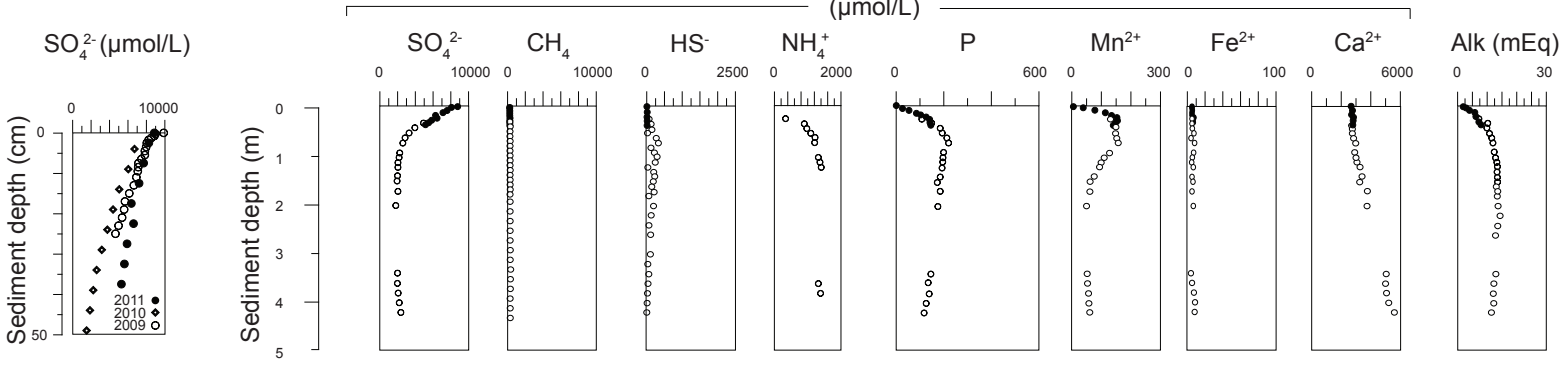


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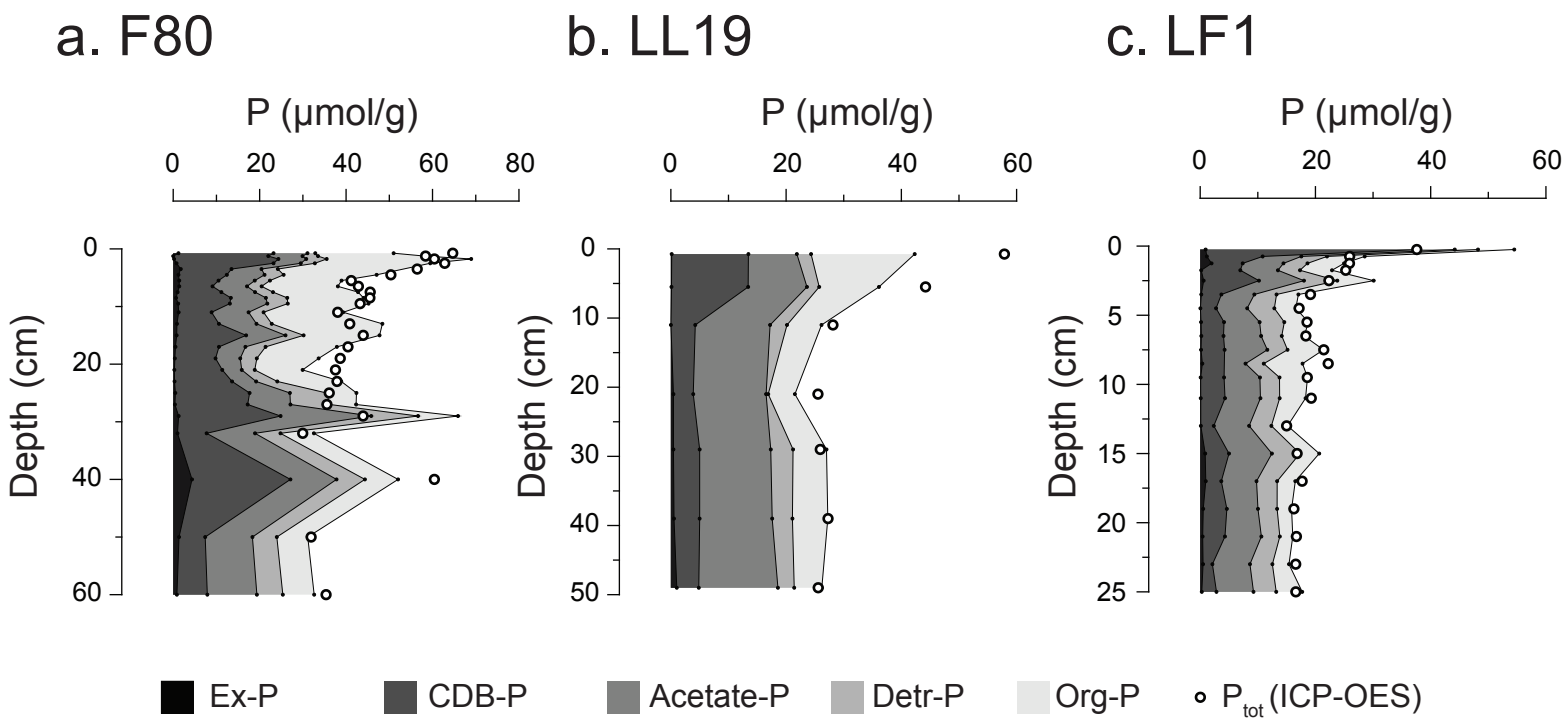


Figure 7.pdf

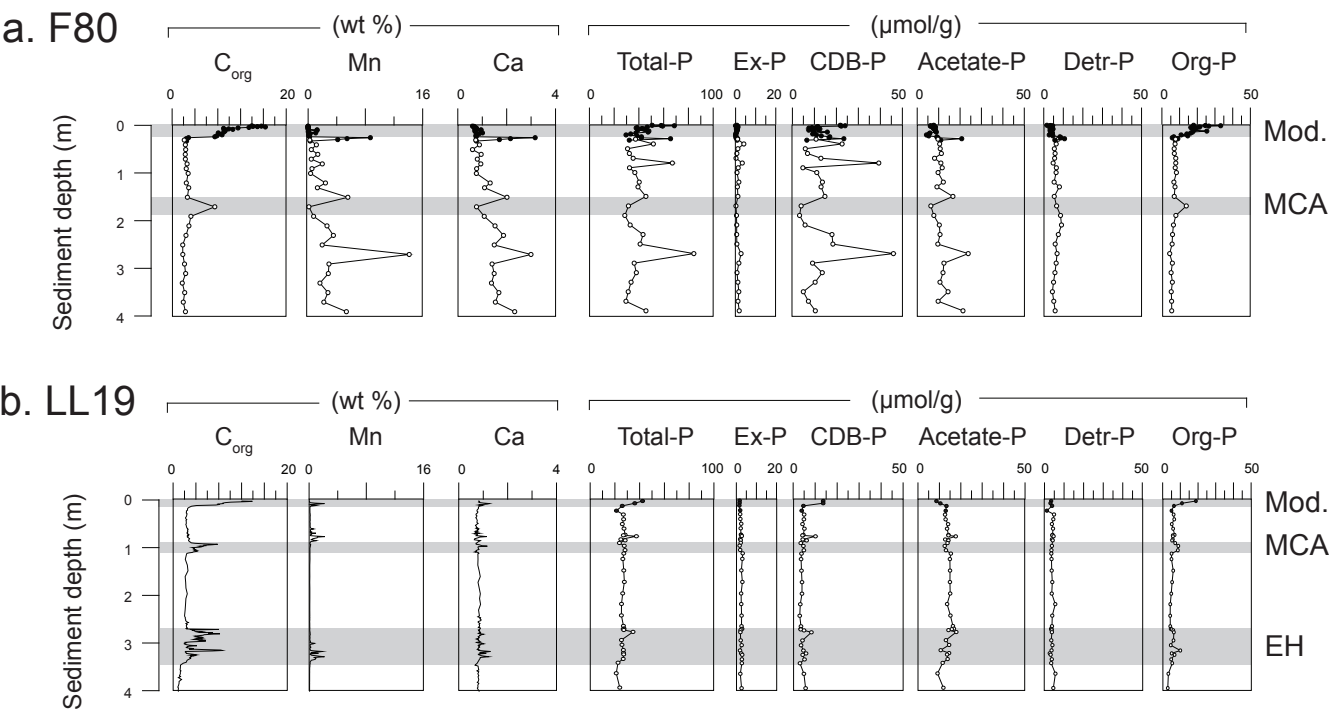
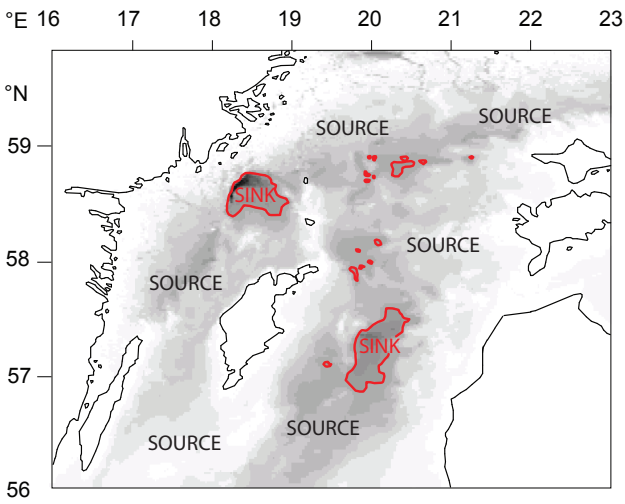
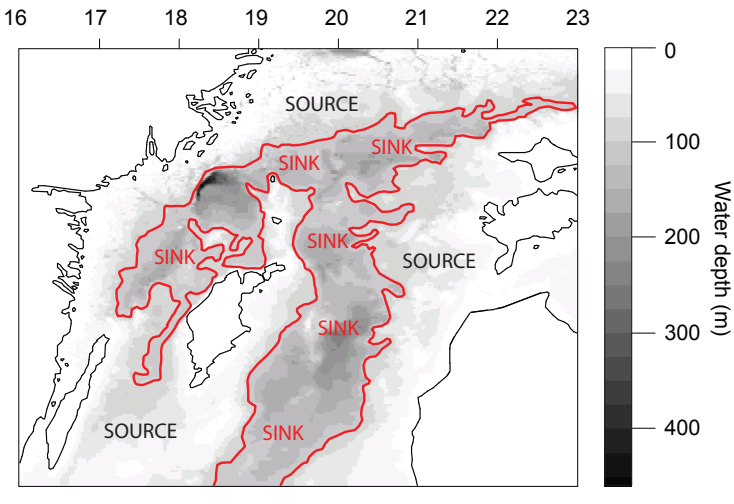


Figure 8.pdf



Oxic interval, 175 m



Anoxic interval, 100 m